

Bronchodilatory Activity of *Vitis vinifera* Leaf Hydroalcoholic Extract in Rat

Mohammad Kazem Gharib Naseri^{*1} and Akbar Heidari²

¹Dept. of Physiology and ²School of Pharmacy, Jondi Shapour Ahwaz University of Medical Sciences, Ahwaz, Iran

Received 13 July 2005; revised 8 January 2006; accepted 14 January 2006

ABSTRACT

Several reports have shown the various effects of grape (*Vitis vinifera*) seed extract such as antioxidant, hypotensive, hypolipidemic and vasodilatory effects. The aim of present study was to investigate the effect of grape leaf hydroalcoholic extract on isolated rat tracheal contractions induced by KCl and acetylcholine. The trachea was removed from male adult Sprague-Dawley rat and placed in an organ bath containing Krebs-Henseleit solution and contractions were recorded isometrically. The results demonstrate that the grape leaf extract at 0.5, 1, 2, 4 and 8 mg/ml significantly reduces the tracheal contractions induced by KCl (60 mM) dose-dependently ($P < 0.0001$). Acetylcholine (55 μ M)-induced tracheal contractions were also attenuated at the same concentration of the extract ($P < 0.0001$). The grape leaf extract induced relaxation in the KCl-induced contraction in trachea was unaffected neither by nitric oxide (NO) synthase inhibitor (L-NAME, 100 μ M) nor by beta-adrenoceptor antagonist (propranolol 1 μ M). Our results suggest that the bronchodilatory effect of grape leaf extract is mediated via the voltage dependent calcium channels. Furthermore, the beta-adrenergic and NO are not involved. *Iran. Biomed. J. 10 (2): 79-83, 2006*

Keywords: Grape leaf, Bronchodilatation, Rat

INTRODUCTION

Grape (*Vitis vinifera*) from Vitaceae is a perennial woody vine native to Asia Minor and then introduced in Europe [1]. Its leaves are consumed in some traditional foods (Dolmathes) and used for diarrhea, vomiting and varicose treatment [2]. The pharmacological properties of grape seeds are well investigated and it is believed that flavonoids (from polyphenols) are its most potent constituents. Grape seed extract reduces blood lipids in rabbit [3] with no side effects after long term consumption [4]. The procyanidins in grape seed induces endothelial-dependent vasorelaxation in human artery [5]. This effect is carried out through nitric oxide (NO) and enhancement of cGMP [6,7]. Procyanidins of grape seed extract show several effects, for instance, protective effect against oxidative stress [8], cataract [9], colon cancer [10] and the increase in plasma antioxidant activity [11]. Recently, the vasorelaxatory effect of grape leaf extract on isolated rat aorta was shown to

be dependent on endothelium integrity, and production of NO and cGMP [12]. The aim of the present study was to investigate the effect of grape leaf hydroalcoholic extract (GLHE) on rat isolated trachea.

MATERIALS AND METHODS

Acetylcholine, propranolol and L-NAME were purchased from Sigma (USA) and other chemicals were from Merck (Germany). All chemicals were dissolved in the Krebs-Henseleit solution. Grape leaves were collected from the Jondi Shapour Ahwaz University of Medical Sciences Campus (Ahwaz, Iran) in April 2004. The collected leaves were identified by Dr. Siahpoosh from Department of Pharmacogenosy, Faculty of Pharmacy, Jondi Shapour Ahwaz University of Medical Sciences. A voucher specimen was deposited in the herbarium of the same department under number A06390001M. Grape leaves were dried under

*Corresponding Author; Fax: (+98-611) 333 2036; E-mail: gharibnaseri_m@yahoo.com

shade and powdered by electrical grinder. The powder (50 g) was mixed with 230 ml of alcohol (70%) for 72 h at room temperature and stirred four times daily. The mixture was filtered with Whatman filter paper (No.1) and then the solvent was evaporated at room temperature. The obtained extract powder was 9.5 g (19% extraction ratio) which stored at 4°C until being used.

Animals and tissue preparation. Male Sprague-Dawley rats (200-245 g) were housed in cages with ambient temperature at 20-24°C, 12-h light/dark cycle and free access to water and food. All animal experiments were approved by the Local Animal Ethics Committee of Ahwaz Jondi Shapour University of Medical Sciences. The rats were anaesthetized by ketamine hydrochloride (50 mg/kg, ip) and chest and neck were opened. The trachea (2 cm) was dissected out and its connective tissue was removed in a cold oxygenated Krebs-Henseleit solution. A piece of trachea (5 mm) with 5-6 cartilage rings was cut and mounted between two stainless steel hooks horizontally. The lower hook was fixed at the bottom of the organ bath and upper one was connected to an isometric transducer (UF1 Harvard transducer, UK) connected to an ink-writing curvilinear recorder (Harvard Universal Oscillograph, UK). The organ bath contained Krebs-Henseleit (10 ml, pH 7.4 and at 37°C) with following composition (mM): NaCl, 118; KCl, 4.7; CaCl₂, 2.52; MgSO₄, 1.64; KH₂PO₄, 1.18; NaHCO₃, 7 and glucose, 5.5 bubbled with oxygen. The initial tension was 1.5 g throughout the experiment and equilibrium period was 60 min in which, the bath solution was refreshed every 15 min. After equilibrium period, the trachea was contracted by 60 mM of KCl [13] or by 55 μ M [14] of acetylcholine and when the plateau was achieved, the extract (0.5, 1, 2, 4 and 8 mg/ml) was added non-cumulatively to the organ bath and left to achieve a new plateau. Then, during 10 min, the bath solution was exchanged three times with fresh Krebs-Henseleit solution. The same protocol was repeated but with the higher extract concentration. To investigate the involvement of NO and β -adrenoceptors, first, the inhibitory effect of extract (3 mg/ml) was recorded on KCl-induced contraction. After 15 min and several exchanging the bath solution, the same protocol was repeated in the presence of L-NAME (100 μ M, 10 min) or propranolol (1 μ M, 10 min). At the end of the

experiments, the tissue water was absorbed by filter paper and tissue weighed.

Statistical analysis. Data are expressed as mean \pm SEM of g force/100 mg tissue or in percentage of force changes. Statistical comparisons were made by Student's *t*-test and one-way ANOVA. *P*<0.05 was considered significant.

RESULTS

Effect of GLHE on KCl and acetylcholine-induced tracheal contractions (ACh). Figure 1 shows that non-cumulative concentrations of GLHE (0.5, 1, 2 and 4 mg/ml) have reduced the KCl and ACh-induced tracheal contractions in a dose-dependent manner and significantly (*n* = 8, *P*<0.0001, tested by ANOVA). Each piece of trachea was used only for one of these two spasmogens.

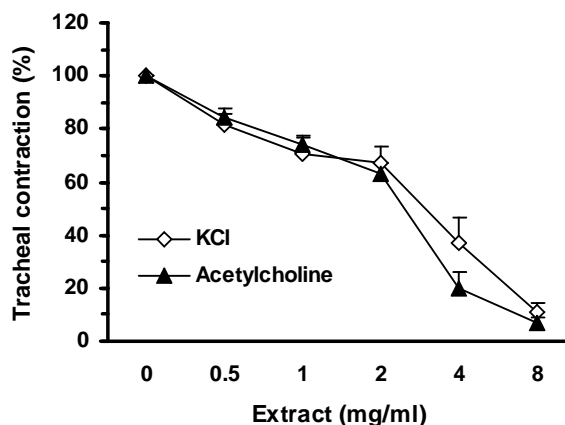


Fig. 1. Effect of non-cumulative concentrations of the grape leaf hydroalcoholic extract on rat tracheal contractions induced by KCl (60 mM) or ACh (55 μ M). Spasmogens-induced contractions in the absence of extract were considered as 100% (*n* = 8). The *P* values of dose-response for the both curves (ANOVA) are <0.0001.

Inhibitory effect of extract on trachea KCl-induced contraction in the presence of L-NAME. Figure 2 shows that the inhibitory effects of GLHE (3 mg/ml) on the KCl-induced contraction are not different in the absence and in the presence of L-NAME [15] as a NO synthase inhibitor (100 μ M, for 10 min, *n* = 8).

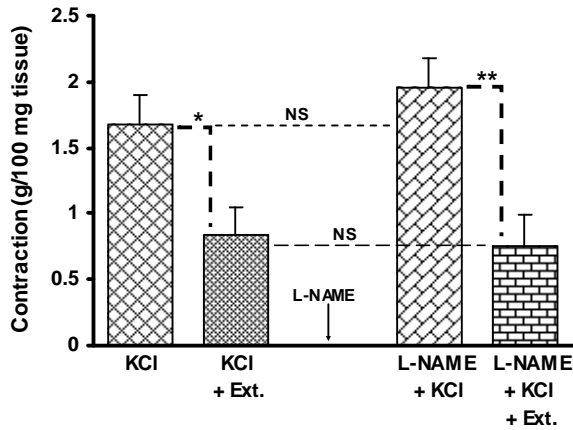


Fig. 2. The comparison of spasmolytic effect of grape leaf hydroalcoholic extract (Ext. 3 mg/ml) on contraction induced by KCl (60 mM) in absence and in the presence (10 min) of the nitric oxide synthase inhibitor (L-NAME, 100 μ M) in rat trachea ($n = 8$; *, $P < 0.05$; **, $P < 0.0001$ and NS, non-significant).

Inhibitory effect of extract on trachea KCl-induced contraction in the presence of propranolol. As Figure 3 shows, the inhibitory effects of GLHE (3 mg/ml) on the KCl-induced contraction are not different in the absence and in the presence of propranolol [16] as a β -adrenoceptor antagonist (1 μ M, for 10 min, $n = 8$).

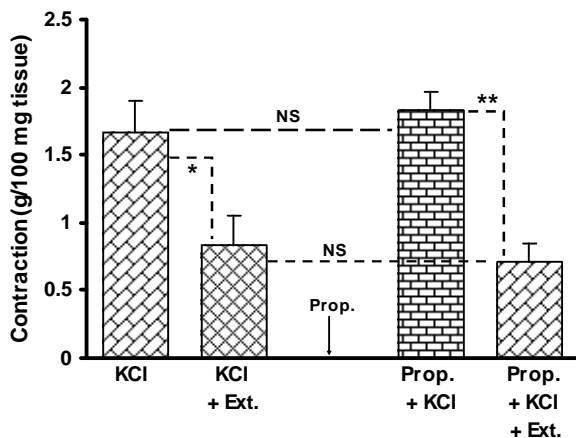


Fig. 3. Spasmolytic effect of grape leaf hydroalcoholic extract (Ext. = 3 mg/ml) on contraction induced by KCl (60 mM) in absence and in the presence (10 min) of β -adrenoceptor antagonist, propranolol, (Prop. = 1 μ M) in rat trachea ($n = 8$, *, $P < 0.05$; **, $P < 0.0001$ and NS = non significant).

Study of the cholinergic antagonistic effect of extract. The extract inhibited the contraction induced by acetylcholine to study whether this inhibitory effect is due to antagonistic effect on acetylcholine receptors or is just due to inhibiting

the contracting effect of acetylcholine, the following protocol was carried out. The trachea was contracted by KCl (30 mM) and after achievement of the plateau, ACh (27 μ M) was added to organ bath. The ACh-induced contraction was superimposed on KCl-induced contraction. The tissue was washed several times and was left for 20 min. In the next stage, atropine was first added to bath for 10 min and then the previous protocol was repeated. Figure 4 shows that only KCl induces contraction but not ACh. Meanwhile, applying the extract (4 mg/ml) abolished this contraction.

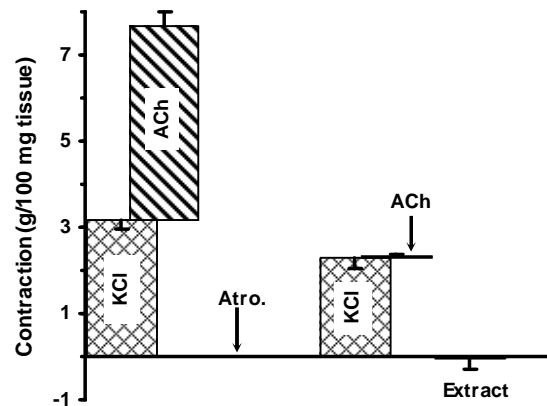


Fig. 4. Effect of atropine (Atro. = 30 μ M) on rat trachea superimposed contractions induced by KCl (30 mM) and acetylcholine (ACh = 27 μ M). Atropine has only prevented the contractile effect of acetylcholine but not the KCl effect. Extract (4 mg/ml) has abolished the contractile effect of KCl ($n = 7$).

DISCUSSION

The present study showed that GLHE reduces the tracheal contractions induced by KCl and acetylcholine. In this bronchodilating effect, neither NO nor β -adrenoceptors were involved. This spasmolytic effect was reversible due to the fact that, this effect was abolished when the bath solution was exchanged with fresh solution. Therefore, it seems that extract effects occur on the smooth muscle cell membrane. The depolarization of smooth muscle cells by KCl causes the activation of voltage dependent calcium channels (VDCC), which ultimately induces the contraction [16-18]. The existence of L-type calcium channels in the rat trachea has been reported [18] and supported by the inhibitory effect of verapamil on rat trachea contraction induced by KCl [19]. Furthermore, it is reported that the same extract reduces the rat aorta contracted by KCl [12] which support that, the

VDCC are involved in the spasmolytic effect of GLHE. In the vasorelaxatory effect of the extract, NO and cGMP was involved but in trachea the involvement of NO was not the case, since the spasmolytic effect of extract was unaffected by L-NAME. However, It has been reported that, the rat trachea produces NO and cGMP without NO synthase activation [20]. Therefore, it was possible that grape leaf flavonoids, for instance, quercetin [21] promotes NO production and induces the tracheal relaxation but without NO synthase activation. The inhibitory effect of quercetin on uterus and intestine contractility has been also reported [22, 23]. The β -adrenoceptors activation by isoproterenol causes tracheal relaxation [24] but the ineffectiveness of propranolol to reduce the extract spasmolytic activity indicates that, these receptors are not involved. Acetylcholine depolarizes the tracheal smooth muscle via increasing intracellular calcium concentration [25]. Inhibitory effect of extract on ACh-induced tracheal contraction, could be an evidence for anticholinergic property of extract. In the presence of atropine, KCl induced contraction, however, ACh was unable to induce more contraction. Meanwhile, adding extract abolished the contraction induced by KCl. This result supports that, extract reduces tracheal contractility, possibly, through VDCC and the cholinergic receptors are not involved directly in this bronchodilatory effect of extract. On the other hand, it has been reported that atropine reverses the ACh-induced relaxation in rat aorta precontracted by phenylephrine but the vasorelaxatory effect of the same extract was unaffected by atropine [12]. This report supports that GLHE reduces the ACh-induced tracheal contraction via blocking the events occurred by coupling ACh to its receptors rather than blocking cholinergic receptors.

In conclusion, these data suggest that the GLHE inhibits the rat tracheal contractions induced by KCl and ACh via blocking the VDCC on the smooth muscle cells membrane.

ACKNOWLEDGMENTS

The authors wish to thank the Jondi Shapour Ahwaz University of Medical Sciences (Ahwaz, Iran) for supporting this work financially.

REFERENCES

- Bombardelli, E. and Morazzoni, P. (1995) *Vitis vinifera* L. *Fitoterapia* 66 (4): 291-317.
- Zargari, A. (1993) Medicinal plants. Tehran University Publications, Tehran, Iran.
- Yu, H., Zhao, X., Xu, G., and Wang, S.E. (2003) Effect of grape seed extracts on blood lipids in rabbits model with hyperlipidemia. *Wei Sheng Yan Jiu* 31 (2): 114-116.
- Rays, S., Bagchi, D., Lim, P.M., Bagchi, M., Gross, S.M., Kothari, S.C., Preuss, H.G., and Stohs, S.J. (2001) Acute and long-term safety evaluation of a novel IH636 grape seed proanthocyanidin extract. *Res. Commun. Mol. Pathol. Pharmacol.* 109 (3-4): 165-197.
- Aldini, G., Carini, M., Piccoli, A., Rossoni, G. and Maffei Facino, R. (2003) Procyanidins from grape seeds protect endothelial cells from peroxynitrite damage and enhance endothelium-dependent relaxation in human artery: new evidences for cardio-protection. *Life Sci.* 73 (22): 2883-2898.
- Fitzpatrick, D.F., Bing, B., Maggi, D.A., Fleming, R.C. and O'Malley, R.M. (2002) Vasodilating procyanidins derived from grape seed. *Ann. N.Y. Acad. Sci.* 957: 78-89.
- Kim, S.H., Kang, K.W., Kim, K.W. and Kim, N.D. (2000) Procyanidins in crataegus extract evoke endothelium-dependent vasorelaxation in rat aorta. *Life Sci.* 67 (2): 121-131.
- Sato, M., Maulik, G., Ray, P.S., Bagchi, D. and Das, D.K. (1999) Cardioprotective effects of grape seed proanthocyanidins against ischemic reperfusion injury. *J. Mol. Cell Cardiol.* 31(6): 1289-1297.
- Yamakoshi, J., Saito, M., Kataoka, S. and Tokutake, S. (2002) Procyanidin-rich extract from grape seeds prevents cataract formation in hereditary cataractous (ICR/f) rats. *J. Agric. Food Chem.* 50 (17): 4983-4988.
- Singletary, K.W. and Meline, B. (2001) Effect of grape seed proanthocyanidins on colon aberrant crypts and breast tumors in a rat dual-organ tumor model. *Nutr. Cancer* 39 (2): 252-258.
- Koga, T., Moro, K., Nakamori, K., Yamakoshi, J., Hosoyama, H., Kataoka, S. and Arigo, T. (1999) Increase of antioxidative potential of rat plasma by oral administration of proanthocyanidin-rich extract from grape seeds. *J. Agric. Food Chem.* 47 (5): 1892-1897.
- Gharib Naseri, M.K., Navid Hamidi, M. and Heidari, A. (2005) Vasorelaxant effects of *Vitis vinifera* leaf extract on isolated rat aorta. *Iran. J. Pharmaceut. Res.* 2: 93-99.
- Teixeira, M.C.L., Coelho, R.R., Leal-Cardoso, J.H. and Criddle, D.N. (2000) Comparative effects of niflumic acid and nefipine on 5-hydroxytryptamine- and acetylcholine-induced contraction of the rat trachea. *Eur. J. Pharmacol* 394: 117-122.

14. Agbonon, A., Eklugadegbeku, K., Aklikokou, K., Essien, K., Akpagana, K. and Gbeassor, M. (2002) The effect of *Mangifera indica* stem bark and *Pluchea ovalis* roots on tracheal smooth muscle *in vitro*. *Fitoterapia* 73: 619-622.
15. Chiu, C.C., Wu, R.J., Lee, C.H., Liou, S.F., Dai, Z.K., Chen, I.J. and Yeh, J.L. (2004) Anti-hypertension effect vanylidilol: a phenylaldehyde alpha/beta-adrenoceptor blocker with endothelium-dependent and K⁺ channels opening-associated vasorelaxant activities. *Pharmacology* 70 (3): 140-151.
16. Liao, J.F., Shi, C.C., Chen, S.Y., Fu, Y.T. and Chen, C.F. (1997) Spasmolytic effect of water extract of *Stemona radix* on the guinea-pig tracheal smooth muscle *in vitro*. *J. Ethnopharmacol.* 57 (1): 57-62.
17. Leal, L.K., Nechio, M., Silveira, E.R., Canuto, K.M., Fontenele, H.B., Ribeiro, R.A. and Viana, G.S. (2003) Anti-inflammatory and smooth muscle relaxant activities of the hydroalcoholic extract and chemical constituents from *Amburana cearensis* A C Smith. *Phytother. Res.* 17 (4): 335-340.
18. Bova, S., Cavalli, M., Cima, L., Luciani, S., Saponara, S., Sgaragli, G., Cargnelli, G. and Fusi, F. (2003) Relaxant and Ca²⁺ channel blocking properties of norbormide on rat non-vascular smooth muscles. *Eur. J. Pharmacol.* 470 (3): 185-191.
19. Chang, K.C., Ko, H.J., Cho, S.D., Yoon, Y.J. and Kim, J.H. (1993) Pharmacological characterization of effects of verapamil and GS 283 on isolated guinea pig and rat trachealis. *Eur. J. Pharmacol.* 236 (1): 51-60.
20. Jia, Y., Zacour, M., Tolloczko, B. and Martin, J.G. (1998) Nitric oxide synthesis by tracheal smooth muscle cells by a nitric oxide synthase-independent pathway. *Am. J. Physiol.* 275: L895-L901.
21. Diaz Lanza, A.M., Elias, R., Maillard, C., Faure, R., De Sotto, M. and Balansard, G. (1989) Flavonoids of 3 cultivars vine leaves, *Vitis vinifera* L. var. tinctoria (Alicante, Carignan, Grand nori). Value in chemical control. *Ann. Pharm. Fr.* 47 (4): 229-234.
22. Zhang, W.J., Chen, B.T., Wang, C.Y., Zhu, Q.H. and MO, Z.X. (2003) Mechanism of quercetin as an antidiarrheal agent. *Di Yi Jun Yi Da Xue Xue Bao* 23(10): 1029-1031.
23. Revuelta, M.P., Hidalgo, A. and Cantabrana, B. (1999) Involvement of cAMP and beta-adrenoceptors in the relaxing effect elicited by flavonoids on rat uterine smooth muscle. *J. Auton. Pharmacol.* 19 (6): 353-358.
24. McGrogan, I., Lu, S., Hipworth, S., Sormaz, L., Eng, R., Preocanin, D. and Daniel, E.E. (1995) Mechanisms of cyclic nucleotide-induced relaxation in canine tracheal smooth muscle. *Am. J. Physiol.* 268 (3 Pt 1): L407-L413.
25. Roux, E., Hyvelin, J.M., Savineau, J.P. and Marthan, R. (1998) Calcium signaling in airway smooth muscle cells is altered by *in vitro* exposure to the aldehyde acrolein. *Am. J. Respir. Cell Mol. Biol.* 19: 437-444.