

Carvacrol and Thymol Attenuate Cytotoxicity Induced by Amyloid β_{25-35} via Activating Protein Kinase C and Inhibiting Oxidative Stress in PC12 Cells

Zahra Azizi¹, Mona Salimi¹, Amir Amanzadeh², Nahid Majelssi¹ and Nasser Naghdi^{1*}

¹Department of Physiology and Pharmacology, Pasteur Institute of Iran, Tehran, Iran;

²Department of Cell Bank, Pasteur Institute of Iran, Tehran, Iran

Received 14 April 2019; accepted 12 June 2019; published online 22 January 2020

ABSTRACT

Background: Our previous findings indicated that carvacrol and thymol alleviate cognitive impairments caused by $A\beta$ in rodent models of AD. In this study, the neuroprotective effects of carvacrol and thymol against $A\beta_{25-35}$ -induced cytotoxicity were evaluated, and the potential mechanisms were determined. **Methods:** PC12 cells were pretreated with $A\beta_{25-35}$ for 2 h, followed by incubation with carvacrol or thymol for additional 48 h. Cell viability was measured by the MTT method. A flurospectrophotometer was employed to observe the intracellular ROS production. PKC activity was analyzed using ELISA. **Results:** Our results indicated that carvacrol and thymol could significantly protect PC12 cells against $A\beta_{25-35}$ -induced cytotoxicity. Furthermore, $A\beta_{25-35}$ could induce intracellular ROS production, while carvacrol and thymol could reverse this effect. Moreover, our findings showed that carvacrol and thymol elevate PKC activity similar to Bryostatin-1, as a PKC activator. **Conclusion:** This study provided the evidence regarding the protective effects of carvacrol and thymol against $A\beta_{25-35}$ -induced cytotoxicity in PC12 cells. The results suggested that the neuroprotective effects of these compounds against $A\beta_{25-35}$ might be through attenuating oxidative damage and increasing the activity of PKC as a memory-related protein. Thus, carvacrol and thymol were found to have therapeutic potential in preventing or modulating AD. **DOI: 10.29252/ibj.24.4.243**

Keywords: Alzheimer's disease, Carvacrol, Thymol, Reactive oxygen species, Protein kinase C

Corresponding Author: Nasser Naghdi

Department of Physiology and Pharmacology, Pasteur Institute of Iran, Tehran 1316943551, Iran; E-mail: naghdi@pasteur.ac.ir

INTRODUCTION

Alzheimer's disease is a neurodegenerative disorder characterized by progressive deterioration of brain structure and function^[1]. As life expectancy is increasing in parallel with the economic development, the risk of AD along with its cost and social burden will be felt even more in the future^[2].

The excessive accumulation of senile plaque in the brain is one of the neuropathological hallmarks of AD^[3]. The $A\beta$ peptide in the plaque cores is a 39-43 amino acid sequence generated from a larger

transmembrane protein, the APP^[4,5]. APP can be processed by two alternative pathways: amyloidogenic and non-amyloidogenic. In the amyloidogenic pathway, β - and γ -secretase cooperatively convert APP into $A\beta$ to form the deposits. In the non-amyloidogenic pathway, α -secretase reroutes the substrate APP toward the production of soluble APP α , which is non-toxic and prevents the formation of amyloid plaques^[5]. PKC has been shown to activate α -secretase, increase the non-toxic soluble APP α release and reduce the secretion of the toxic $A\beta$ peptide^[6]. In this sense, PKC pharmacology would be an interesting area of research for development of cognition-enhancing agents and

List of Abbreviations:

AD, Alzheimer's disease; **APP**, amyloid precursor protein; **$A\beta$** , amyloid β ; **DCF-DA**, 2',7'-dichlorofluoresceine diacetate; **MTT**, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; **PKC**, protein kinase C; **ROS**, reactive oxygen species

therapeutics against memory loss^[7]. On the other hand, oxidative stress has been revealed to contribute to the progression of AD. Some studies have shown that A β plays a crucial role in the accumulation of ROS^[8,9]. High ROS levels cause oxidative stress that may result in cell death, ultimately leading to neurodegenerative diseases including AD^[9,10]. Therefore, therapeutic effort for attenuating the oxidative stress could be beneficial in AD treatment^[11].

Despite of extensive research on AD for more than a century, few effective treatment options have been developed^[12]. Current available drugs used for the treatment of AD transiently alleviate some of the symptoms, but do not modify the disease mechanism or cure it. Therefore, there is an urgent need for the development of new drugs^[13,14]. Nowadays, several medicinal plants and their constituents have been suggested as possible treatments for AD based on their effects on oxidant formation^[15] and processing of APP^[16]. Among the chemical compounds derived from plants, monoterpenes^[2] along with phenolic compounds^[17,18] exhibit diverse pharmacological activities, representing them as potential modulators of AD. In this regard, carvacrol (2-methyl-5-(1-methylethyl) phenol) and thymol (2-isopropyl-5-methylphenol) as two phenolic monoterpenes are abundantly found in aromatic plants, including *Thymus vulgaris* and *Zataria multiflora*^[19]. These cyclic monoterpenes exert protective activities in a variety of pathological states including oxidative stress^[20,21], which might be beneficial in reducing AD symptoms.

Previously, we investigated the effects of carvacrol and thymol in two animal models of AD and observed their effectiveness in alleviating cognitive impairments caused by A β ^[22]. In this study, carvacrol and thymol were used in an *in vitro* model of AD, A β -treated PC12 cells, and their antioxidant activity as well as their effects on PKC activity, as a memory-related enzyme, were studied.

MATERIALS AND METHODS

Chemicals and agents preparation

A β ₂₅₋₃₅ peptide, MTT, DCF-DA, DMSO, thymol and carvacrol were all provided by Sigma-Aldrich Chemical Company (St. Louis, Missouri, USA). RPMI 1640 medium, fetal bovine serum, and horse serum were obtained from Gibco BRL (Grand Island, NY, USA). PKC kinase activity kit and Bryostatin-1 were purchased from Enzo Life Sciences Company (Farmingdale, NY, USA) and Tocris Bioscience (Bristol, UK), respectively. The PC12 cell line (rat pheochromocytoma cells) was provided by the

National Cell Bank, Pasteur Institute of Iran (Tehran). A β ₂₅₋₃₅ peptide was dissolved in RPMI at a concentration of 1 mM and incubated at 37 °C for 4 days to obtain the aggregated form. The solution was stored at -20 °C. The stock solution was diluted to desired concentrations immediately before use. Bryostatin-1 was dissolved in 1% DMSO at the concentration of 10 nM and stored tightly sealed at -20 °C. Carvacrol and thymol were freshly dissolved in 1% DMSO as a stock solution and then diluted with RPMI to desired various concentrations by the serial dilution of stock solution before the experiment.

Cell culture and treatment

PC12 cells were cultured in flasks with RPMI 1640 medium supplemented with 15% fetal bovine serum and 1% antibiotics (penicillin-streptomycin-nystatin) and incubated in a 5% CO₂ atmosphere at 37 °C. Next, 75% of culture medium was replaced with a fresh medium every 48 h. PC12 cells were passaged when the culture was 80% confluent. The cells were subcultured once a week in the ratio of 1:4. Cell culture solutions were prepared under sterile conditions. The cells in exponential growth were seeded on 96-well plates at a density of 2×10^4 cells per well, in 100 μ l of growth medium and allowed to attach for 24 h. The cells were then exposed to the desired concentrations of A β , carvacrol, thymol, or Bryostatin-1 according to each experiment.

Cell viability assay

The MTT assay was used for three different experiments. First, to achieve the appropriate concentration of A β , the cells were treated with different concentrations of A β (30, 50, and 100 μ M) for 48 h. Second, to detect the safety of carvacrol and thymol, cells were exposed to different concentrations of carvacrol and thymol ranging from 1 μ M to 1000 μ M for 48 h. The third stage was an assay in which the cells were treated with A β ₂₅₋₃₅ (50 μ M), followed by indicated concentrations of carvacrol (10, 20, and 50 μ M), thymol (10, 20, and 50 μ M), or Bryostatin-1 (10 nM). The control group was treated with DMSO 1% (v/v). After each of these three experiments and at the end of incubation, cell viability was evaluated by MTT assay^[23]. In Brief, 10 μ l of MTT solution, diluted in sterile PBS (5 mg/ml), was added to the culture plates. The plates were incubated at 37 °C for 4 h. Then the media was carefully removed, and 100 μ l of DMSO as lysis buffer was added to each well. After 60 min, the amount of MTT formazan product was determined by measuring absorbance using a 96-well ELISA microplate reader at a wavelength of 545 nm and a reference wavelength of 630 nm. The results were

expressed as the percentage of MTT reduction, assuming that the absorbance of control was 100%. All experiments were performed three independent times, each in triplicate.

Measurement of ROS generation

The intracellular ROS formation levels were monitored using the fluorescent probe DCF-DA, which readily diffuses through the cell membrane^[24]. Within the cells, the nonfluorescent dye reacts with the intracellular ROS and is converted into DCF, which is a green fluorescent dye. PC12 cells were treated with 50 μ M of A β ₂₅₋₃₅, followed by the various concentrations of carvacrol (10, 20, and 50 μ M), thymol (10, 20, 50 μ M), or 10 nM of Bryostatin-1 for 48 h. At the end of the treatment, the plates were washed twice with PBS. Then the cells were incubated in the presence of DCF-DA at a final concentration of 10 μ M at 37 °C for 50 min. After incubation, DCF intensity was determined with a fluoremeter (BioTeK, Swindon, UK). The excitation and emission wavelengths were 485 nm and 528 nm, respectively. The results were expressed in relation to ROS-associated fluorescence intensity of control cells, which was set to 100%. The control group was treated with DMSO 1% (v/v). All experiments were repeated three independent times, each in triplicate.

Sample preparation of cell lysates

PC12 cells with different treatments were cultured in a dish (100 mm) to reach 80-90% confluence. After the removal of the culture medium, cells were washed twice with cold PBS and lysed with 300 μ l/dish of cool RIPA buffer solution containing 5 M of sodium chloride, 1% Triton X-100, 0.5% sodiumdeoxycholate, 1M of Tris-HCl (pH 7.4), 0.1% SDS, supplemented with protease inhibitor. After 15-min incubation period, the plates were rotated by hand to cover cells with a film of lysis buffer. The cells were then immediately dislodged with a cell scraper, and the cell lysate were collected in a pre-chilled 1.5-ml microcentrifuge tube. In the next step, the microtubes with the cell suspensions were taped vigorously five times and leaved on ice for 15 min. Afterwards, the samples were centrifuged at 16,000 \times g at 4 °C for 15 min to separate the supernatant. At the end, the clear supernatant fractions were transferred to new pre-chilled 1.5-ml microtubes and stored at -70 °C. The protein concentration of each sample was determined using the Bradford method, and equal amounts of proteins were analyzed for PKC activity assay.

Kinase activity assay

The non-radioactive PKC kinase activity assay kit

(ENZO, ADI-EKS-420A) was based on a solid-phase ELISA that utilizes a polyclonal antibody recognizing the phosphorylated form of the substrate. The assay was designed for the analysis of PKC activity in the solution phase. In brief, the wells of the PKC substrate microtiter plate were filled with 50 μ l of kinase assay dilution buffer at room temperature for 10 minutes, and then the liquid was carefully aspirated from the wells. The samples were added to the appropriate wells of the PKC substrate microtiter plate, and the reaction was initiated by adding 10 μ l of diluted ATP to each well. After 90 minutes, the reaction was stopped by discarding the contents of each well. Then 40 μ l of phosphospecific substrate antibody was added to each well, and the plate was incubated at room temperature for 60 minutes. After that, the wells were washed four times with 100 μ l of 1 \times wash buffer, and 40 μ l of diluted anti-rabbit IgG, horseradish peroxidase conjugate, was added to each well. After 30 minutes, the wells were washed four times with 100 μ l of 1 \times wash buffer. Then 60 μ L of 3,3',5,5'-tetramethylbenzidine substrate was added to each well, and the plate was incubated at room temperature for 30 minutes. Finally, 20 μ l of acid stop solution was added to each well. The absorbance was measured at 450 nm using a Bio-Rad microtiter plate reader (USA).

Statistical analysis

All data were presented as means \pm SEM. Statistical analyses were carried out using one-way ANOVA, followed by Tukey's honestly significant difference. $p < 0.05$ was considered to be statistically significant compared with A β ₂₅₋₃₅ treatment or control group.

RESULTS

Determination of A β ₂₅₋₃₅ cytotoxicity

First, to assess the A β cytotoxicity, PC12 cells were treated with different concentrations (30, 50, and 100 μ M) of A β ₂₅₋₃₅. As shown in Figure 1, cell viability measured by MTT assay significantly reduced after A β ₂₅₋₃₅ treatment at different concentrations of A β ₂₅₋₃₅ for 48 h. Based on our results, 50 μ M of A β ₂₅₋₃₅ showed the maximum cytotoxicity, and this concentration was selected for our further experiments.

Safety of carvacrol and thymol

The safe doses of carvacrol and thymol were first determined by measuring the cell viability assay. Carvacrol and thymol at concentrations ranging between 1 and 1000 μ M were added separately to the wells containing PC12 cells for 48 h. The results showed statistically significance difference between

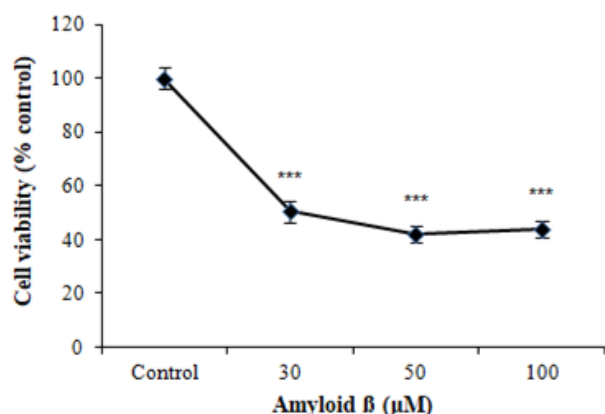


Fig. 1. A β ₂₅₋₃₅-induced cell death in PC12 cells. The cells were incubated with different concentrations of A β ₂₅₋₃₅ (30, 50, and 100 μ M) for 48 h, and the cell viability was measured by MTT assay. The data were presented as the mean \pm SEM, n = 9. *** p < 0.001 vs. control.

the control group (1% DMSO), and the groups were treated with carvacrol at concentrations up to 200 μ M; however, treatment with high doses of carvacrol (500 and 1000 μ M) resulted in PC12 cytotoxicity. Furthermore, exposure to concentrations up to 100 μ M of thymol did not decrease the cell viability compared with the control group (Fig. 2). Our findings indicated that 1-200 μ M concentrations of carvacrol along with 1-100 μ M concentrations of thymol revealed no cytotoxicity against PC12 cells and those used for our further experiments.

Carvacrol and thymol protection against A β -induced cytotoxicity

In order to explore the protective effects of carvacrol and thymol against A β ₂₅₋₃₅-induced cytotoxicity, PC12 cells were incubated with 50 μ M of A β ₂₅₋₃₅, and 30 minutes later, carvacrol or thymol at concentrations of 10, 20, and 50 μ M were added to the cells for 48 h. The cytotoxicity induced by A β ₂₅₋₃₅ was reversed following treatment with carvacrol at concentrations of 10, 20, and 50 μ M by 70%, 80%, and 85%, respectively (Fig. 3). Similarly, thymol could reverse A β toxicity at concentrations of 10, 20, and 50 μ M on PC12 cells. Our findings imply that treatment with carvacrol or thymol at concentrations of 10, 20 and 50 μ M could reverse the cytotoxicity induced by A β ₂₅₋₃₅ on PC12 cells. This effect was similar to Bryostatin-1 that showed significant protection against A β ₂₅₋₃₅-induced cytotoxicity (Fig. 3).

Carvacrol and thymol prevention against A β -induced ROS production

Intracellular accumulation of ROS was determined by DCF-DA assay as described in the material and

methods. A substantial increase in the level of ROS was observed (Fig. 4) in PC12 cells exposed to 50 μ M of A β ₂₅₋₃₅ for 48 h compared to the control group (DMSO 1%). Also, 10 nM of Bryostatin-1 significantly reduced intracellular ROS levels compared to A β group. Carvacrol and thymol at concentrations of 10, 20, and 50 μ M showed a similar effect as Bryostatin-1 (Fig. 4). Altogether, the results demonstrated that carvacrol and thymol had similar behavior toward Bryostatin-1 in the reduction of ROS level.

Induction of PKC activity by carvacrol and thymol in PC12 cells-treated with A β

Enzyme activity was determined using an assay kit obtained from Enzo Life Science (USA). We found that exposure of PC12 cells to 50 μ M of A β ₂₅₋₃₅ for 48 h considerably inhibited PKC activity compared to the control group. As expected, Bryostatin-1 significantly inversed this effect. Treatment with carvacrol or thymol at concentrations of 10, 20, and 50 μ M could also significantly induce the PKC activity (Fig. 5).

DISCUSSION

In our previous work, we demonstrated the potential neuroprotective effects of carvacrol and thymol in AD animal models^[22]. Our present results indicated that, applying carvacrol and thymol attenuated cell death induced by A β ₂₅₋₃₅, and prevented A β ₂₅₋₃₅-induced oxidative stress in PC12 cells. In addition, A β ₂₅₋₃₅ inhibited PKC activation, whereas carvacrol and thymol could reverse this effect similar to Bryostatin-1, as a potent PKC activator.

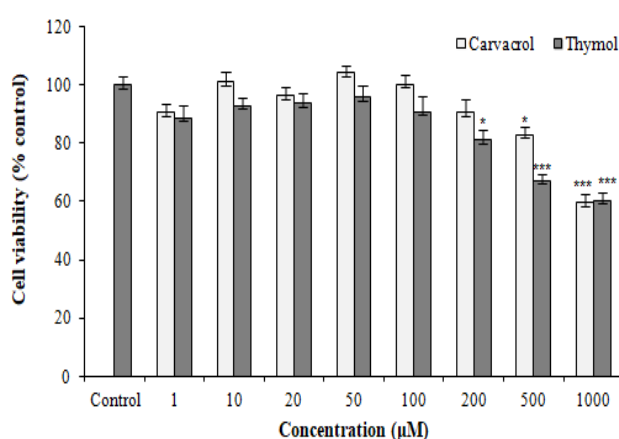


Fig. 2. Concentration-dependent effects of carvacrol and thymol on the cell viability of PC12 cells. The cells were incubated with different concentrations of carvacrol or thymol (1-1000 μ M) for 48 h, and the cell viability was measured by MTT assay. The viability of the control group (DMSO 1%) was set to 100%. The data were presented as the mean \pm SEM, n = 9. * p < 0.05 and *** p < 0.001 vs. control.

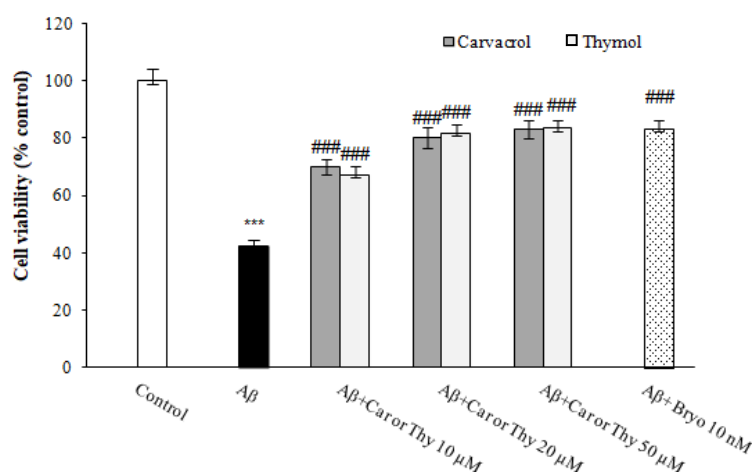


Fig. 3. The protective effects of carvacrol (Car) and thymol (Thy) on A β -induced cytotoxicity in PC12 cells. The cells were incubated with different concentrations (10, 20, and 50 μ M) of carvacrol and thymol and also 10 nM of Bryostatin (Bryo)-1 in the presence of 50 μ M of A β ₂₅₋₃₅ for 48 h. The viability of the control group (DMSO 1%) was set to 100%. The data were presented as the mean \pm SEM. *** p < 0.001 vs. control, ### p < 0.001 vs. A β

A β deposition and subsequent formation of senile plaques in the brain have well been recognized as key events during neuropathogenesis of AD. Therefore, studies on the discovery of drugs inhibiting A β formation and toxicity are of great significance for AD treatment^[25]. Studies have suggested that the neurobiological effects of phytochemicals derived from medicinal plants may contribute to clinical benefits in AD^[14,26]. Carvacrol and thymol, natural monoterpenoid phenols, are obtained from many plants belonging to Lamiaceae family and possess diverse pharmacologic properties including antibacterial^[20], cardioprotective^[21], and anticancer^[27] effects. Neuroprotective effects of these compounds have been reported in a variety of animal models, including

AD^[22,28,29]. Herein, the effects of carvacrol and thymol on A β -induced cytotoxicity were evaluated in PC12 cells. The rats adrenal pheochromocytoma cells line, namely PC12, appears to be an *in vitro* cell model for neurobiology and neurochemistry studies^[30]. PC12 cells were found to respond specifically to the 25-35 fragment of A β as a neurodegenerative factor in AD, stimulating the production of cellular ROS and A β -mediated cell death^[31,32]. The results of MTT assay revealed that treatment with carvacrol or thymol significantly increased the cell viability compared to that of the A β ₂₅₋₃₅ group.

Intracellular oxidative stress resulting from imbalance in ROS production and cellular antioxidant defense mechanisms mediates the damage

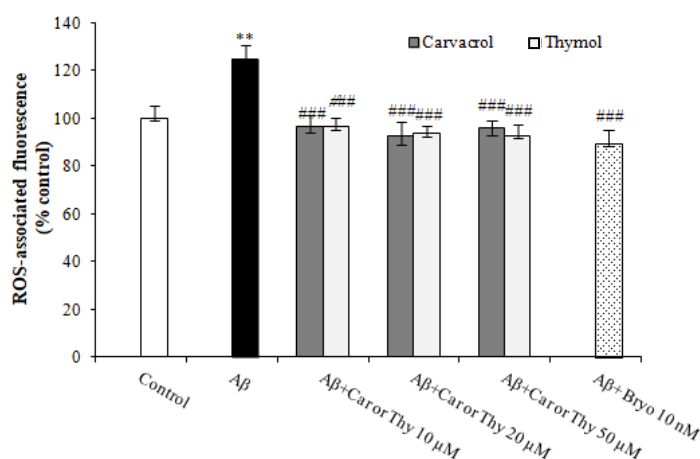


Fig. 4. Effects of carvacrol (Car) and thymol (Thy) on the intracellular accumulation of ROS produced by A β ₂₅₋₃₅ in PC12 cells. The cells were incubated with different concentrations (10, 20, and 50 μ M) of carvacrol and thymol or 10 nM of Bryostatin (Bryo)-1 in the presence of 50 μ M of A β ₂₅₋₃₅ for 48 h. Intracellular ROS was measured based on the peroxide-sensitive DCF. ROS-associated fluorescence of the control group (DMSO 1%) was set to 100%. The data were presented as the mean \pm SEM. ** p < 0.01 vs. control, ### p < 0.001 vs. A β group.

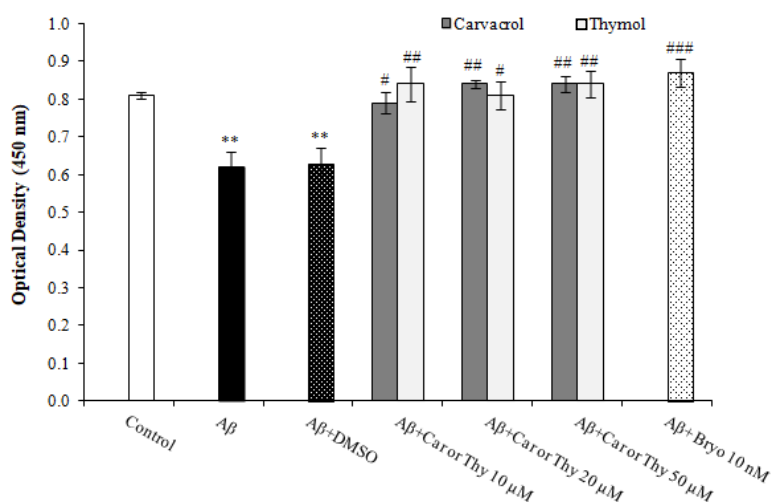


Fig. 5. Carvacrol (Car) and thymol (Thy) effect on PKC pathway inhibited by A β ₂₅₋₃₅. The cells were incubated with 10, 20, and 50 μ M of carvacrol and thymol or 10 nM of Bryostatin (Bryo)-1 following treatment of 50 μ M of A β ₂₅₋₃₅ for 48 h. The phosphorylation of PKC detected in cell lysates of PC12 using an anti-phospho-PKC (pan) antibody. The data were presented as the mean \pm SEM. ** p < 0.05 vs. control, # p < 0.05 vs. A β , ## p < 0.01 vs. A β , ### p < 0.001 vs. A β .

to proteins and nucleic acids, which has direct and deleterious consequences in AD^[33]. Evidence has confirmed that the increased amounts of A β peptide elevate the production of ROS^[34,35]; therefore, antioxidants may be merged as therapeutic strategies to attenuate A β -induced neurotoxicity and improve neurological outcomes in AD^[11]. In this study, when the PC12 cells were incubated with A β ₂₅₋₃₅, the contents of ROS significantly increased. Meanwhile, co-treatment with carvacrol or thymol attenuated the A β ₂₅₋₃₅-induced increase of ROS levels. Therefore, the ability of these compounds to protect PC12 cells could be partially attributed to their antioxidant property. Our results are in accordance with other studies denoting that carvacrol and thymol possess strong anti-oxidative activity^[20,21]. Many investigations have reported that antioxidant agents may slow the progression of AD^[10]. For instance, Asadbegi *et al.*^[36] have disclosed that thymol, a natural antioxidant, can be therapeutic against high risk factors for AD. In the present study, administering carvacrol and thymol could effectively ameliorate the induction of oxidative stress in PC12 cells. As oxidative stress is a process always associated with AD, carvacrol and thymol could be considered as potential candidates for AD treatment.

PKC as a phospholipid-dependent protein kinase is a crucial player in various cellular functions either in neuronal and non-neuronal cells^[37]. It is well established that signaling deficits of PKC pathway have an important role in the pathophysiology of neurodegenerative disorders including AD^[38,39]. On the other hand, PKC activators are regarded as a potential candidate for the treatment of AD because of APP processing activation through α -secretase^[6,40]. In this

regard, it has been shown that Bryostatin-1 reduces amyloid plaques in the brain of transgenic AD mice, improves behavioral outcomes and consequently decreases the death rate^[7]. Consistent with these data, our findings obtained from PKC activity assay revealed that carvacrol and thymol reversed A β -triggered PKC inhibition, representing a new mechanism of action for carvacrol and thymol in reducing A β -induced neurotoxicity. Interestingly, Bryostatin-1 acted in the similar way, suggesting the possible modulating role for carvacrol and thymol in PKC activity.

The results of the current study revealed that the protective effects of carvacrol and thymol against A β -induced cytotoxicity are not only attributable to their antioxidant properties but also they may occur through other mechanisms such as PKC stimulation. In accordance with our results, Chen *et al.*^[41] have displayed that the cardioprotective effects of carvacrol may be attributed to its antioxidant property through the activation of the MAPK/ERK signaling pathway, as the common downstream effectors of PKC^[40]. Therefore, considering the possible neuroprotective role of PKC in AD, it is reasonable to suggest administration of carvacrol and thymol as potential disease-modifying agents.

Given the involvement of multiple factors in the pathogenesis of AD as well as the aberrant processing of APP, AD treatment requires multiple drug therapies to address the varied pathological aspects. New potential approaches have been developed to target multiple sites in the brain with single molecular entities for treatment of AD^[42,43]. The results of the present study showed that carvacrol and thymol may present multi-functional properties, including antioxidant

activity and PKC activity modulation. Our results are consistent with a number of previous studies, introduced numerous bio-molecules belonging to terpenoid^[44] or phenolic structures^[45,46] as novel therapeutic candidates with the properties such as radical scavenging and PKC activation.

Our results provide evidence for the neuroprotective effect of carvacrol and thymol against cytotoxicity induced by A β ₂₅₋₃₅. The underlying mechanism might be through decreasing ROS formation as well as stimulating PKC activity. Taken together, these two compounds may have therapeutic potential in preventing or modulating AD. However, further studies are required to clarify all possible neuroprotective mechanisms of carvacrol and thymol in AD models.

ACKNOWLEDGEMENTS

This study has been supported by a Ph.D. grant to the first author through the Graduate Department of the Pasteur Institute of Iran. The authors would like to thank Mrs. Tahereh Bozorgmehr, research assistant from the University of British Columbia for final revision of the manuscript.

CONFLICT OF INTEREST. None declared.

REFERENCES

- Selkoe DJ. Alzheimer's disease results from the cerebral accumulation and cytotoxicity of amyloid beta-protein. *Journal of Alzheimer's disease* 2001; **3**(1): 75-80.
- Habtemariam S. Iridoids and other monoterpenes in the Alzheimer's brain: recent development and future prospects. *Molecules* 2018; **23**(1): pii: E117.
- Duyckaerts C, Delatour B, Potier MC. Classification and basic pathology of Alzheimer disease. *Acta neuropathologica* 2009; **118**(1): 5-36.
- Haass C, Koo EH, Mellon A, Hung AY, Selkoe DJ. Targeting of cell-surface beta-amyloid precursor protein to lysosomes: alternative processing into amyloid-bearing fragments. *Nature* 1992; **357**(6378): 500-503.
- Selkoe DJ. The cell biology of beta-amyloid precursor protein and presenilin in Alzheimer's disease. *Trends in cell biology* 1998; **8**(11): 447-453.
- Etcheberrigaray R, Tan M, Dewachter I, Kuiperi C, Van der Auwera I, Wera S, Qiao L, Bank B, Nelson TJ, Kozikowski AP, Van Leuven F, Alkon DL. Therapeutic effects of PKC activators in Alzheimer's disease transgenic mice. *Proceedings of the national academy of sciences of the United States of America* 2004; **101**(30): 11141-11146.
- Sun MK, Alkon DL. Pharmacology of protein kinase C activators: cognition-enhancing and anti dementic therapeutics. *Pharmacology and therapeutics* 2010; **127**(1): 66-77.
- Leuner K, Schütt T, Kurz CH, Eckert SH, Schiller C, Occhipinti A, Mai S, Jendrach M, Eckert GP, Kruse SE, Palmiter RD, Brandt U, Dröse S, Wittig I, Willem M, Haass C, Reichert AS, Müller WE. Mitochondrion-derived reactive oxygen species lead to enhanced amyloid beta formation. *Antioxidants and redox signaling* 2012; **16**(12): 1421-1433.
- Swomley AM, Butterfield DA. Oxidative stress in Alzheimer's disease and mild cognitive impairment: evidence from human data provided by redox proteomics. *Archives of toxicology* 2015; **89**(10): 1669-1680.
- Behl C, Moosmann B. Oxidative nerve cell death in Alzheimer's disease and stroke: antioxidants as neuroprotective compounds. *Biological chemistry* 2002; **383**(3-4): 521-536.
- Aliev G, Obrenovich ME, Reddy VP, Shenk JC, Moreira PI, Nunomura A, Zhu X, Smith MA, Perry G. Antioxidant therapy in Alzheimer's disease: theory and practice. *Mini-reviews in medicinal chemistry* 2008; **8**(13): 1395-1406.
- Galimberti D, Scarpini E. Disease-modifying treatments for Alzheimer's disease. *Therapeutic advances in neurological disorders* 2011; **4**(4): 203-216.
- Essa MM, Vijayan K, Castellano-Gonzalez G, Memon MA, Braidy N, Guillemin GJ. Neuroprotective effect of natural products against Alzheimer's disease. *Neurochemical research* 2012; **37**(9): 1829-1842.
- Kim MH, Kim SH, Yang WM. Mechanisms of action of phytochemicals from medicinal herbs in the treatment of Alzheimer's disease. *Planta medica* 2014; **80**(15): 1249-1258.
- Tewari D, Stankiewicz AM, Mocan A, Sah, AN, Tzvetkov NT, Huminiecki L, Horbańczuk JO, Atanasov AG. Ethnopharmacological approaches for dementia therapy and significance of natural products and herbal drugs. *Frontiers in aging neuroscience* 2018; **10**: 3.
- Lakey-Beitia J, Berrocal R, Rao KS, Durant AA. Polyphenols as therapeutic molecules in Alzheimer's disease through modulating amyloid pathways. *Molecular neurobiology* 2015; **51**(2): 466-479.
- Syarifah-Noratiqah S, Naina-Mohamed I, Zulfarina MS, Qodriyah HM. Natural polyphenols in the treatment of Alzheimer's disease. *Current drug targets* 2018; **19**(8): 927-937.
- Sawikr Y, Yarla NS, Peluso I, Kamal MA, Aliev G, Bishayee A. Neuroinflammation in Alzheimer's disease: The preventive and therapeutic potential of polyphenolic nutraceuticals. *Advances in protein chemistry and structural biology* 2017; **108**: 33-57.
- Baser KH. Biological and pharmacological activities of carvacrol and carvacrol bearing essential oils. *Current pharmaceutical design* 2008; **14**(29): 3106-3119.
- Wei HK, Xue HX, Zhou ZX, Peng J. A carvacrol-thymol blend decreased intestinal oxidative stress and influenced selected microbes without changing the messenger RNA levels of tight junction proteins in jejunal mucosa of weaning piglets. *Animal* 2017; **11**(2): 193-201.
- El-Sayed el-SM, Mansour AM, Abdul-Hameed MS.

- Thymol and carvacrol prevent doxorubicin-induced cardiotoxicity by abrogation of oxidative stress, inflammation, and apoptosis in rats. *Journal of biochemical and molecular toxicology* 2016; **30**(1): 37-44.
22. Azizi Z, Ebrahimi S, Saadatfar E, Kamalinejad M, Majlessi N. Cognitive-enhancing activity of thymol and carvacrol in two rat models of dementia. *Behavioural pharmacology* 2012; **23**(3): 241-249.
 23. Liu Y, Peterson DA, Kimura H, Schubert D. Mechanism of cellular 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide (MTT) reduction. *Journal of neurochemistry* 1997; **69**(2): 581-593.
 24. LeBel CP, Ischiropoulos H, Bondy SC. Evaluation of the probe 2',7'-dichlorofluorescein as an indicator of reactive oxygen species formation and oxidative stress. *Chemical research in toxicology* 1992; **5**(2): 227-231.
 25. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 2002; **297**(5580): 353-356.
 26. Parvez MK. Natural or plant products for the treatment of neurological disorders: Current knowledge. *Current drug metabolism* 2018; **19**(5): 424-428.
 27. Arigesavan K, Sudhandiran G. Carvacrol exhibits anti-oxidant and anti-inflammatory effects against 1, 2-dimethyl hydrazine plus dextran sodium sulfate induced inflammation associated carcinogenicity in the colon of Fischer 344 rats. *Biochemical and biophysical research communications* 2015; **461**(2): 314-320.
 28. FangFang Li H, Qin T, Li M, Ma S. Thymol improves high-fat diet-induced cognitive deficits in mice via ameliorating brain insulin resistance and up regulating NRF2/HO-1 pathway. *Metabolic brain disease* 2017; **32**(2): 385-393.
 29. Wang P, Luo Q, Qiao H, Ding H, Cao Y, Yu J, Liu R, Zhang Q, Zhu H, Qu L. The Neuroprotective effects of carvacrol on ethanol-induced hippocampal neurons impairment via the antioxidative and anti apoptotic pathways. *Oxidative medicine and cellular longevity* 2017; **2017**: 4079425.
 30. Greene LA, Tischler AS. Establishment of a noradrenergic clonal line of rat adrenal pheochromocytoma cells which respond to nerve growth factor. *Proceedings of the national academy of sciences of the United States of America* 1976; **73**(7): 2424-2428.
 31. Yankner BA, Duffy LK, Kirschner DA. Neurotrophic and neurotoxic effects of amyloid beta protein: reversal by tachykinin neuropeptides. *Science* 1990; **250**(4978): 279-282.
 32. Shearman MS, Ragan CI, Iversen LL. Inhibition of PC12 cell redox activity is a specific, early indicator of the mechanism of beta-amyloid-mediated cell death. *Proceedings of the national academy of sciences of the United States of America* 1994; **91**(4): 1470-1474.
 33. Reddy VP, Zhu X, Perry G, Smith MA. Oxidative stress in diabetes and Alzheimer's disease. *Journal of Alzheimers disease* 2009; **16**(4): 763-774.
 34. Bennett SP, Boyd TD, Norden M, Padmanabhan J, Neame P, Wefes I, Potter H. A novel technique for simultaneous bilateral brain infusions in a mouse model of neurodegenerative disease. *Journal of neuroscience methods* 2009; **184**(2): 320-326.
 35. Kaminsky YG, Marlatt MW, Smith MA, Kosenko EA. Sub cellular and metabolic examination of amyloid-beta peptides in Alzheimer disease pathogenesis: evidence for Abeta(25-35). *Experimental neurology* 2010; **221**(1): 26-37.
 36. Asadbegi M, Komaki A, Salehi I, Yaghmaei P, Ebrahim-Habibi A, Shahidi S, Sarihi A, Soleimani Asl S, Golipoor Z. Effects of thymol on amyloid- β -induced impairments in hippocampal synaptic plasticity in rats fed a high-fat diet. *Brain research bulletin* 2018; **137**: 338-350.
 37. Majewski H, Iannazzo L. Protein kinase C: a physiological mediator of enhanced transmitter output. *Progress in neurobiology* 1998; **55**(5): 463-475.
 38. Cole G, Dobkins KR, Hansen LA, Terry RD, Saitoh T. Decreased levels of protein kinase C in Alzheimer brain. *Brain research* 1988; **452**(1-2): 165-174.
 39. Wang HY, Pisano MR, Friedman E. Attenuated protein kinase C activity and translocation in Alzheimer's disease brain. *Neurobiology aging* 1994; **15**(3): 293-298.
 40. Sun MK, Alkon DL. The "memory kinases": roles of PKC isoforms in signal processing and memory formation. *Progress in molecular biology and translational science* 2014; **122**: 31-59.
 41. Chen Y, Ba L, Huang, W, Liu Y, Pan H, Mingyao E, Shi P, Wang Y, Li S, Qi H, Sun H, Cao Y. Role of carvacrol in cardioprotection against myocardial ischemia/reperfusion injury in rats through activation of MAPK/ERK and Akt/eNOS signaling pathways. *European journal of pharmacology* 2017; **796**: 90-100.
 42. Bolognesi ML, Rosini M, Andrisano V, Bartolini M, Minarini A, Tumiatti V, Melchiorre C. MTDL design strategy in the context of Alzheimer's disease: from lipocrine to memoquin and beyond. *Current pharmaceutical design* 2009; **15**(6): 601-613.
 43. Zhang HY. One-compound-multiple-targets strategy to combat Alzheimer's disease. *FEBS letters* 2005; **579**(24): 5260-5264.
 44. Ji ZH, Liu C, Zhao H, Yu XY. Neuroprotective effect of Biatractylenolide against memory impairment in D-Galactose-induced aging mice. *Journal of molecular neuroscience* 2015; **55**(3): 678-683.
 45. Rebai O, Belkhir M, Sanchez-Gomez MV, Matute C, Fattouch S, Amri M. Differential molecular targets for neuroprotective effect of chlorogenic acid and its related compounds against glutamate induced excitotoxicity and oxidative stress in rat cortical neurons. *Neurochemical research* 2017; **42**(12): 3559-3572.
 46. Levites Y, Amit T, Youdim MB, Mandel S. Involvement of protein kinase C activation and cell survival/ cell cycle genes in green tea polyphenol (-)-epigallocatechin 3-gallate neuroprotective action. *Journal of biological chemistry* 2002; **277**(34): 30574-30580.