Evaluation of mRNA Expression of *CD244* and Its Adapter Molecules in CD8⁺ T Cells in Acute Leukemia

Maryam Mohammadi¹, Hossein Asgarian-Omran^{1,2}, Ahmad Najafi¹, Reza Valadan³, Hossein Karami⁴, Mohammad Naderisoraki⁴, Ehsan Zaboli^{2,5}, Mohammad Eslami^{2,5}, Mohsen Tehrani^{1,2*}

¹Department of Immunology, School of Medicine, Mazandaran University of Medical Sciences, Sari, Iran; ²Gastrointestinal Cancer Research Center, Mazandaran University of Medical Sciences, Sari, Iran; ³Molecular and Cell-Biology Research Center, Mazandaran University of Medical Sciences, Sari, Iran; ⁴Thalassemia Research Center (TRC), Hemoglobinopathy Institute, Mazandaran University of Medical Sciences, Sari, Mazandaran Iran; ⁵Department of Hematology and Oncology, Imam Khomeini hospital, Mazandaran University of Medical Sciences, Sari, Iran; ⁴Thalassemia Research Center

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Mohammadi M, Asgarian-Omran H, Najafi A, Valadan R, Karami H, Naderisoraki M, Zaboli E, Eslami M, Tehrani M. Evaluation of mRNA Expression of *CD244* and Its Adapter Molecules in CD8⁺ T Cells in Acute Leukemia. *Iranian biomedical journal* 2022; 27(4): 214-218. adapter molecules, in CD8⁺ T cells in acute leukemia. **Methods**: Blood samples were obtained from 21 ALL and 6 AML patients and 20 control subjects. Relative gene expression of *CD244*, *SAP*, *EAT-2*, and LncRNA-GSTT1-AS1 were evaluated using qRT-PCR. **Peculte:** Expression of *CD244*, *SAP*, and *EAT*, 2 were significantly lower in CD8⁺

Background: This study investigated the role of the ICR, CD244, and its

ABSTRACT

Results: Expression of *CD244*, *SAP*, and *EAT-2* were significantly lower in CD8⁺ T cells from ALL patients than those from control subjects. Interestingly, the expression of *SAP* was much lower than that of *CD244*, indicating a lower ratio of *SAP* to *CD244*. Also, *SAP* expression was significantly lower in AML patients compared to the control group. Expression of LncRNA-GSTT1-AS1 showed no significant difference in ALL and AML patients compared to control subjects.

Conclusion: The low *SAP/CD244* expression ratio in CD8⁺ T cells in ALL suggests an inhibitory role for *CD244* in ALL. *DOI: 10.61186/ibj.3843*

Keywords: Acute myeloid leukemia, CD244, EAT-2

Corresponding Author: Mohsen Tehrani

Department of Immunology, School of Medicine, Mazandaran University of Medical Sciences, Sari, P.O.Box: 48471-91971, Iran; Tel.: (+98-11) 33543081; Fax: (+98-11) 33543248; E-mail: drmtehrani@gmail.com

INTRODUCTION

cute leukemia, including ALL and AML, are two common leukemias throughout the world^[1,2]. Treatments for leukemia include chemotherapy, radiation therapy, and hematopoietic stem cell transplantation, which have many side effects and limited efficacy^[1]. One of the problems in chronic infections and cancers, which the immune system suffers, is the dysfunction of T cells, known as T-cell exhaustion^[3]. Exhausted T cells show overexpression of some ICRs, which bind to their ligands on the surface of antigen-presenting and tumor cells. The most important ICRs are PD-1, Tim-3, LAG-3, TIGIT, and CD244. Blockade of these receptors, socalled immune checkpoint blockade, using monoclonal antibodies or other approaches, improved T cell function and slowed progression of the disease in animal models of cancer, as well as in human cancers^[4]. In leukemia, however, the role of ICRs has not fully been investigated; thus, immune checkpoint blockade therapy in leukemias has not yet been approved by the US Food and Drug Administration (FDA).

CD244, a member of the SLAM family, is expressed on the surface of several immune cells, including Tcells, NK cells, basophils, monocytes, dendritic cells, and myeloid-derived suppressor cells. Its ligand, CD48, is expressed on the surface of various hematopoietic cells^[5]. Once CD244 binds to its ligand, its ITSM motifs binds to the adapter molecule, SAP, which leads to an

List of Abbreviations:

ALL: acute lymphoblastic leukemia; AML: acute myeloid leukemia; cDNA: complementary DNA; EAT-2: EWS/FLI1-activated transcript 2; ICR: immune-checkpoint receptor; ITSM: immune receptor tyrosine-based switch motif; LncRNA: long noncoding RNA; PBMC: peripheral blood mononuclear cell; PD-1: programmed cell death protein 1; qRT-PCR: quantitative reverse transcription polymerase chain reaction; SAP: immune receptor tyrosine-based switch motif-associated protein; SLAM: signaling lymphocyte activation molecule

activating signal. However, it can bind to the other adapter molecule, EAT-2, resulting in an activating or inhibitory signal^[6,7]. It is supposed that expression levels, availability, and competitive binding of the adapter molecules, SAP and EAT-2, determine whether CD244 mediates an activating or inhibitory signal, as adequate concentrations of functional SAP leads to CD244 activating signals^[4]. Therefore, the ratio of SAP to CD244 plays an important role in determining an activating or inhibitory role for this molecule^[4]. CD244 has been shown to be overexpressed in the exhausted T cells in chronic infections, such as in hepatitis B^[8], as well as in some malignancies, including multiple myeloma^[9] and AML^[10]. Of note, blockade of CD244, using monoclonal antibodies, increased the population of virus-specific T cells and expression of CD107a, perforin, IFN- γ , TNF- α , and IL-6 in *in vitro* models^[4]. Study of CD244 and its adapter molecules, SAP and EAT-2, in CD8⁺ T cells in acute leukemia would be helpful to further understand T cell exhaustion-related pathways and to find therapeutic targets for acute leukemia. Hence, this study aimed to investigate the expression of CD244 and its related adapter molecules in CD8⁺ T cells in acute leukemia^[6].

MATERIALS AND METHODS

Patients and controls

The sample size was calculated based on previous studies^[11-13]. Twenty-one ALL patients and six AML patients, referred to the Outpatient Clinic or the Hematology and Oncology Wards of Imam Khomeini Hospital and Bou-Ali Sina Hospital (both affiliated to Mazandaran University of Medical Sciences, Sari, Iran), were included in the study. Diagnosis of the disease was carried out via clinical evaluations, blood cell count, and morphology, along with the immunophenotyping of blood cells in the peripheral blood sample or the bone marrow, according to World Health Organization criteria^[14]. A questionnaire, including (WHO) demographic, clinical and laboratory information, was completed separately for each study subject. Twenty healthy volunteers were also included in the study as a control group. Patients with chronic viral diseases, such as hepatitis B, hepatitis C and HIV, those with a history of autoimmune diseases or malignancies other than ALL and AML, or taking chemotherapy or immunesuppressive drugs, were excluded from the study. In total, 21 ALL (8 males and 13 females, mean age 15.94 years) and 6 AML (5 males and 1 female, mean age 45.66 years) patients, as well as 20 control subjects (9 males and 11 females, mean age 22.5 years) participated in the study. An amount of 8-10 mL of heparinized peripheral blood sample was taken from each study subject.

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Heparinized peripheral blood samples were collected from patients and controls, and PBMCs were isolated by density gradient centrifugation on Lymphosep (Biosera, Nuaille, France). Isolated PBMCs were washed twice with RPMI-1640 culture medium (Biosera). Viability of the isolated cells was determined by trypan blue staining. CD8⁺ T cells were then purified from PBMCs using the magnetic-activated cell separation by means of CD8 microbeads (Miltenyi Biotec, Germany). To check the purity of the isolated CD8⁺ T cells, 2×10^5 cells were stained with anti-CD8-FITC (Clone SK1, 0.125 µg/test, eBioscience, San Diego, US) and anti-CD3-PE (Clone UCHT1, 0.06 µg/test, eBioscience). Appropriate amounts of isotype-matched control antibodies were also used to subtract the background staining. Samples were then analyzed on a Partec PAS flow cytometer system (Partec GmBH, Munster, Germany) using the FlowMax software (version 2.82).

Quantitative reverse transcription polymerase chain reaction

Total RNA was isolated from CD8⁺ T cells using the FavorPrep Blood/Cultured Cell Total RNA Mini Kit (Favorgen, Taiwan) based on the manufacturer's protocol. The quantity and quality of the isolated RNA were confirmed by nano-spectrophotometer and electrophoresis, respectively. The cDNA was synthesized using a cDNA synthesis kit (Yekta Tajhiz Azma, Tehran, Iran). qRT-PCR was performed using Real Q Plus 2× Master Mix (High Rox, Ampliqon, Denmark) reagent on an ABI Step one plus Real-Time system (Applied Biosystems, USA) with specific primers for CD244, SAP, EAT-2, and LncRNA-GSTT1-AS1, as well as ACTB, as a housekeeping gene. PCR was performed using 10 pmol of each forward and reverse primer, 1 µL of cDNA, 10 µL of Master mix, and 7 µL of PCR grade water and amplified at 95 °C for initial denaturation, followed by 40 cycles at 95 °C for 15 seconds, 60 °C for 30 seconds, and extension at 72 °C for 30 seconds. After normalizing to ACTB, relative expression levels of CD244, SAP, EAT-2, and LncRNA-GSTT1-AS1 was determined using the $2^{-\Delta\Delta Ct}$ method^[15].

Statistical analysis

Statistical analyses were performed using SPSS 18 software. Results are shown as mean \pm SEM. Using Kolmogorov-Smirnov test, normal distribution of the studied parameters was checked, and parametric or non-parametric tests were used. P < 0.05 was considered statistically significant.

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RESULTS

mRNA expression of CD244 and its adapter molecules in patients and controls

Expression of CD244 and EAT-2 in patients with ALL was significantly lower than the control group (p =0.0225 and p = 0.0375, respectively), while their expression was not significantly different between AML patients and the control group (p = 0.7785 and p =0.5670; Fig. 1A and 1B). SAP expression in both ALL and AML patients was significantly lower than the control group (p = 0.0031 and p = 0.0155, Fig. 1C).

LncRNA-GSTT1-AS1 expression in patients and controls

The relative mRNA expression of LncRNA-GSTT1-AS1, a long noncoding RNA, related to CD244 expression, known as LncRNA-CD244,^[16] was evaluated in CD8⁺ T cells from patients with ALL and AML, as well as the control group. The results of qRT-PCR showed that although its expression was lower in both ALL and AML patients than the control group, the differences were not statistically significant (p = 0.3273and p = 0.5457, respectively; Fig. 1D).

Correlations between CD244 and SAP expression in **ALL** patients

Correlation analysis of the expression level of CD244 with that of its adapter molecules showed a direct correlation between CD244 and SAP expression in ALL patients (r = 0.53; p = 0.013; Fig. 2A). However, the correlations between the expression level of CD244 with that of EAT-2 and LncRNA-GSTT1-AS1 were not statistically significant (Fig. 2B and 2C).

DISCUSSION

CD244 is expressed on the surface of NK cells, $\gamma\delta$ T cells, dendritic cells, and other immune cells^[4]. Basically, CD244 has been recognized as an ICR; however, its adapter molecules, including SAP and EAT-2, play important roles in determining whether CD244 produces an activating or inhibitory signal^[4]. In this study, we focused on acute leukemia, including ALL and AML, and examined the relative mRNA expression of CD244 and its downstream adapter molecules, SAP and EAT-2, in CD8⁺ T cells.

Previously, it has been shown that CD244 is overexpressed on exhausted CD8+ T cells in chronic infections, and its expression correlates with that of PD-1^[7,17]. CD244 also showed an overexpression on the exhausted CD8⁺ T cells in some human cancers, like melanoma^[18,19] and multiple myeloma^[9]. In the present study, in ALL patients, CD244 expression was significantly lower in CD8⁺ T cells than the control

subjects, while it was not significantly different between AML patients and controls. Since these results were not in line with the previous studies, we then measured the relative expression of SAP and EAT-2 in CD8⁺ T cells in ALL and AML patients. SAP binds to ITSM domainin the cytoplasmic part of CD244, which induces

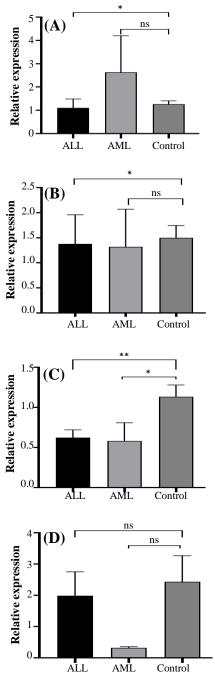


Fig. 1. Relative mRNA expression of CD244 (A), EAT-2 (B), SAP (C), and LncRNA-GSTT1-AS-1 (Lnc-CD244; D) in ALL and AML patients and control subjects. The mRNA expression of CD244, SAP, EAT-2, and LncRNA-GSTT1-AS-1 was determined using qRT-PCR, normalized to that of ACTB, and expressed as relative expression. Vertical bars represent mean ± SEM (*p < 0.05; **p < 0.01)

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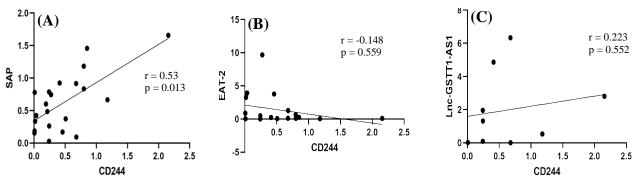


Fig. 2. Correlations between mRNA expressions of *CD244* and its adapter molecules in ALL patients. Pearson's correlation analyses between *CD244* and *SAP* (A), *CD244* and *EAT-2* (B), and *CD244* and LncRNA-GSTT1-AS1 (C) gene expressions in ALL patients.

an activating signal. In the absence of sufficient concentrations of SAP, CD244 binds to phosphatases, and inhibitory signal transduction occurs. Alternatively, CD244 could bind to EAT-2, which is associated with activating or inhibitory signal transduction^[7]. In our study, although both CD244 and SAP showed reduced expression in CD8+ T cells in ALL patients, SAP expression was much lower than that of CD244. Also, expression of CD244 and SAP were positively correlated with each other. As mentioned above, it has been shown that signaling function of CD244 depends on the concentration of its downstream adapter molecules. In the presence of adequate expression of SAP, it is more likely that this adapter molecule binds to the ITSM domain of the CD244 molecule, resulting in an activating signal; however, in the absence of enough expression of SAP, other adapter molecules in the downstream of CD244, such as phosphatases, bind to the CD244 tyrosine domain, leading to an inhibitory signal^[4]. Since our results indicated that SAP expression in CD8⁺ T cells from ALL patients was much less than that of CD244, we assumed that the ratio of CD244 to SAP expression was lower in CD8⁺ T-cells in ALL than in control subjects, thus the receptor would exhibit an inhibitory function.

In this study, we measured the expression level of the LncRNA-GSTT1-AS1 in CD8⁺ T cells from ALL and AML patients. LncRNAs are a recently discovered class of RNAs with a length of more than 200 base pairs that do not encode a protein^[20]. Many LncRNAs have been identified in mammalian genomes that regulate the expression of genes related to T-cell differentiation^[21]. A previous study also found out that in tuberculosis, as a chronic infection, the expression of CD244 and then LncRNA-GSTT1-AS1, known as LncRNA-CD244, increased in CD8⁺ T cells, which led to decreased IFN- γ and TNF- α production and impaired immune response to infection^[16]. Accordingly, to study the role of LncRNA-GSTT1-AS1 in T-cell exhaustion in hematologic malignancies, we evaluated its expression level in CD8⁺ T cells from ALL and AML patients.

However, our results showed that the expression of LncRNA-GSTT1-AS1 was not significantly different in CD8⁺ T cells from ALL and AML patients compared to those from control subjects. Based on the results, *SAP* expression was significantly lower in AML patients than in control subjects, while *CD244* and *EAT-2* did not show significant differences between the patients and controls. This finding might be because only six AML patients were enrolled in our study. This is a limitation for this study, which suggests future studies enrolling larger numbers of AML patients.

In conclusion, this study demonstrated a low SAP/CD244 expression ratio in CD8⁺ T cells in ALL patients. Therefore, it can be assumed that CD244 signaling plays an inhibitory role in CD8⁺ T cells in ALL patients, which is in line with T-cell exhaustion in ALL. Further studies are required to perform functional assays on CD8⁺ T cells to elucidate the exact role of CD244 in acute leukemia.

DECLARATIONS

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Ethical statement

All the patients and healthy individuals signed the consent form, which was based on the rules of the Ethics Committee of Mazandaran University of Medical Sciences, Sari, Iran (ethical code: IR.MAZUMS. IMAMHOSPITAL.REC.1399.8204).

Data availability

The raw data supporting the conclusions of this article are available from the corresponding author upon reasonable request.

Author contributions

MM: carried out the assays and contributed to data

collection and analysis, and prepared the manuscript; HAO: designed and conducted the research; AN and RV: helped in PCR optimization and analysis of PCR data; HK, EZ, and MT: provided the samples; MT: designed and conducted the research and prepared the manuscript. All authors read and approved the final version of manuscript.

Conflict of interest

None declared.

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REFERENCES

- 1. Terwilliger T, Abdul-Hay M. Acute lymphoblastic leukemia: a comprehensive review and 2017 update. *Blood cancer journal* 2017; **7**(6): e577.
- 2. Meyers J, Yu Y, Kaye JA, Davis KL. Medicare fee-forservice enrollees with primary acute myeloid leukemia: an analysis of treatment patterns, survival, and healthcare resource utilization and costs. *Applied health economics and health policy* 2013; **11**(3): 275-286.
- Wherry EJ, Kurachi M. Molecular and cellular insights into T cell exhaustion. *Nature reviews immunology* 2015; 15(8): 486-499.
- Agresta L, Lehn M, Lampe K, Cantrell R, Hennies C, Szabo S, Wise-Draper T, Conforti L, Hoebe K, Janssen EM. CD244 represents a new therapeutic target in head and neck squamous cell carcinoma. *Journal for immunotherapy of cancer* 2020; 8(1):e000245.
- Chen R, Relouzat F, Roncagalli R, Aoukaty A, Tan R, Latour S, Veillette A. Molecular dissection of 2B4 signaling: implications for signal transduction by SLAMrelated receptors. *Molecular and cellular biology* 2004; 24(12): 5144-5156.
- Wang N, Calpe S, Westcott J, Castro W, Ma C, Engel P, Schatzle J, Terhorst C. The adapters EAT-2A and -2B are positive regulators of CD244- and CD84-dependent NK cell functions in the C57BL/6 mouse. *The journal of immunology* 2010; **185**(10): 5683-5687.
- Roncagalli R, Taylor JER, Zhang S, Shi X, Chen R, Cruz-Munoz ME, Yin L, Latour S, Veillette A. Negative regulation of natural killer cell function by EAT-2, a SAP-related adaptor. *Nature immunology* 2005; 6(10): 1002-1010.
- Raziorrouh B, Schraut W, Gerlach T, Nowack D, Grüner NH, Ulsenheimer A, Zachoval R, Wächtler M, Spannagl M, Haas J, Diepolder HM, Jung MC. The immunoregulatory role of CD244 in chronic hepatitis B infection and its inhibitory potential on virus-specific CD8⁺ T-cell function. *Hepatology* 2010; **52**(6): 1934-1947.
- Zelle-Rieser C, Thangavadivel S, Biedermann R, Brunner A, Stoitzner P, Willenbacher E, Greil R, Jöhrer K. T cells in multiple myeloma display features of exhaustion and senescence at the tumor site. *Journal of hematology and oncology* 2016; 9(1): 116.

- Tan J, Chen S, Lu Y, Yao D, Xu L, Zhang Y, Yang L, Jie Chen J, Lai J, Yu Z, Zhu K, Li Y. Higher PD-1 expression concurrent with exhausted CD8⁺ T cells in patients with de novo acute myeloid leukemia. *Chinese journal of cancer research* 2017; **29**(5): 463-470.
- Chauvin JM, Pagliano O, Fourcade J, Sun Z, Wang H, Sander C, Kirkwood JM, Chen THT, Maurer M, Korman AJ, Zarour HM. TIGIT and PD-1 impair tumor antigenspecific CD8⁺ T cells in melanoma patients. *The journal of clinical investigation* 2015; **125**(5): 2046-2058.
- Toor SM, Murshed K, Al-Dhaheri M, Khawar M, Abu Nada M, Elkord E. Immune checkpoints in circulating and tumor-infiltrating CD4⁺ T Cell subsets in colorectal cancer patients. *Frontiers in immunology* 2019; 10:2936.
- Gros A, Robbins PF, Yao X, Li YF, Turcotte S, Tran E, Wunderlich JR, Mixon A, Farid S, Dudley ME, Hanada KI, Almeida JR, Darko S, Douek DC, Yang JC, Rosenberg SA. PD-1 identifies the patient-specific CD8⁺ tumor-reactive repertoire infiltrating human tumors. *The journal of clinical investigation* 2014; **124**(5): 2246-2259.
- Swerdlow SH, Campo E, Pileri SA, Harris NL, Stein H, Siebert R, Advani R, Ghielmini M, Salles GA, Zelenetz AD, Jaffe ES. The 2016 revision of the World Health Organization classification of lymphoid neoplasms. *Blood* 2016; **127**(20): 2375-2390.
- 15. Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods* 2001; **25**(4): 402-408.
- 16. Wang Y, Zhong H, Xie X, Chen CY, Huang D, Shen L, Zhang H, Chen ZW, Zeng G. Long noncoding RNA derived from CD244 signaling epigenetically controls CD8⁺ T-cell immune responses in tuberculosis infection. *Proceedings of the national academy of sciences of the United States of America* 2015; **112**(29): E3883-E3892.
- Schlaphoff V, Lunemann S, Suneetha PV, Jaroszewicz J, Grabowski J, Dietz J, Helfritz F, Bektas H, Sarrazin C, Manns MP, Cornberg M, Wedemeyer H. Dual function of the NK cell receptor 2B4 (CD244) in the regulation of HCV-specific CD8⁺ T cells. *PLoS pathogens* 2011; 7(5): e1002045.
- Lichtenegger FS, Schnorfeil FM, Emmerig K, Neitz JS, Beck B, Draenert R, Hiddemann W, Subklewe M. Pseudo-Exhaustion Of CD8⁺ T cells in AML. *Blood* 2013; **122**(21): 2615.
- Baitsch L, Baumgaertner P, Devêvre E, Raghav SK, Legat A, Barba L, Wieckowski S, Bouzourene H, Deplancke B, Romero P, Rufer N, Speiser DE. Exhaustion of tumor-specific CD8⁺ T cells in metastases from melanoma patients. *The journal of clinical investigation* 2011; **121**(6): 2350-2360.
- Rinn JL, Chang HY. Genome regulation by long noncoding RNAs. *Annual review of biochemistry* 2012; 81: 145-166.
- Hu G, Tang Q, Sharma S, Yu F, Escobar TM, Muljo SA, Zhu J, Zhao K. Expression and regulation of intergenic long noncoding RNAs during T cell development and differentiation. *Nature immunology* 2013; 14(11): 1190-1198.

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