

Validating *MARK2* Gene Polymorphism as a Predictor of Response to Lithium Treatment in Bipolar Patients

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OPEN ACCESS

Received: 26 October 2021
Accepted: 16 November 2021
Published online: 26 December 2021

ABSTRACT

Background: Lithium is a therapeutic option for the treatment of the acute phase of the bipolar disorder and long-term management of this disorder. However, it is estimated that 10 to 60% of patients do not properly respond to this medication.

Methods: To investigate the role of *MARK2* gene in response to lithium, we genotyped the *MARK2* rs10792421 polymorphism in Iranian bipolar patients using ARMS-PCR.

Results: Results of this study showed a significant association of this polymorphism with response to lithium. The A allele was more frequent in the responder than the non-responder group and also in the semi-responder group compared to the non-responder group in the codominant model of analysis. AA and AG genotypes were more frequent in both the responder and semi-responder groups compared to the non-responder group in dominant model of analysis.

Conclusion: Based on the findings of the current study, the rs10792421 variant of *MARK2* gene could be considered as a potential biomarker for predicting the treatment outcome of bipolar disorder type 1 in Iranian population. **DOI:** 10.52547/ibj.26.2.110

Citation:
Aghabozorg Afjeh SS, Shams J, Hamednia S, Boshehri B, Amini A, Omrani A, Omrani MD. Validating *MARK2* Gene Polymorphism as a Predictor of Response to Lithium Treatment in Bipolar Patients. *Iranian biomedical journal* 2022; 26(2): 110-115.

Keywords: Biomarkers, Bipolar, Lithium, Genotyping

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INTRODUCTION

Lithium, as a well-established therapeutic option, has long been used for the management of severe mental illnesses^[1]. Regarding mood-stabilizing properties, lithium has been known as the main medical option for the acute and long-term treatment of bipolar disorder^[2]. However, it is estimated that 10 to 60% of patients do not show an appropriate response to the medication, and the introduction of new medications has not improved

these numbers^[3]. Nonadherence of patients to lithium confers a high risk of relapse and may make them susceptible to rebound depression and mania^[4,5]. Regarding the high rate of nonadherence and side effects caused by an inappropriate dose of lithium treatment, the determination of the right dose of this drug is considered a significant treatment challenge^[6]. Long-term studies support the effect of genetic variations in the various responses of individuals to medications^[7]. Understanding the molecular pathway of the function of medications would help to find

List of Abbreviations:

ARMS-PCR, amplification Refractory Mutation System-PCR; **GSK3-β**, glycogen synthase kinase; **MARK**, microtubule affinity regulating kinase; **PINK1**, PTEN 1-stimulating kinase

variants that alter the therapeutic response.

One of the significant actions of lithium is investigated in tauopathies^[8]. Evidence has also shown that lithium is prescribed as a neuroprotective agent in some neurodegenerative disorders such as Parkinson's or Alzheimer's diseases^[9]. Due to the absence of gliosis (neurodegeneration marker), bipolar disorder is not considered as a classic neurodegenerative disorder, but has specific patterns of glial loss and elevated choline levels^[8], and is considered as a neurodevelopmental disorder^[10]. Many studies have suggested that lithium may target agents affecting neurotransmission and signal transduction, which results in cell-surface receptor modulation and the subsequent effect on the activity of major regulatory systems. These events may result in activation of relevant transcription factors and their target genes' expression^[11]. Serine-threonine protein kinase-related cascades is among the main influenced signaling pathways. These cascades are reported to be activated by lithium^[12]. Serine-threonine protein kinases are involved in the establishment of neuronal polarity, neurite outgrowth, and regulation of microtubule dynamics, axogenesis, neuronal migration, and inhibition of microtubule-dependent accumulation^[13,14]. Among them, MARK family of protein kinases has been under focus as exerts important roles in phosphorylation of certain sites in tau protein. This phosphorylation is found to be increased in abnormal tau of Alzheimer's brain tissue, resulting in detachment of tau and microtubules^[14].

Several studies have reported similar pathophysiological processes in Alzheimer's disease and bipolar disorder. Further, impairments of episodic memory and executive dysfunction are shared between these disorders^[15-17]. Therefore, it is expected that these two disorders represent some common genetic basis and overlap in some pathogenic signaling pathways. The *MARK2* gene has been reported to be associated with Alzheimer's disease and bipolar disorder. This gene is widely expressed in the brain throughout the life and has shown spatial and temporal relationships with both neurodevelopmental and neurodegenerative processes^[10]. Activated *MARK2*, along with neurofibrillary tangles, is located in the same area in the brains of Alzheimer's patients. Also, more target areas of *MARK2* on tau in transgenic mice of the Tauopathies model emphasize the importance of *MARK2* in this disease^[18].

Lithium is prescribed in Alzheimer's disease to modulate tau phosphorylation and amyloid- β production, as well as improve memory impairment^[19]. Lithium has been shown to reduce GSK3- β activity^[20]. It has been demonstrated that GSK3- β is responsible

for *MARK2* phosphorylation and activation, which give rise to tau phosphorylation at Ser-262^[21,22]. Tau phosphorylation at this particular site is mainly mediated by a GSK-3 β *MARK2* pathway^[23]. Hence, common variants of *MARK2* gene could be investigated as the candidate modulators of lithium therapeutic action in Alzheimer's disease and bipolar disorder.

One diverse study has identified *MARK2* as an upstream regulator of PINK1, which raises a new perspective on the mitochondrial transport status of neurons^[24]. The motor proteins dynein (retrograde) and kinesin (anterograde) are responsible for mitochondria transport within a normal neuron. A molecular switch, PINK1, modulates the ability to carry mitochondria between dynein and kinesin. *MARK2* regulates the direction of neuronal mitochondrial movement by cleavage and binding/phosphorylation of PINK1^[25,26]. Apart from the important role of *MARK2* protein in the brain, especially in the pathophysiology of neurodevelopmental disorders, due to its serine-threonine kinase activity, *MARK2* protein may also play a role in the lithium metabolic pathway^[12]. It has been established that lithium activates the serine-threonine kinase Akt-1^[27]. Overexpression of *MARK2* has been shown to provoke PI3K/AKT/NF κ B signaling pathways and induces cell apoptosis^[28]. This evidence also supports the role of *MARK2* variants in the lithium's mechanism of action and the patients' various responses to the medications. Regarding the importance of *MARK2* gene in the pathophysiology of bipolar disorder and also in the lithium's mechanism of action, the association of one of these gene variants, rs10792421, was investigated with different responses to lithium treatment in bipolar type 1 patients.

MATERIALS AND METHODS

Study subjects

The sample size was assessed using the usual expectations of this type of study, which is 95% CI ($\alpha = 0.05$), 80% power ($\beta = 20\%$), and minimum extreme odds ratio to detect 2.0. According to data from the Iranome database (www.iranome.com), the hypothetical percentage of controls with exposure to risk alleles was set at 0.28. All patients were diagnosed by psychiatrists at Imam Hossein Hospital of Tehran and Razi Hospital of Urmia according to the Mental Illness Diagnosis and Statistics Manual, 4th Edition Text Revision (DSM-IV-TR) and DSM-5^[29]. Due to the naturalistic design of the study, patients receiving other pharmacological treatments and had stable doses of other treatments for at least four weeks prior to the start of lithium were included in the study. Various

patients' responses to lithium carbonate drugs were calculated using the Clinical Global Impression Severity (CGI-S) scale. Although the CGI-S is a common clinical scale for psychiatric disorders, it relies heavily on the clinical understanding of a patient's mental state by a psychiatrist and includes two subtypes. The first subclass ranges from 1 (no illness) to 7 (very serious illness) and the second subclass measures global improvement of patients from the beginning of their disorder. The severity of the illness was assessed by a psychiatrist through diagnosing the current state of the illness at the start of treatment. Patients were eventually divided into three groups: responders (R; n = 101), semi-responders (SR; n = 91), and non-responders (NR; n = 11). The information collected from all cases includes social demographic information (age, gender, ethnicity, marriage history, vocational status, and education); lithium carbonate consumption data (amount, side effects, and age of start); codependence: all other drugs, including alcohol and tobacco; current treatment and medication; medical history, including height and weight for calculating BMI; family history and drug-related problems. All patients were treated with lithium and followed for at least six months. Information on dose and drug consumption behavior, including results of blood lithium levels, routinely collected in the clinic as part of routine clinical care, was also collected.

Genotyping methods

Peripheral blood (4 ml) was collected from all patients in EDTA tubes. Salting out method was employed to extract DNA from samples. 4P-ARMS-PCR method was used for genotyping the *MARK2* gene, rs10792421, in Flex Thermo Cycler (Analytik Jena, Germany). Each amplification reaction contained 2× Master Mix Red (Ampliqon, Denmark), 5 pmol/l of inner primers, 10 pmol/l of outer primers, 150 ng of DNA, and 12.5 μ l of Taq DNA Polymerase. The PCR program included 35 cycles of initial denaturation at 95 °C for 5 minutes, 45 seconds at 95 °C (denaturation), 45 seconds at 55 °C (annealing), 45 seconds at 72 °C (elongation), and final extension at 72 °C for 5 minutes. The rs10792421 allele amplification produced 210, 280, and 441 bp bands for the A allele, G allele, and external primers, respectively. PCR products were further approved by random sequencing of about 15% of the genotypes using the ABI 3730 XL DNA Analyzer (Macrogen, South Korea).

Statistical analysis

The chi-square test was used to compare the genotypes of the patients to determine their suitability for the Hardy-Weinberg equilibrium. The association

between allele and genotype frequency and response to lithium therapy was investigated using the chi-square test in three different genetic models, including recessive, dominant, and co-dominant. The results of the association analysis are shown as odds ratios and their 95% CI. *p* value less than 0.05 was considered as statistically significant in all analyses.

RESULTS

The association between rs10792421 and patients' responses to lithium was investigated in 203 bipolar I patients through Pearson's chi-squared analysis. The obtained results proved a significant association of rs10792421 different genotypes with patients' responses to lithium in bipolar I patients. In R group, as shown in Table 1, the A allele was more frequent compared to NR group in codominant model of analysis (OR: 11.40; 95% CI: 1.28–101.37; *p* = 0.028). AA and AG genotypes were more frequent in R group compared to NR group in the dominant model of analysis (OR: 4.86; 95% CI: 1.35–17.55; *p* = 0.017). Based on Table 2, in SR, the A allele was more frequent in R group when compared with NR group in codominant model of analysis (OR: 9.33; 95% CI: 1.04–84.09; *p* = 0.042). AA and AG genotypes were more frequent in the SR group compared NR group in the dominant model of analysis (OR: 4.87; 95% CI: 1.33–17.75; *p* = 0.018).

DISCUSSION

MARK2 kinase is recruited in a variety of neuronal processes such as polarity, motility, neuronal migration, and also tau protein phosphorylation^[25,30], which is essential for cortical formation^[31]. The immature neurons' migration is considered as an essential step for cortical formation^[31]. Bipolar disorder has been identified as a neurodevelopmental disorder based on obtained results of cortical cell migration incompetence in patients^[32,33]. Abnormal neuronal migration can also be observed later in the life of patients with Alzheimer's disease^[17]. Tauopathy is recognized as one of the pathophysiological symptoms of Alzheimer's disease^[34]. In a 2013 study, Gu *et al.*^[13] found that *MARK2* enhances tau phosphorylation *in vitro* and reported certain interactions between *MARK2* and tau in the brain tissue of postmortem patients with Alzheimer's disease. The role of tauopathy in bipolar disorder has also been investigated. A research on cerebrospinal fluid in youth bipolar patients and a similar study in elderly patients and mild cognitive disorders detected no proof of

Table 1. Association analysis of rs10792421 with response to lithium carbonate in responder group (n=112)

Model of inheritance	Genotypes	NR number (%)	R number (%)	OR (95% CI)	p value
Codominant	G/G	6 (54.4)	20 (19.8)	1.00	
	A/G	4 (36.4)	43 (42.2)	3.23 (0.82-12.72)	0.028
	A/A	1 (9.1)	38 (37.6)	11.40 (1.28-101.37)	
Dominant	G/G	6 (54.5)	20 (19.8)	1.00	
	A/G-A/A	5 (45.5)	81 (80.2)	4.86 (1.35-17.55)	0.017
Recessive	G/G-A/G	10 (90.9)	63 (62.4)	1.00	
	A/A	1 (9.1)	38 (37.6)	6.03 (0.74-49.00)	0.038
Overdominant	G/G-A/A	7 (63.6)	58 (57.4)	1.00	
	A/G	4 (36.4)	43 (42.6)	1.30 (0.36-4.71)	0.69
Log-additive	---	---	---	3.32 (1.28-8.62)	0.0074

NR, non-responders; R, responders

of tauopathy^[35,36]. However, in another study, it has been demonstrated that the overall proportion of tau-phosphorylated reduced in bipolar patients who carry the high-risk allele variant of the previously studied *CACNA1C* gene^[35]. This reduction was not observed in healthy individuals carrying the same allele. Further studies are needed to study the association of tau phosphorylation by regulatory genes, such as the *MARK2* gene, with bipolar disease to confirm previous findings and attribute them to bipolar disorder^[36].

Lithium has various molecular targets, including GSK3-β inhibitor^[37]. There is conflicting evidence that GSK3-β ultimately inhibits or activates *MARK2*^[21,38]. As a result, it is not yet clear whether lithium therapy can reduce or increase tau phosphorylation in carriers of the common variant of the *MARK2* gene^[10]. Lithium has been found to regulate apoptosis through NF-kappa B and mitogen-activated protein kinase-dependent pathways^[39]. NF-kappa B is an important mediator and an essential transcription factor in inflammatory signaling and is involved in the relapse/remission course of bipolar disorder^[40,41]. One of the *MARK2*

gene variants, rs10792421, has been indicated to alter the NF-kappa B motif^[42]. It has also been reported that this variant may act in a similar way in both Alzheimer's disease and bipolar disorder. These results make this variant a potential marker in the pathophysiology of bipolar disorder and lithium's mechanism of action.

The current study investigated the association of rs10792421 of *MARK2* gene with the response to lithium medication in bipolar type 1 patients. Analyses showed that the AA genotype of this variant is associated with better response to lithium treatment in bipolar type 1 patients, as in comparative studies, AA genotype was more frequent than GG and AG + GG genotypes in the responder to lithium group, and also in the moderate responder to lithium group than the non-responder group. Therefore, the A allele of this variant can serve as a low-risk allele for treatment with the standard dose of lithium in bipolar patients.

Overall, it could be concluded that patients with AA genotype may benefit more from the standard dose of lithium than those with GG or AG genotypes.

Table 2. Association analysis of rs10792421 with response to lithium carbonate in semi-responders

Model of inheritance	Genotypes	NR number (%)	R number (%)	OR (95% CI)	p value
Codominant	G/G	6 (54.4)	18 (19.8)	1.00	
	A/G	4 (36.4)	45 (49.5)	3.75 (0.95-14.88)	0.042
	A/A	1 (9.1)	28 (30.8)	9.33 (1.04-17.75)	
Dominant	G/G	6 (54.5)	18 (19.8)	1.00	
	A/G-A/A	5 (45.5)	73 (80.2)	4.87 (1.33-17.75)	0.018
Recessive	G/G-A/G	10 (90.9)	63 (69.2)	1.00	
	A/A	1 (9.1)	28 (30.8)	4.44 (0.54-36.41)	0.098
Overdominant	G/G-A/A	7 (63.6)	46 (50.5)	1.00	
	A/G	4 (36.4)	45 (49.5)	1.71 (0.47-6.25)	0.41
Log-additive	---	---	---	3.30 (1.21-9.02)	0.012

NR, non-responders; R, responders

Moreover, patients with GG genotype of rs1079241 have more challenges with the routine treatment of lithium and are required to be carefully monitored during their treatment to avoid harmful side effects, suicide, or relapse of disorder.

DECLARATIONS

Ethical statement

The above-mentioned sampling protocols were approved by the Institutional Review Board of Shahid Beheshti Medical University, Tehran, Iran (ethical code: IR.SBMU.MSP.REC.1398.344). All participants signed consents in accordance with the Declaration of the World Medical Association in Helsinki.

Data availability

The analyzed data sets generated during the study are available from the corresponding author on reasonable request.

Author contributions

SSAA and MDO made substantial contributions to the conception of the study and supervised the study. AA performed the laboratory assessment. JS, SH, and BB collected samples and analyzed the data. AO wrote the manuscript. All authors have approved the final manuscript and have agreed both to be personally accountable for their own contributions and to ensure that questions related to the accuracy or integrity of any part of the work are appropriately investigated, resolved, and the resolution documented in the literature.

Conflict of interest

None declared.

Funding/support

This study was financially supported by Shahid Beheshti University of Medical Sciences, Tehran, Iran.

REFERENCES

1. Cade J F J. Lithium salts in the treatment of psychotic excitement. *The medical journal of Australia* 1949; **2**(10): 349-s2.
2. Nivoli AM, Colom F, Murru A, Pacchiarotti I, Castro-Loli P, González-Pinto A, Fountoulakis KN, Vieta E. New treatment guidelines for acute bipolar depression: a systematic review. *Journal of affective disorders* 2011; **129**(1-3): 14-26.
3. Moreira J, Noé G, Rangarajan S, Courtin C, Etain B, Geoffroy PA, Laplanche JL, Vidal M, Bellivier F, Marie-Claire C. Lithium effects on serine-threonine kinases activity: High throughput kinomic profiling of lymphoblastoid cell lines from excellent-responders and non-responders bipolar patients. *The world journal of biological psychiatry* 2020; **21**(4): 317-324.
4. Suppes T, Baldessarini RJ, Faedda GL, Tohen. Risk of recurrence following discontinuation of lithium treatment in bipolar disorder. *Archives of general psychiatry* 1991; **48**(12): 1082-1088.
5. Goodwin GM. Recurrence of mania after lithium withdrawal. Implications for the use of lithium in the treatment of bipolar affective disorder. *The British journal of psychiatry* 1994; **164**(2): 149-152.
6. Jawad I, Watson S, Haddad PM, Talbot PS, McAllister-Williams RH. Medication nonadherence in bipolar disorder: a narrative review. *The therapeutic advances in psychopharmacology* 2018; **8**(12): 349-363.
7. Ahmed S, Zhou Z, Zhou J, Chen SQ. Pharmacogenomics of Drug Metabolizing Enzymes and Transporters: Relevance to Precision Medicine. *Genomics, proteomics and bioinformatics* 2016; **14**(5): 298-313.
8. Machado-Vieira R, Manji HK, Zarate CA Jr. The role of lithium in the treatment of bipolar disorder: convergent evidence for neurotrophic effects as a unifying hypothesis. *Bipolar disorders* 2009; **2**(Suppl 2): 92-109.
9. Forlenza OV, De-Paula VJ, Diniz BS. Neuroprotective effects of lithium: implications for the treatment of Alzheimer's disease and related neurodegenerative disorders. *ACS chemical neuroscience* 2014; **5**(6): 443-450.
10. Drange OK, Smeland OB, Shadrin AA, Finseth PI, Witoelar A, Frei O. Psychiatric Genomics Consortium Bipolar Disorder Working Group, Wang Y, Hassani S, Djurovic S, Dale AM, Andreassen OA. Genetic overlap between Alzheimer's disease and bipolar disorder implicates the MARK2 and VAC14 genes. *Frontiers in neuroscience* 2019; **13**: 220.
11. Pasquali L, Busceti C L, Fulceri F, Paparelli A, Fornai F. Intracellular pathways underlying the effects of lithium. *Behavioural pharmacology* 2010; **21**(5-6): 473-492.
12. Chalecka-Franaszek E, Chuang DM. Lithium activates the serine/threonine kinase Akt-1 and suppresses glutamate-induced inhibition of Akt-1 activity in neurons. *Proceedings of the national academy of sciences of the United States of America* 1999; **96**(15): 8745-8750.
13. Gu GJ, Lund H, Wu D, Blokzijl A, Classon C, von Euler G, Landegren U, Sunnemark D, Kamali-Moghaddam M. Role of individual MARK isoforms in phosphorylation of tau at Ser²⁶² in Alzheimer's disease. *Neuromolecular medicine* 2013; **15**(3): 458-469.
14. Biernat J, Wu YZ, Timm T, Zheng-Fischhofer Q, Mandelkow E, Meijer L, Mandelkow EM. Protein kinase MARK/PAR-1 is required for neurite outgrowth and establishment of neuronal polarity. *Molecular biology of the cell* 2020; **13**: 4013-4028.
15. Martino DJ, Samamé C, Ibañez A, Strejilevich SA. Neurocognitive functioning in the premorbid stage and in the first episode of bipolar disorder: a systematic review. *Psychiatry research* 2015; **226**(1): 23-30.
16. Gold CA, Budson AE. Memory loss in Alzheimer's

disease: implications for development of therapeutics. *Expert review of neurotherapeutics* 2008; **8**(12):1879-91.

17. Godefroy O, Bakchine S, Verny M, Delabrousse-Mayoux JP, Roussel M, Pere JJ, REFLEX study group. Characteristics of Alzheimer's disease patients with severe executive disorders. *Journal of Alzheimers disease* 2016; **51**(3): 815-825.
18. Matenia D, Mandelkow E M. The tau of MARK: a polarized view of the cytoskeleton. *Trends in biochemical sciences* 2009; **34**(7): 332-342.
19. Zhang X, Heng X, Li T, Li L, Yang D, Zhang X, Du Y, Doody RS, Le W. Long-term treatment with lithium alleviates memory deficits and reduces amyloid-beta production in an aged Alzheimer's disease transgenic mouse model. *Journal of Alzheimers disease* 2011; **24**(4): 739-749.
20. Matsunaga S, Kishi T, Annas P, Basun H, Hampel H, Iwata N. Lithium as a treatment for Alzheimer's disease: A systematic review and meta-analysis. *Journal of Alzheimers disease* 2010; **48**(2): 403-410.
21. Kosuga S, Tashiro E, Kajioka T, Ueki M, Shimizu Y, Imoto M. GSK-3beta directly phosphorylates and activates MARK2/PAR-1. *The Journal of biological chemistry* 2005; **280**(52): 42715-42722.
22. Kaidanovich-Beilin O, Woodgett JR. GSK-3: Functional Insights from Cell Biology and Animal Models. *Frontiers in molecular neuroscience* 2011; **4**: 40.
23. Hanger DP, Noble W. Functional implications of glycogen synthase kinase-3-mediated tau phosphorylation. *International journal of Alzheimers disease* 2011; **2011**: 352805.
24. Matenia D, Hempp C, Timm T, Eikhof A, Mandelkow EM. Microtubule affinity-regulating kinase 2 (MARK2) turns on phosphatase and tensin homolog (PTEN)-induced kinase 1 (PINK1) at Thr-313, a mutation site in Parkinson disease: effects on mitochondrial transport. *The journal of biological chemistry* 2012; **287**(11): 8174-8186.
25. Matenia D, Mandelkow EM. Emerging modes of PINK1 signaling: another task for MARK2. *Frontiers in molecular neuroscience* 2014; **7**: 37.
26. Weihofen A, Thomas KJ, Ostaszewski BL, Cookson MR, Selkoe DJ. Pink1 forms a multiprotein complex with Miro and Milton, linking Pink1 function to mitochondrial trafficking. *Biochemistry* 2009; **48**(9): 2045-2052.
27. Reiner O, Shmueli A, Sapir T. Neuronal migration and neurodegeneration: 2 Sides of the same coin. *Cerebral cortex* 2009; **19**: 42-48.
28. Wei X, Xu L, Jeddo SF, Li K, Li X, Li J. MARK2 enhances cisplatin resistance via PI3K/AKT/NF- κ B signaling pathway in osteosarcoma cells. *American journal of translational research* 2020; **12**(5):1807-1823.
29. Regier DA, Kuhl EA, Kupfer DJ. The DSM-5: classification and criteria changes. *World psychiatry* 2013; **12**(2): 92-98.
30. Insolera R, Chen S, Shi SH. Par proteins and neuronal polarity. *Developmental neurobiology* 2011; **71**(6): 483-494.
31. Kon E, Cossard A, Jossin Y. Neuronal polarity in the embryonic mammalian cerebral cortex. *Frontiers in cellular neuroscience* 2017; **11**: 163.
32. Sanches M, Keshavan MS, Brambilla P, Soares JC. Neurodevelopmental basis of bipolar disorder: A critical appraisal. *Progress in neuro-psychopharmacology and biological psychiatry* 2008; **32**(7): 1617-1627.
33. O'Shea KS, McInnis MG. Neurodevelopmental origins of bipolar disorder: iPSC models. *Molecular and cellular neurosciences* 2016; **73**: 63-83.
34. Jack CR Jr, Knopman D S, Jagust W J, Petersen RC, Weiner MW, Aisen PS, Shaw LM, Vemuri P, Wiste H J, Weigand SD, Lesnick TG, Pankratz VS, Donohue MC, Trojanowski JQ. Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *The lancet neurology* 2013; **12**(2): 207-216.
35. Forlenza OV, Aprahamian I, Radanovic M, Talib LL, Camargo MZ, Stella F, Machado-Vieira R, Gattaz WF. Cognitive impairment in late-life bipolar disorder is not associated with Alzheimer's disease pathological signature in the cerebrospinal fluid. *Bipolar disorders* 2016; **18**(1): 63-70.
36. Jakobsson J, Pålsson E, Sellgren C, Rydberg F, Ekman A, Zetterberg H, Blennow K, Landén M. CACNA1C polymorphism and altered phosphorylation of tau in bipolar disorder. *The British journal of psychiatry* 2016; **208**(2): 195-196.
37. Freland L, Beaulieu JM. Inhibition of GSK3 by lithium, from single molecules to signaling networks. *Frontiers in molecular neuroscience* 2012; **5**: 14.
38. Timm T, Balusamy K, Li X, Biernat J, Mandelkow E, Mandelkow E M. Glycogen Synthase Kinase (GSK) 3 β directly phosphorylates serine 212 in the regulatory loop and inhibits microtubule affinity-regulating kinase (MARK) 2. *The journal of biological chemistry* 2008; **2839**(27): 18873-18882.
39. Németh ZH, Deitch EA, Szabó C, Fekete Z, Hauser CJ, Haskó G. Lithium induces NF- κ B activation and interleukin-8 production in human intestinal epithelial cells. *The journal of biological chemistry* 2002; **277**(10): 7713-7719.
40. Miklowitz DJ, Portnoff LC, Armstrong CC, Keenan-Miller D., Breen EC, Muscatell K A, Eisenberger N I, Irwin MR. Inflammatory cytokines and nuclear factor- κ B activation in adolescents with bipolar and major depressive disorders. *Psychiatry research* 2016; **241**: 315-322.
41. Slavich GM, Irwin MR. From stress to inflammation and major depressive disorder: a social signal transduction theory of depression. *Psychological bulletin* 2014; **140**(3): 774-815.
42. Ward LD, Kellis M. HaploReg: a resource for exploring chromatin states, conservation, and regulatory motif alterations within sets of genetically linked variants. *Nucleic acids research* 2012; **40**: 930-934.