

***HOTAIR* Induces the Downregulation of *miR-200* Family Members in Gastric Cancer Cell Lines**

Fatemeh Bossaghzadeh¹, Mohammadreza Hajjari^{2*},
Abdolkarim Sheikhi³, Iman Salahshourifar¹, Shiva Irani¹

¹Department of Biology, Science and research Branch, Islamic Azad University, Tehran, Iran; ²Department of Biology, Faculty of Science, Shahid Chamran University of Ahvaz, Ahvaz, Iran; ³Department of Immunology, Faculty of Medicine, Dezful University of Medical Sciences, Dezful, Iran

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ABSTRACT

Background: Gastric cancer is the fourth most common human malignancy and the second reason for cancer morbidity worldwide. LncRNA *HOTAIR* has recently emerged as a promoter of metastasis in various cancer types, including GC, through the EMT process. However, the exact mechanism of *HOTAIR* in promoting EMT is unknown. Aberrant expression of the *miR-200* family has been linked to the occurrence and development of various types of malignant tumors. This study investigates the correlation between the *HOTAIR* and *miR-200* family gene expression patterns in GC cell lines. We investigated the *miR-200* and *HOTAIR* due to their common molecular features in the EMT process. **Methods:** AGS and MKN45 cell lines were transfected with si-*HOTAIR*, along with a negative control. The effect of *HOTAIR* knockdown was also analyzed on cell viability and also on the expression of *miR-200* family members, including *miR-200a*, *-200b*, and *-200c*, in cell lines using qRT-PCR. Statistical analysis was performed to find the potential correlation between the expression level of *HOTAIR* and miRs. **Results:** Our results showed significant increased *miR-200* family expression level in transfected AGS and MKN45 GC cells (fold changes > 2; $p < 0.001$). Moreover, a negative correlation was observed between *HOTAIR* and *miR-200* expression levels in GC cell lines ($p < 0.05$). **Conclusion:** Our findings showed a significant association between *miR-200* family and *HOTAIR* expression levels in GC cell lines. Taken together, the *HOTAIR-miR-200* axis seems to play a vital role in human GC, suggesting a potential therapeutic target in future GC treatment. **DOI: 10.52547/ibj.26.1.77**

Keywords: Gene expression, Long noncoding RNA, *HOTAIR*, MicroRNAs

Corresponding Author: Mohammadreza Hajjari

Department of Biology, Faculty of Science, Shahid Chamran University of Ahvaz, Ahvaz, Iran; Tel.: (+98-61) 3331045;

E-mail: Mohamad.hajjari@gmail.com or m-hajjari@scu.ac.ir

INTRODUCTION

Gastric cancer is considered as one of the most common human malignancies and cancer morbidities worldwide^[1]. Various treatment approaches such as surgery, chemotherapy and targeted therapy have continuously been developed to manage the GC patients, although the GC prognosis has still remained poor^[2-5]. Tumor metastasis is an intricate process in which cancer cells move toward secondary organs far from the original tumor site. Metastatic

cancer cells maintain their epithelial characteristics to some extents, showing mesenchymal characteristics such as invasion or distraction. This biological process is called EMT, which is characterized by the lack of E-cadherin expression, as well as N-cadherin and vimentin upregulation^[6].

EMT is a process that is regulated by a number of signaling pathways and transcriptional/post-transcriptional factors^[7], involving both miRNA and lncRNA families^[8]. Several studies have shown the involvement of epigenetic modifications, including

List of Abbreviations:

EMT, epithelial-mesenchymal transition; **GC**, gastric cancer; ***HOTAIR***, HOX transcript antisense RNA; **lncRNA**, long noncoding RNA; ***miR-200***, microRNA-200; **miRNA**, microRNA; **ncRNAs**, noncoding RNA; **si-*HOTAIR***, siRNA targeted against *HOTAIR*; **siNC**, siRNA negative control

histone modifications, DNA methylation, lncRNAs, and miRNAs in the EMT process of cancer cells^[9-12]. Therefore, the accumulation of genetic and epigenetic modifications can contribute to the etiology of GC^[13]. It has been demonstrated that the major part of the human genome (98%) is non-coding DNA, which does not code for amino acids. MiRNAs and lncRNAs are two main ncRNA families and are able to control basic cellular processes through various mechanisms^[14]. NcRNAs are classified into two major categories: structural ncRNAs and regulatory ncRNAs^[15]. LncRNAs containing more than 200 nucleotides^[15], have different roles in the regulation of gene expression^[16]. Furthermore, miRNAs are short endogenous ncRNAs of around 18–25 nucleotides length^[17,18] that can suppress target mRNAs involved in various biological pathways^[19].

Among lncRNAs, the lncRNA *HOTAIR* is a featured one recognized in 2007^[20]. This lncRNA can repress different target genes, such as miRNAs, through an epigenetic process by recruiting polycomb repressive complex 2. Its role in different cancers, particularly gastric tumors, has been proposed previously. Silencing of *HOTAIR* has been suggested to prevent GC cell migration, invasion, and metastasis and inverses the EMT process in GC cells^[5]. The involvement of *HOTAIR* in EMT process through epigenetically silencing of miRNAs, such as miR-34a, has previously been reported^[5]. Nevertheless, this EMT regulatory action of *HOTAIR* on GC cells through miRNAs has not completely been understood yet.

Among miRNAs, *miR-200* family includes featured members acting as EMT regulatory factors due to their ability to cease Wnt/ β -catenin pathway in GC cells^[21,22]. The transcripts go through dynamic regulatory epigenetic changes associated with EMT or MET phenotypes during tumor progression^[9]. It has been indicated that the *miR-200* family determines the epithelial phenotype of cancer cells by targeting the E-cadherin repressors ZEB1 and ZEB2 proteins^[22]. By this mechanism, *miR-200* can inhibit the EMT process, which is involved in carcinogenesis.

Based on the epigenetic regulatory effect of *HOTAIR* on different miRNAs and its potential role in EMT process and due to the regulatory effect of *miR-200* on EMT markers, our study was designed to find a correlation between the expression of *HOTAIR* and *miR-200* members. The current study was aimed to find the silencing effect of *HOTAIR* on *miR-200* members involved in EMT pathway. We found that *HOTAIR* knockdown increased the *miR-200* family level in GC cell lines. Our findings provide new insights into the molecular mechanisms of lncRNAs-

mediated regulation of miRNAs involved in the EMT pathway. To our knowledge, this is the first study that reveals a correlation between the *HOTAIR* and *miR-200* family in GC.

MATERIALS AND METHODS

Cell lines and culture conditions

AGS and MKN45 human GC cell lines were prepared from the Pasteur Institute of Iran (Tehran, Iran). Cells were cultured in RPMI 1640 medium (Gibco, USA), supplemented with 10% FBS (Invitrogen, USA), and 1% penicillin and streptomycin (Invitrogen). The cells were then incubated at 37 °C, 5% CO₂, and saturated humidity. The cells were sub-cultured when grown to suitable confluence. Cell growth was monitored using an inverted microscope. Cells in the logarithmic growth phase were cultured for further analysis.

Knockdown of *HOTAIR* in AGS and MKN45 cell lines

For *HOTAIR* gene silencing, AGS and MKN45 cells were transfected with 50 nM of si-*HOTAIR*; siNC was provided from Sigma (UK). The siRNAs included *HOTAIR* siRNA-SASI (Hs02_00380445) and negative control (MISSION® siRNA Universal Negative Control #1, SIGMA/SIC001). AGS and MKN45 cells were grown in 24-well plates to 50% confluence and subsequently transfected with Lipofectamine 2000 (Invitrogen) according to the manufacturer's protocol. The experiment was repeated twice. Cells were plated until they reach 70-90% confluency at the time of transfection (AGS: 60 × 10³ cells and MKN45: 110 × 10³ cells) and then harvested at 48 h after transfection. Finally, the *HOTAIR* gene knockdown was assessed by real-time PCR.

Proliferation assay

Cell viability assay was performed using MTT kit (Atocell, England), which is used to measure cellular metabolic activity as an indicator of cell viability based on the manufacturer's guidelines. The cell lines were plated on a 96-well plate and after 24 h of seeding, AGS and MKN45 cells were transfected with 50 nM of si-*HOTAIR* along with a siNC and MOCK. MTT solution was added to each well after 48 hours and incubated in humidified atmosphere with 5% CO₂ at 37 °C for 4 h. Colorimetric detection was carried out at 490 nm using an ELISA-reader (Bio-Rad, USA). Four replicate wells were set up in each group, and experiments were repeated two times.

RNA extraction and cDNA synthesis from miRNAs/total RNAs

Total RNAs were extracted from the cultured cells using RNX (CinnaGen, Iran) based on the manufacturer's guidelines. The extracted RNA was treated with DNaseI (Sigma) and then was stored at -80 °C. Using a Nanodrop spectrophotometer (Epoch, BioTek- USA), RNA purity was determined based on A260/280 nm absorbance ratio. In addition, RNA integrity was assessed using electrophoresis on a 1% agarose gel containing SafeStain (CinnaGen). Then RNA was reverse transcribed into cDNA using cDNA Synthesis Kit (Takara, Japan) following the manufacturer's guidelines. The qPCR BONmiR kit (Cat no. BN-0011.17) was purchased from BonBiotech Company (Iran). The kit was applied to synthesize cDNA from all miRNAs, and SNORD gene was used as the internal control gene. Instead of using oligo dT and random hexamer, a relatively long sequence (50-70 nucleotides) called specific RTP (Provided from BonBiotech, Iran) was employed to trap miRNA and SNORD gene. In the end, the final amplification from RNAs attached to these pieces was carried out. The mixture was exposed to 16 °C for 10 minutes to bind the miRNAs to RTPs. It was then exposed to 42 °C for 40 minutes to completely amplify the components attached to the RTP. Tests were performed in duplicates.

Real-time PCR

Real-time PCR was performed using SYBR-Green PCR Mix (Takara) in an ABI 7100 system (Applied Biosystems, USA). The total reaction volume was 20 µl containing 1 µl of cDNA, 0.5 µl of forward primer (10 µM), 0.5 µl of reverse primer (10 µM), and 10 µl of 2× SYBR Green PCR super master mix. Conditions for PCR included denaturation at 95 °C for 5 min, 40

cycles of 5 sec at 95 °C, and annealing/extension at 60 °C for 30 s. The housekeeping gene HPRT1 was used as the internal control for *HOTAIR* gene, and SNORD gene was used as the internal control for *miR-200* family. Primers were designed by GenScript online tool (<https://www.genscript.com/tools/pcr-primers-designer>) and Primer-Blast (<https://www.ncbi.nlm.nih.gov/tools/primer-blast/>) according to the cDNA sequences extracted from the Gene bank. The primer sequences are shown in the Table 1. To validate the real-time PCR data, melt curves were plotted, and the accuracy of the curves was confirmed for each analyzed gene and primer dimer fragments. Finally, the expression of genes was assessed by $2^{-\Delta Ct}$ method (Livak method; ΔCt : Ct gene of interest - Ct internal control) to measure the expression level of genes in each sample analyzed. The efficiency of the primers was determined through standard curves plotted by raw data.

Statistical analysis

Statistical analysis was performed using Graphpad Prism 8 software. Data were shown as mean ± standard deviation. Student's t-test (unpaired) and two-way ANOVA were used for data analysis. The difference was statistically significant at $p < 0.05$. Gene expression analysis for the *HOTAIR* and *miR-200* family genes was repeated two times for each sample.

RESULTS

Downregulation of *HOTAIR* expression levels in AGS and MKN45 GC by si-*HOTAIR*

The knockdown of *HOTAIR* led into the decreased number of cells. There was a statistically significant

Table 1. Primers used in this study

Target gene	Primer name	Sequence
<i>HOTAIR</i>	F	5'-GAAAGGTCCTGCTCCGCTTC-3'
	R	5'-TCCTCTCGCCGCCGCTG-3'
<i>HPRT1</i>	F	5'-GGACTTTGCTTTCCTTGCTCAG-3'
	R	5'-GTCAAGGGCATATCCTACAACA-3'
<i>miR-200a</i>	F	5'-AACGCTAACACTGTCTGGT-3'
<i>miR-200b</i>	F	5'-TCATCCGCTAATACTGC-3'
<i>miR-200c</i>	F	5'-AGACCGCTAATACTG-3'
<i>SNORD47</i>	F	5'- ATC ACT GTA AAA CCG TT-3'
<i>MiR-200</i> Family	Universal common reverse	[BN-0011.17.5]

Family *miR-200*, *SNORD47* Primers Reference: Exclusively designed by Bon Yakhteh Company, Tehran, Iran; *HOTAIR*, *HPRT1* Primers Reference: GenScript online tool; F, forward; R, reverse

reduction of si-HOTAIR-transfected cells growth compared to siNC and MOCK groups (for AGS = 0.28 and for MKN45 = 0.23; $p < 0.0001$; Fig. 1). After the treatment of AGS and MKN45 GC cell lines with si-HOTAIR, the expression level of HOTAIR was evaluated by qRT-PCR. As shown in Figure 2A, the expression analysis of HOTAIR in AGS and MKN45 transfected by si-HOTAIR showed the decreased expression level of HOTAIR in AGS and MKN45 (fold change = 0.42 and fold change = 0.57; $p = 0.0156$ and $p < 0.05$, respectively).

Altered gene expression of miR-200 family after HOTAIR downregulation

We hypothesized that HOTAIR can induce EMT through the downregulation of miR-200 family members targeting the E-cadherin repressors ZEB1 and ZEB2 proteins (Fig. 3). To examine the possible regulatory role of HOTAIR on miR-200 family expression in AGS and MKN45 GC cell lines, the expression levels of three members of this family, including miR-200a/-200b/-200c, were analyzed. Gene expression was evaluated in AGS and MKN45 transfected with si-HOTAIR compared to the negative controls. RT-qPCR analysis of miR-200 family expression levels was performed, as well. Our data revealed that the expression of miR-200 family was upregulated in both AGS and MKN45. In other words, the knockdown of HOTAIR led to the increased level of all three genes of the miR-200 family (Table 2). As shown in AGS cell line, the most differentiated expression miRNA was the miR-200a (fold change: 3; $p < 0.001$), while the least differentiated one was miR-200b gene (fold change: 2.1; $p < 0.001$), as represented in Figure 2B). Also, in MKN45 cell line, miR-200b gene showed the most change (fold change: 2.4; $p < 0.001$), while miR-200c gene reached the least change of expression (fold change: 2.1; $p < 0.001$; Fig. 2C).

Correlations between the expression levels of HOTAIR and miR-200 family

Using Pearson's correlation coefficient, we found a correlation between the expression levels of HOTAIR and miR-200 family members in GC cell lines. Statistical significant inverse correlations were observed between HOTAIR and miR-200 family expression in GC cell lines. The expression levels of HOTAIR and miR-200a gene had a significant negative correlation in AGS cell line ($p < 0.001$; $r = -0.9934$; Fig. 4A). Furthermore, the expression level of miR-200a/c and HOTAIR were negatively correlated in MKN45 cell line ($r = -0.9797$ [Fig. 4B] and $r = -0.9800$ [Fig. 4C], respectively $p < 0.05$).

DISCUSSION

GC is known as one of the most lethal malignancies. Several studies have investigated the role of various genes in the progression of this type of cancer^[6-10,23]. LncRNAs have recently been identified as novel regulators of transcriptional and epigenetic networks^[24]. These RNAs achieve their biological functions through their interactions with multiple signaling molecules, including proteins^[25] and miRNAs^[26]. Although being expressed in special types of cells and at specific developmental stage, lncRNAs are not often associated with different kinds of cancers^[27].

Since the introduction of HOTAIR by Rinn *et al.*^[20] in 2007, numerous studies have reported that the overexpression of lncRNA HOTAIR is associated with various cancer types^[28-30], suggesting the role of HOTAIR as an oncogene in a variety of human cancers. Previous studies have revealed that HOTAIR is upregulated in GC tumor tissues^[23,31-33]. Our results confirmed that HOTAIR is a likely therapeutic target in the treatment of GC. However, the function of HOTAIR in GC has remained largely unknown. HOTAIR has been indicated to be able to induce EMT through repressing target genes^[27,34]. In this study, we found the potential mechanism of action of HOTAIR in the downregulation of miR-200 family members as one of the featured markers of EMT.

MicRNAs have been proved to function as important regulatory RNA molecules in tumorigenesis processes^[35]. MiR-200 is one of the major positive regulators in the maintenance of the epithelial phenotype via the repression of ZEB1^[36]. The miR-200

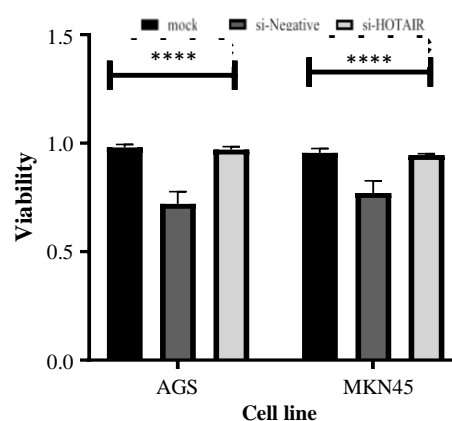


Fig. 1. Effect of knockdown on cell viability. MTT assay was performed to evaluate the cell proliferation in si-HOTAIR-transfected AGS and MKN45 cell lines. The data show a statistically significant decrease in the growth of si-HOTAIR-transfected cells compared to siNC and MOCK groups ($^{****} p < 0.0001$ vs. NC).

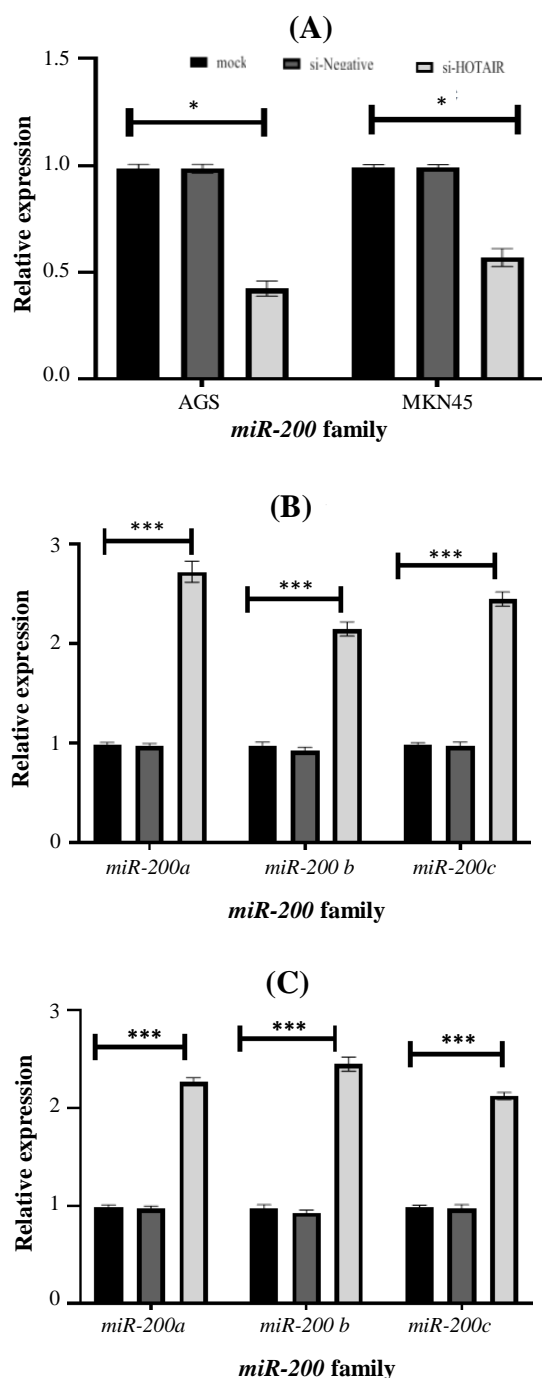


Fig. 2. RT-qPCR indicating the downregulation of *HOTAIR* expression by si-*HOTAIR*. (A) Expression levels of *HOTAIR* in AGS and MKN45 cells transfected with si-*HOTAIR*; (B) expression levels of *miR-200c/-200a/-200b* in AGS cells transfected with si-*HOTAIR*; (C) expression levels of *miR-200a/-200b/-200c* in MKN45 cells transfected with si-*HOTAIR*. Data are expressed as mean \pm SD. * $p < 0.05$ and *** $p < 0.001$.

family is involved in carcinogenesis through the regulation of EMT and is regulated through DNA methylation as a gene-silencing approach^[37]. A few

studies have indicated that *miR-200* family can be downregulated in epithelial cancers, particularly GC^[38-41]. The role of *miR-200* has previously been reported as a tumor suppressor factor for the inhibition of the EMT process and tumor growth of GC through targeting *ZEB1* and *ZEB2*^[22].

The role of *HOTAIR* in EMT regulation of GC has demonstrated in various investigations^[5,34,42]. *HOTAIR* epigenetically represses numerous factors including *miR-34a*; and contributes to the process of GC cell EMT^[5,41] by recruiting the PRC2^[5]. Previous surveys have also indicated the role of *HOTAIR*^[27,34] and *miR-200* family^[9,41] in promoting EMT program in GC. As *HOTAIR* can silence the expression of *miR-200* family, we hypothesize a correlation between *HOTAIR* and *miR-200* in GC. Our findings suggested a novel mechanism mediating shifts between EMT and MET programs through *HOTAIR* and *miR-200* members. Therefore, *miR-200* and *HOTAIR* were investigated in this study due to their common molecular features in EMT process.

Our results showed that *HOTAIR* knockdown increases the expression level of *miR-200a*, *miR-200b*, and *miR-200c* in both AGS and MKN45 GC cell lines. In this study, we explored the correlation between *HOTAIR* knockdown and *miR-200* family expression.

Our data signified that the level of *miR-200* family in GC cell lines is closely associated with the *HOTAIR* expression level. A negative correlation was also observed between *HOTAIR* and *miR-200* expression

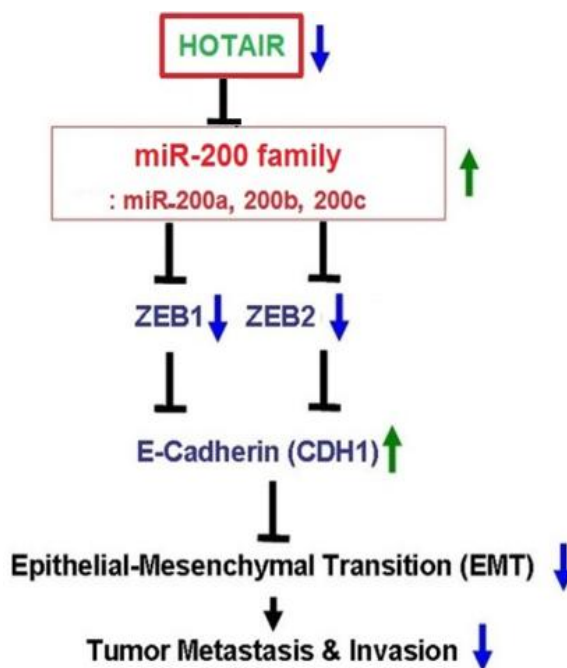
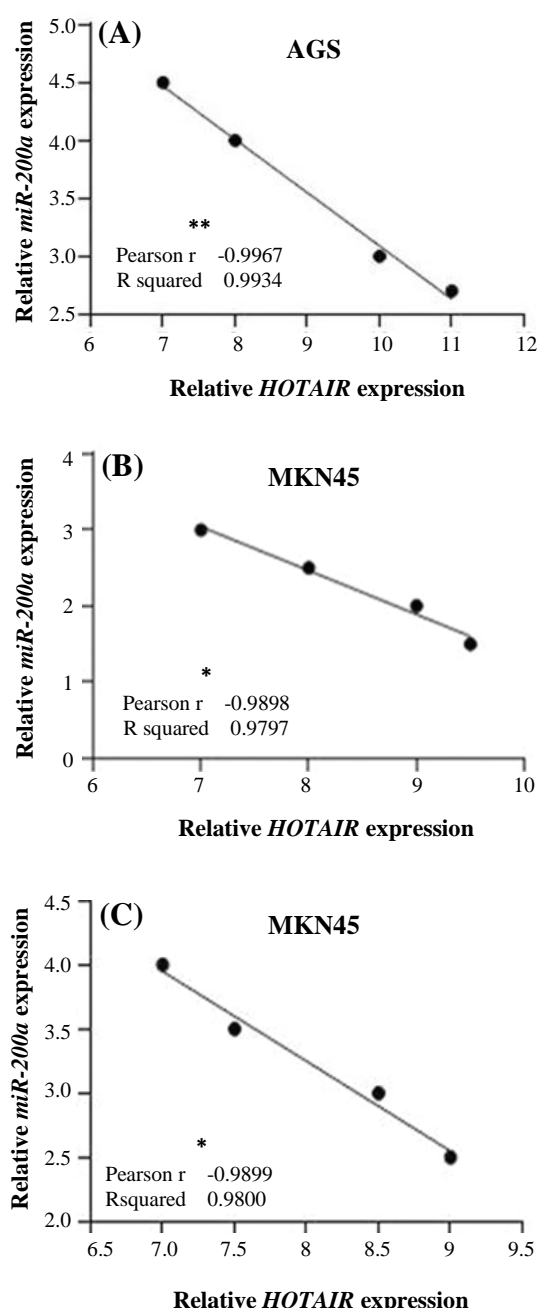


Fig. 3. Potential mechanism involving *HOTAIR* and *miR-200* family members in the EMT process.

Table 2. Fold change and *p* value of expression analysis of *miR-200* family in the loss of function study of *HOTAIR* in different GC cell lines

Gene	MKN45	AGS	<i>p</i> value
<i>miR-200a</i>	FG: 2.3	FG: 3	0.001
<i>miR-200b</i>	FG: 2.4	FG: 2.1	0.001
<i>miR-200c</i>	FG: 2.1	FG: 2.4	0.001

FG, fold change

**Fig. 4.** Correlation between the expression level of *HOTAIR* and *miR-200* in AGS and MKN45 GC cell lines. Pearson's correlation coefficient revealed the expression levels of *HOTAIR* and (A) *miR-200a* in AGS cell line (***p* < 0.01). (B and C) *miR-200a/-200c* in MKN45 cell line were negatively correlated (**p* < 0.05).

levels in GC cell lines. As *miR-200* family determines the epithelial phenotype of cancer cells by targeting the E-cadherin repressors ZEB1 and ZEB2 proteins^[9], our results displays that *HOTAIR* induces the migration and metastasis of GC cells through *miR-200* family, thereby affecting EMT. We speculate that *HOTAIR* has an effect on the EMT through this mechanism (Fig. 3). However, further studies are needed to reveal the exact regulatory mechanism of *HOTAIR* on this miR family. Overexpression of *HOTAIR* as well as interaction analyses between *HOTAIR* and regulatory factors/elements can help the researchers to find more about this axis.

Our investigation highlights the need for understanding the mechanism involving biological processes and the regulatory interactions between ncRNAs and coding transcripts, which may be beneficial to GC treatment. *HOTAIR-miR-200* axis seems to play an important role in human GC, reflecting a promising therapeutic target in future GC treatment. To our knowledge, this is the first study reporting the *HOTAIR-miR-200* axis in GC. However, the exact mechanism should be explored by more surveys in future.

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CONFLICT OF INTEREST. None declared.

REFERENCES

1. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *International journal of cancer* 2015; **136**(5): e359-e386.
2. Newton AD, Datta J, Loaiza-Bonilla A, Karakousis GC, Roses RE. Neoadjuvant therapy for gastric cancer: current evidence and future directions. *Journal of gastrointestinal oncology* 2015; **6**(5): 534.

3. Hu QJ, Ito S, Yanagihara K, Mimori k. Molecular mechanism of peritoneal dissemination in gastric cancer. *Journal of cancer metastasis and treatment* 2018; **4**: 39.
4. Shimizu D, Kanda M, Kodera Y. Emerging evidence of the molecular landscape specific for hematogenous metastasis from gastric cancer. *World journal of gastrointestinal oncology* 2018; **10**(6): 124-136.
5. Liu Y, Sun M, Xia R, Zhang EB, Liu X-h, Zhang ZH, Xu TPp, De W, Liu BRr, Wang ZX. Linc HOTAIR epigenetically silences miR34a by binding to PRC2 to promote the epithelial-to-mesenchymal transition in human gastric cancer. *Cell death and disease* 2015; **6**(7): e1802.
6. Kang Y, Massagué J. Epithelial-mesenchymal transitions: twist in development and metastasis. *Cell* 2004; **118**(3): 277-279.
7. Serrano-Gomez SJ, Maziveyi M, Alahari SK. Regulation of epithelial-mesenchymal transition through epigenetic and post-translational modifications. *Molecular cancer* 2016; **15**: 18.
8. Guo F, Kerrigan BCP, Yang D, Hu L, Shmulevich I, Sood AK, Xue F, Zhang W. Post-transcriptional regulatory network of epithelial-to-mesenchymal and mesenchymal-to-epithelial transitions. *Journal of hematology and oncology* 2014; **7**: 19.
9. Davalos V, Moutinho C, Villanueva A, Boque R, Silva P, Carneiro F, Esteller M. Dynamic epigenetic regulation of the microRNA-200 family mediates epithelial and mesenchymal transitions in human tumorigenesis. *Oncogene* 2012; **31**(16): 2062-2074.
10. Kiesslich T, Pichler M, Neureiter D. Epigenetic control of epithelial-mesenchymal-transition in human cancer. *Molecular and clinical oncology* 2013; **1**(1): 3-11.
11. Tam WL, Weinberg RA. The epigenetics of epithelial-mesenchymal plasticity in cancer. *Nature medicine* 2013; **19**(11): 1438-1449.
12. Costa FF. Epigenomics in cancer management. *Cancer management and research* 2010; **2**: 255-265.
13. Nobili S, Bruno L, Landini I, Napoli , Bechi P, Tonelli F, Rubio CA, Mini E, Nesi G. Genomic and genetic alterations influence the progression of gastric cancer. *World journal of gastroenterology* 2011; **17**(3): 290-293.
14. Xue M, Zhuo Y, Shan B. MicroRNAs, long noncoding RNAs, and their functions in human disease. *Methods in molecular biology* 2017; **1617**: 1-25.
15. Dahariya S, Paddibhatla I, Kumar S, Raghuwanshi S, Palapati A, Gutti RK. Long non-coding RNA: Classification, biogenesis and functions in blood cells. *Molecular immunology* 2019; **112**: 82-92.
16. Wang KC, Yang YW, Liu B, Sanyal A, Corces-Zimmerman R, Chen Y, Lajoie BR, Protacio A, Flynn RA, Gupta RA, Wysocka J, Lei M, Dekker J, Helms JA, Chang HY. A long noncoding RNA maintains active chromatin to coordinate homeotic gene expression. *Nature* 2011; **472**(7341): 120-124.
17. Ha M, Kim VN. Regulation of microRNA biogenesis. *Nature reviews molecular cell biology* 2014; **15**(8): 509-524.
18. Voinnet O. Origin, biogenesis, and activity of plant microRNAs. *Cell* 2009; **136**(4): 669-687.
19. Ahadi A. Dysregulation of miRNAs as a signature for diagnosis and prognosis of gastric cancer and their involvement in the mechanism underlying gastric carcinogenesis and progression. *IUBMB life* 2020; **72**(5): 884-898.
20. Rinn JL, Kertesz M, Wang JK, Squazzo SL, Xu X, Bruggmann SA, Goodnough LH, Helms JA, Farnham PJ, Segal E, Chang HY. Functional demarcation of active and silent chromatin domains in human HOX loci by noncoding RNAs. *cell* 2007; **129**(7): 1311-1323.
21. Su J, Zhang A, Shi Z, Ma F, Pu P, Wang T, Zhang J, Kang Ch, Zhang Q. MicroRNA-200a suppresses the Wnt/ β -catenin signaling pathway by interacting with β -catenin. *International journal of oncology* 2012; **40**(4): 1162-1170.
22. Cong N, Du P, Zhang A, Shen F, Su J, Pu P, Wang T, Zjang J, Kang Ch, Zhang Q. Downregulated microRNA-200a promotes EMT and tumor growth through the wnt/ β -catenin pathway by targeting the E-cadherin repressors ZEB1/ZEB2 in gastric adenocarcinoma. *Oncology reports* 2013; **29**(4): 1579-1587.
23. Hajjari M, Behmanesh M, Sadeghizadeh M, Zeinoddini M. Up-regulation of HOTAIR long non-coding RNA in human gastric adenocarcinoma tissues. *Medical oncology* 2013; **30**(3): 670.
24. Sui C-j, Zhou Y-m, Shen W-f, Dai BH, Lu JJ, Zhang MF, Yang JM. Long noncoding RNA GIHCG promotes hepatocellular carcinoma progression through epigenetically regulating miR-200b/a/429. *Journal of molecular medicine* 2016; **94**(11): 1281-1296.
25. Ferre F, Colantoni A, Helmer-Citterich M. Revealing protein-lncRNA interaction. *Briefings in bioinformatics* 2016; **17**(1): 106-116.
26. Paraskevopoulou MD, Hatzigeorgiou AG. Analyzing miRNA-lncRNA interactions. *Methods in molecular biology* 2016; **1402**: 271-286.
27. Lee NK, Lee JH, Park CH, Yu D, Lee Y, Cheong J-H, Noh SH, Lee SK. Long non-coding RNA HOTAIR promotes carcinogenesis and invasion of gastric adenocarcinoma. *Biochemical and biophysical research communications* 2014; **451**(2): 171-178.
28. Battistelli C, Cicchini C, Santangelo L, Tramontano A, Grassi L, Gonzalez FJ, Nonno V, Grassi G, Amicone L, Tripodi M. The Snail repressor recruits EZH2 to specific genomic sites through the enrollment of the lncRNA HOTAIR in epithelial-to-mesenchymal transition. *Oncogene* 2017; **36**(7): 942-955.
29. Zhang JX, Han L, Bao Z-S, Wang Y-Y, Chen LY, Yan W, Liu N, YU SZ, Pu PY, You YP, Jiang T, Kang CS, Cooperative Group ChG. HOTAIR, a cell cycle-associated long noncoding RNA and a strong predictor of survival, is preferentially expressed in classical and mesenchymal glioma. *Neuro-oncology* 2013; **15**(12): 1595-1603.
30. Hajjari M, Salavaty A. HOTAIR: an oncogenic long non-coding RNA in different cancers. *Cancer biology and medicine* 2015; **12**(1): 1-9.

31. Pan W, Liu L, Wei J, Ge Y, Zhang J, Chen H, Zhou L, Yuan Q, Zhou Ch, Yang M. A functional lncRNA HOTAIR genetic variant contributes to gastric cancer susceptibility. *Molecular carcinogenesis* 2016; **55**(1): 90-96.
32. Zhao W, Dong S, Duan B, Chen P, Shi L, Gao H, Qi H. HOTAIR is a predictive and prognostic biomarker for patients with advanced gastric adenocarcinoma receiving fluorouracil and platinum combination chemotherapy. *American journal of translational research* 2015; **7**(7): 1295-1302.
33. Bhan A, Mandal SS. LncRNA HOTAIR: A master regulator of chromatin dynamics and cancer. *Biochimica et biophysica acta* 2015; **1856**(1): 151-164.
34. Song Y, Wang R, Li L-W, Wang Y-F, Wang Q-X, Zhang Q. Long non-coding RNA HOTAIR mediates the switching of histone H3 lysine 27 acetylation to methylation to promote epithelial-to-mesenchymal transition in gastric cancer. *International journal of oncology* 2019; **54**(1): 77-86.
35. Corté H, Manceau G, Blons H, Laurent-Puig P. MicroRNA and colorectal cancer. *Digestive and liver disease* 2012; **44**(3): 195-200.
36. Burk U, Schubert J, Wellner U, Schmalhofer O, Vincan E, Spaderna S, Brabletz Th. A reciprocal repression between ZEB1 and members of the miR-200 family promotes EMT and invasion in cancer cells. *EMBO reports* 2008; **9**(6): 582-589.
37. Neves R, Scheel C, Weinhold S, Honisch E, Iwaniuk KM, Trompeter H-I, Niederacher D, Wernet P, Uhrberg M. Role of DNA methylation in miR-200c/141 cluster silencing in invasive breast cancer cells. *BMC research notes* 2010; **3**: 219.
38. Men D, Liang Y, Chen L. Decreased expression of microRNA-200b is an independent unfavorable prognostic factor for glioma patients. *Cancer epidemiology* 2014; **38**(2): 152-156.
39. Paterson EL, Kazenwadel J, Bert AG, Khew-Goodall Y, Ruszkiewicz A, Goodall GJ. Down-regulation of the miRNA-200 family at the invasive front of colorectal cancers with degraded basement membrane indicates EMT is involved in cancer progression. *Neoplasia* 2013; **15**(2): 180-IN22.
40. Soubani O, Ali AS, Logna F, Ali Sh, Philip PA, Sarkar FH. Re-expression of miR-200 by novel approaches regulates the expression of PTEN and MT1-MMP in pancreatic cancer. *Carcinogenesis* 2012; **33**(8): 1563-1571.
41. Ning X, Shi Z, Liu X, Zhang A, Han L, Jiang Km, Kang Ch, Zhang Q. DNMT1 and EZH2 mediated methylation silences the microRNA-200b/a/429 gene and promotes tumor progression. *Cancer letters* 2015; **359**(2): 198-205.
42. Xu ZY, Yu QM, Du YA, Yang LT, Dong RZ, Huang L, Yu PF, Cheng XD. Knockdown of long non-coding RNA HOTAIR suppresses tumor invasion and reverses epithelial-mesenchymal transition in gastric cancer. *International journal of biological sciences* 2013; **9**(6): 587-597.