# **Antioxidant Enzymes and Oxidative Stress in the Erythrocytes of Iron Deficiency Anemic Patients Supplemented with Vitamins**

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## **ABSTRACT**

**Background:** Iron deficiency anemia is one of the major causes of morbidity and mortality worldwide. Evidences from epidemiological and clinical studies suggest a possible correlation between antioxidant levels and the anemic disease risk. The present work is to investigate antioxidant levels and lipid peroxidation in anemic patients. **Methods:** A number of 30 patients (15 males and 15 females) were selected for the study. Likewise, 30 age- and gender-matched healthy volunteers (15 males and 15 females) were selected with their informed consent. Patients and healthy subjects were supplemented with vitamins C and E for 15 days. The lipid peroxidation both in plasma and erythrocyte lysates was determined by thiobarbituric acid reactive substances and lipid peroxides. The antioxidant vitamins A, C, and E and total antioxidant activity were also analyzed. The antioxidant enzyme superoxide dismutase, catalase, and glutathione peroxidase were also determined. **Results:** Based on analysis, we found that the increase in lipid peroxidation was higher in the anemic subjects before vitamin supplementation, which was statistically significant at P<0.05. The antioxidant enzymes were higher in the patients before antioxidant supplementation when compared with patients after vitamin supplementation. **Conclusion**: Our data revealed higher oxidative stress before vitamin supplementation in iron deficiency anemic patients and after supplementation, lower lipid peroxidation and increased antioxidant vitamins were achieved. *Iran. Biomed. J. 18* (2): 82-87, 2014

Keywords: Iron deficiency anemia, Lipid hydroperoxides (LOOH), Vitamin C, Vitamin E, Thiobarbituric acid reactive substances (TBARS)

### INTRODUCTION

ron deficiency anemia (IDA) is one of the most common causes of morbidity and mortality worldwide, affecting people of all ages in both developed and developing countries. Oxidative stress is known to be a positive contributor for anemia, giving its effects on lipid peroxidation and DNA damage. Reactive oxygen species, such as superoxide, hydrogen peroxide, and hydroxyl radicals are produced during aerobic metabolism [1]. Reactive oxygen species if not removed in a timely manner by an antioxidant system, mammalian cells may encounter oxidative stress that causes destruction of macromolecules and abnormal functions (neurotransmission function and altered immunologic and inflammatory defenses). Ironmediated oxidative damage has been demonstrated in vivo in normal blood cells [2]. The constant autooxidation of hemoglobin generates superoxide radicals, which through spontaneous or enzymatic dismutation yields hydrogen peroxide [1, 2].

There are two main approaches to the use of supplements. They can be used to treat or prevent nutritional deficiency and to reduce the risk of non-deficiency disease and promote optimal health. The chemical versatility of iron has made it one of the most commonly used metals in biological systems. The iron deficiency continues to be a widespread condition affecting millions of people throughout the world [3]. In mammals, multiple physiological processes, including oxygen transport, respiration, DNA synthesis, formation of some neurotransmitters and hormones, xenobiotic metabolism, and certain aspects of host defense use iron-containing proteins [3-8].

The present study was designed to measure the oxidative stress parameters in iron deficiency anemic individuals expressed through plasma thiobarbituric acid reactive substances (TBARS) and lipid hydroperoxides (LOOH), superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx) before vitamins C and E supplementation and also to find out the changes in lipid peroxidation and antioxidant parameters after supplementation.

## MATERIALS AND METHODS

The study group included individuals who divided into two groups: IDA patients (15 male and 15 female) and healthy controls (15 male and 15 female). Healthy controls were selected from Bangalore University staff and students, while anemic individuals were selected from K.C. General Hospital, Malleswaram, Bangalore, India. The healthy individuals and anemic patients were evaluated before and after vitamin supplementation. Height and weight were measured to determine respective body mass index (BMI, kg/m²) and surface area. The age ranged from 20 to 50 years. Criteria for IDA were hemoglobin concentration <11.5 g/dl in women and <13 g/dl in men, plasma iron concentration <45 μg/dl, and total iron-binding capacity >60 μmol/l.

Vitamins C and E (400 mg once daily) was supplemented to anemic as well as healthy controls for 15 days. Blood (5 ml) from normal and anemic individuals was collected before and after vitamin supplementation in a sterile vial containing ethylene diamine tetraacetic acetate. Plasma was collected by centrifugation at 1,500 ×g for 15 min. The red blood cells (RBC) pellet was washed three times with chilled saline and lysed with chilled distilled water in a ratio of 1:4. The lysed RBC was centrifuged to obtain clear RBC hemolysate from cell debris. The hemoglobin level in whole blood was determined by Cyanmethemoglobin method using Beacon Diagnostics kit (Kabilpore, Navasari, India). Iron and total ironbinding capacity were determined by Ferrozine method using Coral Clinical Systems kit, Goa, India.

The lipid peroxidation product, malondialdehyde in plasma, was determined according to Buege and Aust [9] and LOOH was determined according to Jiang et al. [10]. CAT activity in hemolyzed RBC lysates was assayed in the RBC lysates by the method of Sinha [11], and SOD activity in hemolyzed RBC was determined by the method of Kakkar et al. [12], GPx was analyzed in RBC lysates by the method of Rotruck and co-workers [13] and glutathione content by the method of Beutler et al. [14]. Vitamin A was estimated by the methods of Bessey et al. [15], vitamin C (Ascorbic acid) by the method of Natelson [16] using dinitrophenyl hydrazine, and vitamin E by the method of Baker and Frank [17]. Total antioxidant activity was determined according to Benzie and Strain's method [18]. For statistical analysis, the packaged program SPSS (Statistical package for social sciences) for windows (version 13.0, Chicago, IL. USA) was used.

#### **RESULTS**

The biochemical parameters of iron deficiency anemic patients and healthy controls were determined before and after vitamin supplementation (Table 1). IDA patients were divided into male and female groups with respect to vitamin supplementation i.e., before and after vitamin supplementation. BMI both in cases and controls was in normal range, whereas the hemoglobin levels were lower in anemic group as compared to controls. The female patients showed lower hemoglobin levels as compared to male patients. However, after vitamin supplementation, a slight increase in hemoglobin levels was noticed among cases (both male and female group).

The oxidant parameters including TBARS and LOOH were increased before supplementation in cases as compared with controls, but their levels were found to be lowered after vitamin supplementation both in male and female cases. The enzymatic antioxidant parameter, CAT, was increased before vitamin supplementation; however, its level decreased after vitamin supplementation. Similarly, SOD and GPx levels were increased after vitamin supplementation.

The non-enzymatic antioxidant vitamins (A, C, and E) and total antioxidant activity showed increased levels after vitamin supplementation as compared to before vitamin supplementation. However, the levels of non-enzymatic antioxidants were lower than those of healthy controls. Male patients showed slight increased antioxidants as compared to female patients.

Based on independent *t*-test, BMI did not show any significance. Hemoglobin showed a highly significant relation when total case before supplementation was compared to after supplementation. Likewise, highly significant relation was obtained in male cases when before and after vitamin supplementation was compared. However, female cases did not indicate any significance.

The lipid peroxidation parameters did not show any significance except LOOH in females when before and after vitamin supplementation were compared. Total case, male, and female showed a significant relation of enzymatic antioxidants SOD and GPx when before vitamin supplementation group were compared with after vitamin supplementation group. However, CAT in male cases indicated a significant correlation when before and after vitamin supplementations were compared.

The independent t-test was carried out to find the significance between case and control which showed a highly significant (P<0.005) in all the oxidants, enzymatic, and non-enzymatic antioxidant parameters. The anemic case and control were compared before and after vitamin supplementation except CAT and SOD after vitamin supplementation. However, genderwise comparison i.e., males to females before and after vitamin supplementation did not show any statistical significance.

Table 1. Biochemical parameters of anemic patients and controls before and after Vitamin Supplementation

Tests	Suppl	Total	Sig.	Anemic patients				
				Male	Sig.	Female	Sig.	Control
BMI	before after	$21.00 \pm 0.38$ $21.61 \pm 0.08$	0.651	$20.45 \pm 1.03  20.18 \pm 1.20$	0.374	$21.36 \pm 1.70$ $22.49 \pm 1.98$	0.628	$20.48 \pm 1.07^{*}$ $20.68 \pm 1.57^{*}$
Hb	before after	$9.21 \pm 0.04 \\ 10.53 \pm 0.02$	0.000	$9.96 \pm 0.19$ $11.79 \pm 0.16$	0.000	$8.46 \pm 0.09$ $9.27 \pm 0.88$	0.576	$14.71 \pm 0.83^*  15.47 \pm 0.63^*$
Lipid peroxidation parameters:								
TBARS	before after	$6.10 \pm 0.42$ $4.63 \pm 0.73$	0.150	$6.39 \pm 0.10$ $5.14 \pm 2.49$	0.139	$6.14 \pm 0.79 5.43 \pm 0.62$	0.482	$3.21 \pm 1.46^*$ $3.54 \pm 2.42^*$
LOOH	before after	$3.42 \pm 0.82$ $3.31 \pm 0.70$	0.466	$3.12 \pm 0.20$ $2.99 \pm 0.21$	0.875	$7.18 \pm 0.02 \\ 11.86 \pm 0.95$	0.000	$2.51 \pm 1.46^*$ $2.24 \pm 2.42^*$
Enzymatic antioxidant parameters:								
CAT	before after	$833.24 \pm 55.27$ $676.06 \pm 56.08$	0.549	$855.66 \pm 58.07$ $565.17 \pm 34.66$	0.034	$818.29 \pm 53.57$ $744.02 \pm 65.93$	0.331	$358.62 \pm 19.34^{*}$ $180.88 \pm 26.05$
SOD	before after	$48.14 \pm 1.62$ $23.51 \pm 0.52$	0.020	$46.04 \pm 1.74$ $23.14 \pm 2.53$	0.000	$49.55 \pm 1.18$ $23.74 \pm 1.65$	0.000	$58.41 \pm 9.07^*$ $64.75 \pm 5.14$
GPx	before after	$1.30 \pm 0.07$ $3.68 \pm 0.17$	0.000	$1.20 \pm 1.83$ $3.55 \pm 2.08$	0.001	$\begin{array}{c} 1.37 \pm 0.25 \\ 3.77 \pm 0.26 \end{array}$	0.000	$3.85 \pm 1.32^*$ $4.21 \pm 0.43^*$
Non-enzymatic antioxidant parameters:								
VITA	before after	$56.88 \pm 0.83$ $55.92 \pm 0.90$	0.262	$60.04 \pm 1.64$ $58.72 \pm 1.31$	0.057	$54.78 \pm 6.89$ $54.20 \pm 7.60$	0.544	$75.53 \pm 15.65^*$ $82.26 \pm 19.63^*$
VITC	before after	$0.44 \pm 0.03 \\ 0.55 \pm 0.09$	0.015	$0.53 \pm 0.07$ $0.56 \pm 0.03$	0.347	$0.38 \pm 0.03$ $0.54 \pm 0.06$	0.009	$0.48 \pm 0.26^*$ $0.74 \pm 0.25^*$
VITE	before after	$0.52 \pm 0.01$ $0.51 \pm 0.017$	0.455	$0.51 \pm 0.13 \\ 0.55 \pm 0.14$	0.431	$0.52 \pm 0.12$ $0.49 \pm 0.14$	0.333	$1.12 \pm 0.22^* \\ 1.22 \pm 0.34^*$
TAA	before after	$517.00 \pm 11.89$ $581.43 \pm 20.66$	0.009	$533.34 \pm 16.03$ $607.42 \pm 22.22$	0.105	$506.11 \pm 8.17$ $565.50 \pm 18.13$	0.025	$675.17 \pm 12.41^*$ $852.22 \pm 14.36^*$

Italics and \* denote significance; case versus control before and after vitamin supplementation (P<0.005). Suppl, vitamin supplemented group; BMI, body mass index; Hb, hemoglobin; TBARS, thiobarbituric acid reactive substances; LOOH, lipid hydroperoxide; CAT, catalase; SOD, superoxide dismutase; GPx, glutathione peroxidase; VIT, vitmain; TAA, total antioxidant activity.

#### DISCUSSION

IDA is one of the most frequent causes of medical visits because of its high incidence in children, young women, and elderly people, especially if malnutrition is present. It is also one of the leading sings in many diseases or is the first evidence of disease observed in routine blood cell enumeration. In pathological state, an increase in the reactive free radicals creates an imbalance in this ration, thereby macromolecules vulnerable to oxidative damage. The mammalian erythrocyte functions as an oxygen transporter, thus it is permanently in danger of oxidative damage. In order to avert this danger, the erythrocyte is well equipped with antioxidants and antioxidative enzymes [3, 4, 8, 19-21].

Lipid peroxidation has been accepted as being involved in the pathogenesis of various disorders and diseases [22]. Its inhibition is one of the important

roles for antioxidants, as it induces disruption of membrane organization causing changes in fluidity and permeability, inhibition of metabolic processes, and alteration of ion transport. Furthermore, LOOH and their secondary reaction products, such as aldehydes, are cytotoxic, pro-inflammatory, and capable of modifying proteins and DNA bases [22]. Kumerova et al. [23] found that the antioxidant defenses were decreased, and the lipid peroxidation was increased in IDA patients. Tekin et al. [24] discovered that there was no significant difference in SOD and CAT activities in IDA patients relative to controls. In this study, we measured the oxidant and antioxidant status in IDA patients with age- and gender-matched healthy controls before and after vitamin supplementation. As a result of oxygen carrying function, erythrocytes accumulate oxidative damage during their life span in circulation. Protein aggregation, enzyme degradation, and lipid peroxidation have been shown to accompany

red cell aging and are accelerated in conditions such as diabetes and anemia [22]. The impairment of antioxidant defenses and the increase of oxidized lipids or proteins or susceptibility to oxidative stress are well known [23-26].

Isler et al. [27] reported that iron supplementation was used to recover the antioxidant defense system. Also, Bartal et al. [28] showed that although IDA cells are more susceptible to oxidation, they maintain good capacity for recovery. Kurtoglu et al. [29] demonstrated that oxidative stress was decreased after 6 week of iron supplementation, and the condition remained without significant changes until the end of the study, suggesting that oxidant-antioxidant status is regulated at the same time as hemoglobin normalization. They also found that decreased lipid peroxidation, which was expressed as lower malondialdehyde levels in both groups of IDA patients receiving iron supplements and oxidative stress, was lower both at the time of hemoglobin normalization and at the time of saturation of iron body stores.

Nagababu et al. [30] reported that severe anemic RBC undergoes more oxidative stress than normal cells. The damage to the red cell explains that the shorter red cell life-span associated with anemia. Troost et al. [31] study showed that iron ingestion in vivo induces intestinal oxidative damage as indicated by the occurrence of lipid peroxidation. The magnitude of the TBARS concentration in the intestinal fluid samples as a result of iron ingestion was remarkably high. TBARS concentration in the intestinal fluid samples after the iron challenge was roughly 3-5 times higher than those observed in plasma in healthy adults. Our findings were in agreement with those of Bartal et al. [28], Kuypers et al. [32], Chiu and Kuypers [33], and Amirkhizi et al. [34] that reported erythrocytes in IDA were more susceptible to oxidation, but had good capacity for recovery. They also reported a significant decrease in TBARS levels in anemic patients after iron supplementation with ferrous sulfate [32-34]. Although erythrocytes possess highly active antioxidant enzymes, such as SOD, GPx, and CAT compared to other cell types, our results showed that females had slightly increased SOD levels before and after vitamin supplementation as compared to males. Interestingly, GPx also showed increased levels after vitamin supplementation in females as compared to males. decreased levels showed after supplementation in both male and female anemic groups.

A treated group of nine IDA patients with iron, containing drugs, vitamins, and folic acid in the presence of either vitamin C (500 mg) or combination of Geritamin (30,000 UL vitamin A and 0.07 g vitamin E) plus vitamin C (0.5 g) daily for 3 months, explaining that after therapy with iron in IDA patients,

only the level of plasma ceruloplasmin was normalized. Malondialdehyde was found to remain high, the activities of SOD and GPx remained lower, and CAT was found to remain very high as before iron treatment. Our study illustrated that repletion of IDA patients with iron promoted oxidative stress. The causative factor responsible for such effects was hydroxyl radical produced by excess iron. Iron treatment may give rise to increased free iron concentration in cells, which may induce free radical production. In IDA patients treated with iron along with the antioxidant vitamins A, E, and ascorbic acid, the oxidative stress was reduced and the activity of SOD was normalized [35].

Antioxidant vitamins and trace elements contribute to maintain an effective immune response. Maintaining adequate antioxidant status may provide a useful approach in attenuating cell injury and dysfunction. Beneficial effects of oral supplementation of vitamin E (500 mg/day for 6 months) are improvement of anemia correction, atherosclerosis prevention, and increase in osmotic resistance of RBC. A significant decrease in TBARS and increase in nitric oxide, vitamin E, vitamin C, CAT, and SOD were observed on supplementation of vitamin E (400 IU/day for 2 months), which reflected a proper absorption of vitamin by the body [33-35].

By contrast, on supplementation of 100 mg iron form (fumarate) and 500 mg vitamin C to pregnant women showed increased lipid peroxidation and decreased vitamin E level. The potential harmful effects of a high supplementation during pregnancy determined, and deleterious effect of the combined iron/vitamin C supply could be related to its prooxidant properties. Iron deficiency is regarded as the major cause of nutritional anemia, insufficient levels of vitamins B<sub>12</sub>, A, C, and E folate, and riboflavin has been found to be associated with this deficiency. Some clinical trials suggested that vitamin E might have a role as a potential erythropoietic agent in the anemic patients suffering from various types of inherited hemolytic anemia, chronic renal failure patients on hemodialysis, and mildly anemic apparently healthy subjects with borderline anemia. Vitamin E has also the potential to be effectively used for preventing and/or treating some types of human anemia due to its putative role in promoting erythropoiesis, enhancing the integrity and stability of erythrocyte membrane proteins and lipids, and reducing the oxidative stressinduced erythrocyte fragility and lysis resulting in increased RBC survival and hemoglobin level [27, 30-

In conclusion, we found that oxidant status was increased in IDA patients; however, it was slightly decreased after supplementation of vitamins C and E. The non-enzymatic antioxidants were also increased

after vitamin supplementation in anemic patients. Our results suggested that peroxidative damage would be prevented in patients on supplementation of vitamins. Future studies are being carried out in our laboratory based on a larger number of samples and iron supplementation to anemic patients.

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#### REFERENCES

- 1. http://en.wikipedia.org/wiki/Dietary\_supplement.
- Fu Y, Cheng W, Ross D, Lei X G. Cellular glutathione peroxidase protects mice against lethal oxidative stress induced by various doses of diquat. *Proc Soc Exp Biol Med.* 1999 Nov; 222: 164-9.
- 3. Eisenstein RS. Iron regulatory proteins and the molecular control of mammalian iron metabolism. *Annu Rev Nutr.* 2000; 20:627-62.
- 4. Kanth VR, Reddy PU, Raju TN. Behavioral, morphological and physiological shift in the rats administered with tryptophan deficient regimen. *Nutr Hosp.* 2006 Sep-Oct; 21(5):596-603.
- 5. Moxness MS, Brunauer LS, Huestis WH. Hemoglobin oxidation products extract phospholipids from the membrane of human erythrocytes. *Biochemistry*. 1996 *Jun*; 35(22):7181-7.
- 6. Pietta PG. Flavonoids as antioxidants. *J Nutr Proct.* 2000 Jul; 63(7):1035-42.
- 7. Wu Z, Zhao Y, Zhao B. Superoxide anion, uncoupling proteins and Alzheimer's disease. *J Clin Biochem Nutr.* 2010 May; 46(3):187-94.
- 8. Oladiji TA. Tissue levels of iron, copper, zinc and magnesium in iron deficient rats. *Biokemistri.* 2003 Jan; 14(1):75-81.
- 9. Buege JA, Aust SD. Microsomal lipid peroxidation. *Methods Enzymol.* 1978; 52:302-10.
- 10. Jiang ZY, Hunt JV, Wolff SP. Ferrous ion oxidation in the presence of xylenol orange for detection of lipid hydroperoxides in low density lipoprotein. *Anal Biochem.* 1992 May; 202(2):384-9.
- 11. Sinha AK. Colorimetric assay of catalase. *Anal Biochem.* 1972 Jun; 47(2):389-94.
- 12. Kakkar P, Das B, Vishwanathan PN. A modified spectrophotometric assay of superoxide dismutase. *Indian J Biochem Biophys. 1984 Apr; 21(2):130-2.*
- 13. Rotruck JT, Pope AL, Ganther HE, Swanson AB,

- Hafeman DG, Hoekstra WG. Selenium: biochemical role as a component of glutathione peroxidase. *Science*. 1973 Feb: 179(4073):588-90.
- 14. Beutler E, Duron O, Kelley BM. Improved method for the determination of blood glutathione. *J Lab and Clin Med. 1963 May; 61(5):882-8.*
- 15. Bessey OA, Lowry OH, Brock MJ, Lopez JA. The determination of vitamin A and carotene in small quantities of blood serum. *J Biol Chem. 1946 Nov;* 166(1):177-88.
- Natelson S. Determination of Ascorbic Acid Using 2, 4-Dinitrophenyl Hydrazine. In: Charles C Thomas editor. Techniques of Clinical Chemistry. 3<sup>rd</sup> ed, USA: Spring Field; 1971. 165-6.
- 17. Baker H, Frank O. Determination of Vitamin E. New York: Wiley, USA. 1968.
- 18. Benzie IFF, Strain JJ. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": The FRAP assay. *Anal Biochem.* 1996 Jul; 239(1):70-6.
- 19. Grzelak A, Kruszewski M, Macierzyńska E, Piotrowski Ł, Pułaski Ł, Rychlik B, et al. The effects of superoxide dismutase knockout on the oxidative stress parameters and survival of mouse erythrocytes. *Cell Mol Biol Lett.* 2009; 14(1):23-34.
- 20. Takami T, Sakaida I. Iron regulation by hepatocytes and free radicals. *J Clin Biochem Nutr. 2011 MAr*; 48(2):103-8.
- 21. Samhan-Arias AK, Tyurina YY, Kagan VE. Lipid antioxidants: free radical scavenging *versus* regulation of enzymatic lipid peroxidation. *J Clin Biochem Nutr.* 2011 Jan: 48(1):91-5.
- 22. Jankowska EA, Rozentryt P, Witkowska A, Nowak J, Hartmann O, Ponikowska B, et al. Iron deficiency: an ominous sign in patients with systolic chronic heart failure. Eur Heart J. 2010. doi:10.1093/eurheartj/ehq158.
- Kumerova A, Lece A, Skesters A, Silova A, Petuhovs V. Anemia and antioxidant defense of red blood cells. *Mater Med Pol. 1998 Jan-Jun; 30(1-2):12-5.*
- 24. Tekin D, Yavuzer S, Tekin m, Akar N, Cin S. Possible effects of antioxidant status on increased platelet aggregation in childhood iron-deficiency anemia. *Pediatr Int.* 2001 Feb; 43(1):74-7.
- 25. Niwano Y, Saito K, Yoshizaki F, Kohno M, Ozawa T. Extensive screening for herbal extracts with potent antioxidant properties. *J Clin Biochem Nutr.* 2011 Jan; 48(1):78-84.
- 26. Niki E, Omata Y, Fukuhara A, Saito Y, Yoshida Y. Assessment of radical scavenging capacity and lipid peroxidation inhibiting capacity of antioxidant. *J Agric Food Chem.* 2008 Sep; 56(18):8255-60.
- 27. Isler M, Delibas N, Guclu M, Gultekin F, Sutcu R, Bahceci M, et al. Superoxide dismutase and glutathione peroxidase in erythrocytes of patients with iron deficiency anemia: effects of different treatment modalities. *Croat Med J. 2002 Feb; 43(1):16-9.*
- 28. Bartal M, Mazor D, Dvlinski A, Meyerstein N. Iron deficiency anemia: recovery from *in vitro* oxidative stress. *Acta Haematol.* 1993; 90:94-8.
- Kurtoglu E, Ugur A, Baltaci A K, Undar L. Effect of iron supplementation on oxidative stress and antioxidant status in iron deficiency anemia. *Biol Trace Elem Res.*

- 2003 Winter; 96(1-3):117-23.
- 30. Nagababu E, Gulyani S, Earley CJ, Cutler RG, Mattson MP, Rifkind JM. Iron deficiency anemia enhances red blood cell oxidative stress. *Free Radic Res.* 2008 Sep; 42(9):824-9.
- 31. Troost FJ, Saris WH, Haenen GR, Bast A, Brummer RJ. New method to study oxidative damage and antioxidants in the human small bowel: effects of iron application. *Am J Physiol Gastrointest Liver Physiol*. 2003 Aug; 285(2):G354-9.
- 32. Kuypers FA, Schoot MA, Scott MD. Phospholipid composition and organization in model beta-thalassemic erythrocytes. *Amer J Hematol.* 1996 Jan; 51(1):45-54.
- 33. Chiu D, Kuypers F, Lubin B. Lipid peroxidation in human red cells. *Semin Hematol.* 1989 Oct; 26(4):257-76
- 34. Amirkhizi F, Siassi F, Minaie S, Djalali M, Rahimi A, Chamari M. Assessment of lipid peroxidation and activities of erythrocytes cytoprotective enzymes in women with iron deficiency anemia. *J Res Med Sci.* 2008; 13(5):248-54.
- 35. Gadjeva V, Kuchukova D, Georgieva R. Vitamin combinations reduce oxidative stress and improve antioxidant status in patients with iron deficiency anemia. *Comp Clin Path.* 2005; 14:99-104.