

Recombinant Production of a Novel Fusion Protein: Listeriolysin O Fragment Fused to S1 Subunit of Pertussis Toxin

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Received 3 June 2019; accepted 6 February 2020; published online 10 February 2020

ABSTRACT

Background: Some resources have suggested that genetically inactivated PTs bear a more protective effect than chemically inactivated products. This study aimed to produce new version of PT, by cloning an inactive PTS1 in a fusion form with N-terminal half of the LLO pore-forming toxin. **Methods:** Deposited pdb structure file of the PT was used to model an extra disulfide bond. Codon-optimized ORF of the PTS1 was used to make recombinant constructs of PTS1 and LLO-PTS1 in the pPSG-IBA35 vector. The recombinant PTS1 and LLO-PTS1 proteins were expressed in *BL21 DE3* and *SHuffle T7* strains of *E. coli* and purified by affinity chromatography. Cytotoxic effects of the recombinant proteins were examined in the MCF-7 cell line. **Results:** The purity of the products proved to be more than 85%, and the efficiency of the disulfide bond formation in *SHuffle T7* strain was higher than *BL21 DE3* strain. No cytotoxicity of the recombinant proteins was observed in MCF-7 cells. Soluble recombinant PTS1 and LLO-PTS1 proteins were produced in *SHuffle T7* strain of *E. coli* with high efficiency of disulfide bonds formation. **Conclusion:** The LLO-PTS1 with corrected disulfide bonds was successfully expressed in *E. coli SHuffle T7* strain. Due to the safety for human cells, this chimeric molecule can be an option to prevent pertussis disease if its immunostimulatory effects would be confirmed in the future. **DOI: 10.29252/ibj.25.1.33**

Keywords: Adjuvant, Cloning, Fusion protein, Pertussis toxin

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INTRODUCTION

Pertussis is a highly contagious respiratory illness caused by *Bordetella pertussis* bacteria^[1]. This disease has been controlled utilizing two types of vaccines: the wP and aP ones^[2]. The aP vaccines are subunit vaccines that contain pure and inactive components of *B. pertussis* cells^[3,4], including detoxified pertussis toxin, filamentous hemagglutinin, pertactin, fimbrial antigens and so on. However, the protectivity of the aP vaccines has not been adequate

so that several outbreaks has been reported from different countries since 2012^[5,6]. The causes of the weakness of aP vaccines are yet unclear because it is not known whether the re-emergence of pertussis is due to the vaccine waning immunity or to the fundamental differences in the nature of the immune response caused by aP vaccines compared with the wP vaccines or natural infection^[7]. After the presentation of toxin-mediated theory about pertussis disease by Margaret Pittman in 1978^[8], it was suggested that a suitable inactivated and immunogenic PT is necessary

List of Abbreviations:

aP, acellular pertussis vaccine; **IPTG**, isopropyl- β -D-1-thiogalactopyranoside; **LLO**, listeriolysin O; **PT**, pertussis toxoid; **PTS1**, pertussis toxin S1 subunit; **SOEing**, synthesis by overlap extension; **wP**, whole-cell vaccine

and sufficient for a pertussis vaccine^[9]. Moreover, a former study reported that fimbria, similar to pertactin, is unnecessary for vaccine-induced immunostimulation^[10]. PT is composed of two subunits: A subunit with ADP-ribosyltransferase activity (S1 subunit), which is the main antigen for protection, and B oligomer, which is responsible for the toxin attachment to the target cells and is an important protective antigen in all available pertussis vaccines^[11].

Currently, the toxoid components in aP vaccines are chemically inactivated by aldehyde agents. However, more immunogenic PTs are inactivated genetically and are expected to have a longer protective effect than current products, because of the better preservation of the antigenic epitopes^[9]. The use of adjuvants in vaccine formulation is also important. Researchers believe that adjuvants can play a critical role in improving immunogenicity in next-generation aP vaccines^[12,13]. LLO molecule has been the subject of many research studies, and its role as an adjuvant against several infection and cancer models has been clearly illustrated. In fact, it has been identified that LLO molecule can act as an adjuvant either in chimeric form or as a separately expressed molecule^[14-18].

In this study, a new mutant of PTS1 carrying an extra disulfide bond was designed. This chimeric molecule (LLO-PTS1) was then expressed in *BL21 DE3* and *SHuffle T7* strains of *E. coli* and characterized further.

MATERIALS AND METHODS

The *E. coli* DH5 α and *BL21 DE3* strains were procured from the Pasteur Institute of Iran (Tehran).

SHuffle®T7 competent cells were prepared from New England Biolabs Inc., USA. DpnI restriction enzyme was obtained from Thermo Fisher Scientific (USA), and other chemicals were provided by molecular biology grade providers (Sigma and Merck Co., Germany).

Disulfide bond prediction

1PRT.pdb file, MODIP software of the National Centre for Biological Sciences Integrated Web ServerV1.0.1^[19], and Disulfide By Design software version 2.12^[20] were used to model, generate files and analyze the new desired disulfide bond in the structure of the pertussis S1 enzymatic subunit. Three-dimensional graphics were generated using YASARA molecular graphics suite^[21].

Gene synthesis

The mutant gene of *PTS1* was codon-optimized and chemically synthesized by Pishgaman Gene Transfer Co. (Iran). The synthetic fragment was received in the pUC57 cloning vector. Mutant PTS1 used in the present study had the following mutations: R9K, R13L, and E129G.

Recombinant construct preparation

A set of primers (PerF and PerR) was applied for the amplification of the *PTS1* gene. Each primer had a 5' arm that was complementary to each side of the vector backbone, as illustrated in Figure 1. The PCR amplification of the *PTS1* fragment was carried out using the above primers to result in *PTS1* contain complementary ends to the pPSG-IBA35 expression

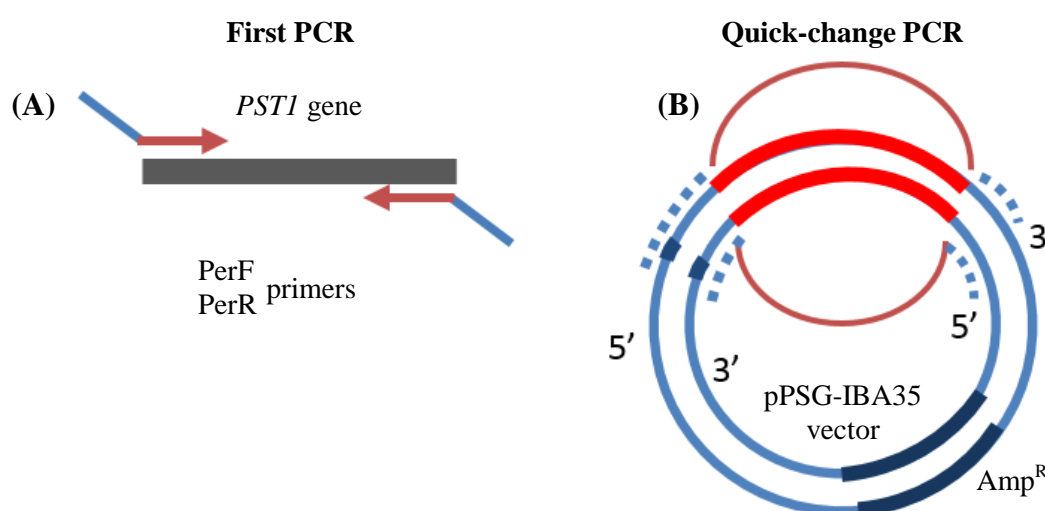


Fig. 1. Design of the mutant PTS1 cloning method. (A) For introducing the *PTS1* gene into pPSG-IBA35 expression vector, two fragments of the multiple cloning site (blue extension) of the vector were added to the inserted gene by a simple PCR. (B) The product of the first PCR was used as a megaprimer in a second quick-change PCR to replace a fragment of the vector by the *PTS1* gene.

vector backbone. The PCR product with the size of about 750 bp was used in a quick-change PCR to synthesize the whole vector with *PTS1* gene inserted between the two arms. Another set of primers employed for the amplification of *LLO* gene (codon 26-266, which is N-terminally his-tagged) had already been sub-cloned in pPSG-IBA35. Codons of the LLO toxin, used in the present study, were codons 22-266. This part of the LLO comprised mostly of the D1 and D2 domains of the protein. Forward primer was complementary to T7 promoter. Reverse primer was hybridized to the last codons of N-terminal half of the *LLO* gene (up to codon 266) and had a 5' arm, which was complementary to the 5'-end of the *PTS1* gene with a Gly2SerGly2 linker (Fig. 2). In addition, the *PTS1* gene was amplified with a set of primers, at the forward with a 5' arm complementary to the last codons of LLO and the linker, and at the reverse with a 5' arm hybridizing to the vector backbone. The PCR amplification of *LLO* and *PTS1* genes with these primers were performed to synthesize two gene fragments. The fragments were fused together using a SOEing PCR protocol, thus leading to a PCR product of about 1500 bp size harboring arms complementary to the pPSG-IBA35 vector backbone. This PCR product was used in a quick-change PCR to synthesize the whole vector with *LLO-PTS1* fusion gene inserted between the two arms. DpnI-treated PCR products were transformed into chemically competent DH5 α bacterial cells using the heat shock method. A single colony from each transformation reaction was cultured in 5-ml LB broth containing 50 $\mu\text{g}\cdot\text{mL}^{-1}$ of ampicillin at 37°C for 16 hours. The recombinant plasmids were

extracted by column method (QIAprep Spin Miniprep kit, QiagenTM, Germany), and the sequence of the construct was confirmed by DNA sequencing.

Recombinant protein expression and purification

Twenty nanograms of the purified plasmids were used to transform *BL21 DE3* and *SHuffle T7* competent cells by the heat shock method. Multiple colonies of *BL21 DE3* and *SHuffle T7* transformants were cultured in a LB medium containing 50 $\mu\text{g}\cdot\text{mL}^{-1}$ of ampicillin at 37 °C and 28 °C, respectively, induced by the addition of 0.1 mM of IPTG when OD^{600nm} reached approximately 0.7. Expression of the recombinant proteins PTS1 and LLO-PTS1 was checked in a number of colonies. The fresh aliquots of recombinant protein expressing clones were used for the inoculation of another 100 mL of the LB medium. Clones were grown until the OD^{600nm} of 0.7, and the expression was induced by adding 0.1 mM of IPTG. After an overnight induction, the bacteria were harvested by centrifugation at 4500 $\times\text{g}$ at room temperature for 10 min. Bacterial pellet was bead-beaten 15 times for 20 seconds with a bench-top vortex with maximum shaking speed (2000 rpm) with 30-second intervals of cooling on ice. Lysis buffer contained Tris (100 mM; pH 8.0), NaCl (300 mM), and glycerol (10%). The soluble fraction was centrifuged, and the supernatant was loaded onto a column filled with Ni-NTA agarose resin (Qiagen) to purify the His-tagged recombinant protein. Equilibration and washing buffers of the chromatography were the same as the lysis buffer. Elution buffer was also the same as the lysis buffer, except for the 300 mM of imidazole. Eluted fractions

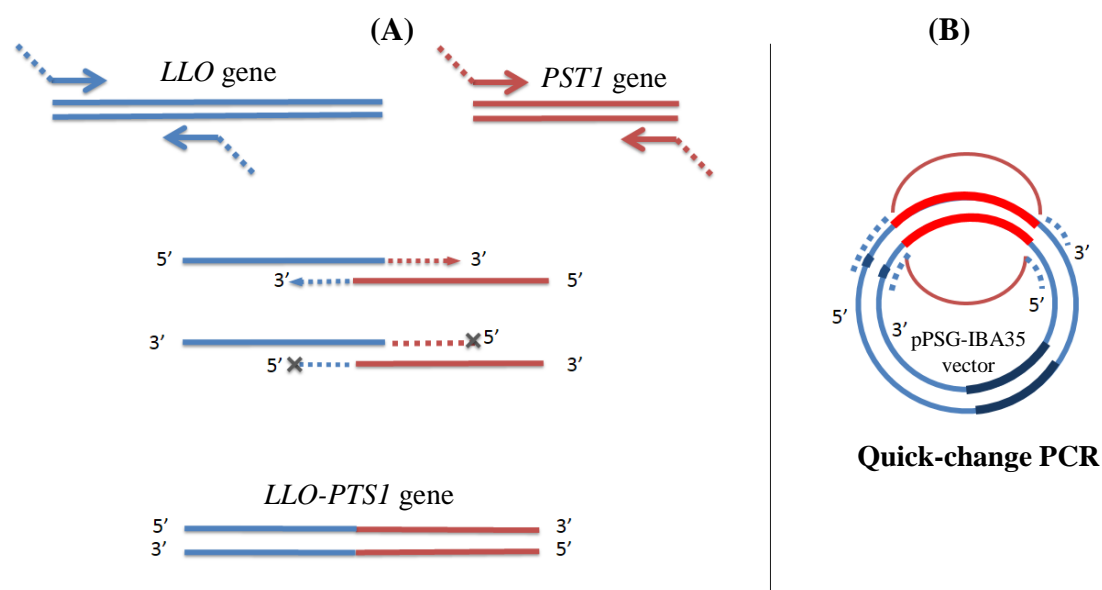


Fig. 2. Design of the fusion LLO-PTS1 cloning method. (A) Two gene fragments were amplified in separate PCRs and ligated together in a SOEing PCR protocol. (B) For introducing the *LLO-PTS1* gene into pPSG-IBA35 expression vector, the product of the SOEing PCR was used as a megaprimer in a second quick-change PCR to replace a fragment of the vector by the *LLO-PTS1* gene.

were analyzed by 12.5% SDS-PAGE for the presence of the recombinant protein, and fractions containing the recombinant protein were dialyzed against the phosphate buffer (pH 8.0) containing 50 mM of NaCl.

Disulfide bond determination

Free SH groups of recombinant proteins purified from *BL21 DE3* and *SHuffle T7 E. coli* strains were examined in a denatured condition (pre-incubating proteins with 6 M of GuHCl for 15 min) using Ellman's reagent. Concentrations of recombinant protein and 5,5'-Dithiobis(2-nitrobenzoic acid) were 2 μ M and 1 mM in a cuvette of 300- μ l volume, respectively. The absorbance of samples at 412 nm was measured, and the number of free SH groups was calculated for the recombinant PTS1 and LLO-PTS1 proteins purified from *BL21 DE3* and *SHuffle T7* strains, as stated before^[22].

Cell culture and MTT assay

Human cell line MCF-7 was purchased from the Pasteur Institute of Iran (Tehran). Cells were cultured and maintained in RPMI-1640 medium supplemented with 10% FBS, streptomycin and penicillin (1%), pH 7.4, in a humidified atmosphere of 95% air plus 5% CO₂ at 37 °C. The anti-proliferative activity of PTS1 and LLO-PTS1 proteins in different concentrations was measured by the MTT assay. Cell viability of each well was calculated as $[A^{570} \text{ of treated cells} / A^{570} \text{ of control untreated cells}] \times 100\%$.

RESULTS

Disulfide bond prediction

The structure of the PTS1 was extracted from 1PRT.pdb file deposited in the PDB database. After an energy minimization step, using the MODIP program and Disulfide By Design, the best disulfide bridges were analyzed, and one of which was selected to introduce into the *PTS1* gene. The distance of the disulfide bond partners from each other in the linear sequence of the protein, ΔG , and dihedral bond is among the parameters concerned with the selection of the best disulfide bond. The potential disulfide bond selected in the present study was Phe53-Asn197 with the numbering of the amino acids based on the first methionine of the PTS1 signal sequence (Fig. 3).

Fusion gene and recombinant construct preparation

Designed *PTS1* gene was chemically synthesized and delivered in the pUC57 cloning vector. PCR of the gene using primers harbouring 5' arms complementary to the *LLO* gene fragment and the pPSG-IBA35 vector

backbone was carried out. In addition, the amplification of the coding sequence of the first 241 amino acids of LLO toxin was performed with designed primers. Quick-change PCR with amplified *PTS1* gene was performed, lanes A and C in Figure 4, to produce recombinant construct pPSG-PTS1. Moreover, the SOEing PCR led to the megaprimer of about 1500 bp, as illustrated in Figure 5 (lane D). Quick-change PCR using this megaprimer was conducted to produce the recombinant construct pPSG-LLO-PTS1.

Expression and purification

Recombinant N-terminal His-tagged PTS1 protein and LLO-PTS1 fusion protein were produced in *BL21 DE3* and *SHuffle T7* strains of *E. coli* in parallel using 0.1 mM concentration of IPTG, as an inducer. The purity and size of the recombinant proteins produced in *BL21 DE3* and *SHuffle T7* strains were the same (lanes D and F; Fig. 4). Yield of the total purified protein was 23.2 mg.L⁻¹ and 13.1 mg.L⁻¹ for the PTS1 protein and 19.4 and 17.4 mg.L⁻¹ for the LLO-PTS1 fusion protein in the two strains, respectively, after the overnight induction of the recombinant protein expression.

Disulfide bond determination of the recombinant proteins

The SH groups titration in denaturing condition indicated that the disulfide bonds of the recombinant mutant PTS1 protein purified from *SHuffle T7* strain have been formed with high efficiency. The ratio of the free SH groups for each molecule was calculated, which proved to be 0.21 and less than 0.05 for proteins

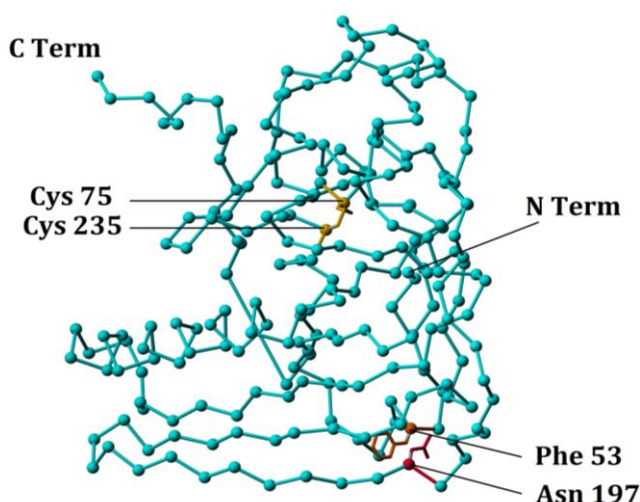


Fig. 3. Modeling of the new disulfide bond in the mutant *PTS1* gene. Two amino acids, Phe53 and Asn197, were chosen to convert to Cys to form a new disulfide bond. Native disulfide bond of the *PTS1* molecule is also shown. The numbering of the amino acids was performed concerning the Met of the signal sequence as the first amino acid.

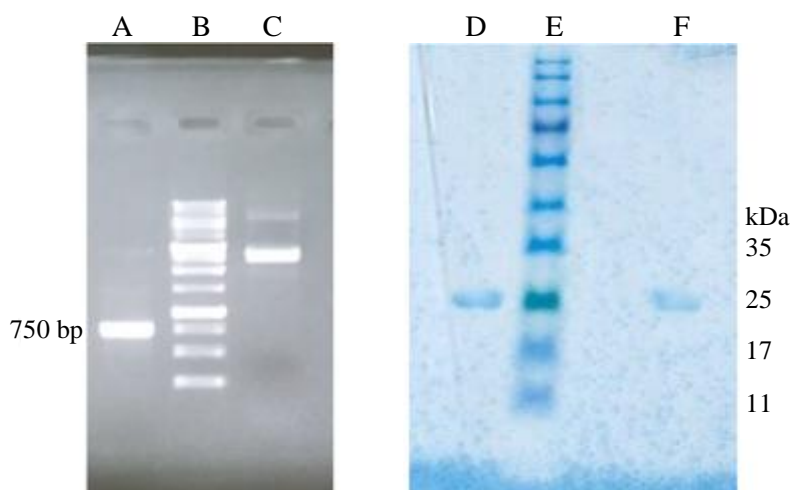


Fig. 4. Quick-change PCR and purity of the mutant PTS1 protein. Quick-change PCR of the mutant *PTS1* gene was performed using the whole gene as megaprimer to introduce the gene into the expression vector pPSG-IBA35. Lane A, *PTS1* gene; lane B, 1 kb DNA size marker; lane C, quick-change PCR product; lane D, mutant PTS1 protein purified from *BL21 DE3* strain; lane E, pre-stained protein size marker; lane F, mutant PTS1 protein purified from *SHuffle T7* strain.

purified from *BL21 DE3* and *SHuffle T7* strains, respectively. Hence, the efficiency of disulfide bond formation in the two proteins was approximately 80% and over 94%, respectively. The same results were procured for the LLO-PTS1 protein.

Cell culture and MTT assay

Serial dilutions of recombinant PTS1 protein and LLO-PTS1 fusion protein were applied to the cultured MCF-7 cells in RPMI 1640 media supplemented with 10% FBS. As shown in Figure 6, the purified recombinant PTS1 protein was not cytotoxic to MCF-7 cell line. The 5-fluorouracil was also tested as the positive control.

DISCUSSION

E. coli has become the most common host for the recombinant protein expression. It is used readily and is economical. In recent years, *SHuffle T7* strain of *E. coli* has been introduced for the cytoplasmic expression of recombinant proteins having disulfide bonds. It benefits from two main features: oxidative environment of cytoplasm and the presence of DsbC chaperone in the cytoplasm^[23]. A few recombinant proteins have successfully been produced in *SHuffle T7* cytoplasm^[24,25]. In the present study, recombinant PTS1 and its fusion form, LLO-PTS1, were produced

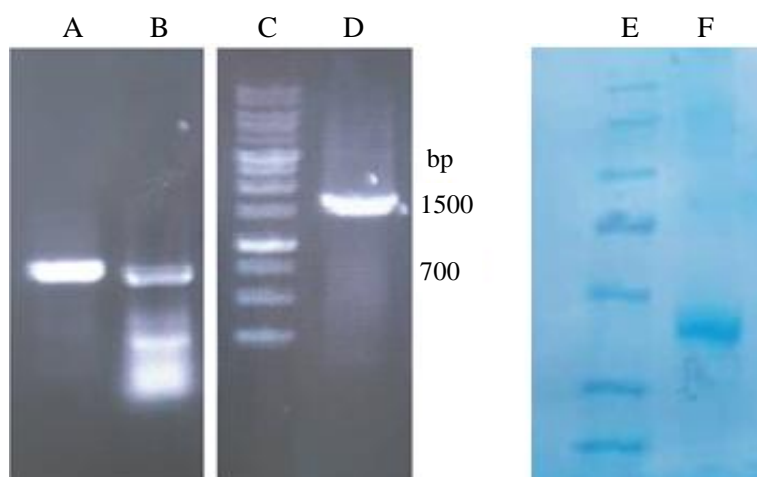


Fig. 5. The SOEing PCR and purity of the LLO-PTS1 protein. SOEing PCR of the mutant *PTS1* gene and *LLO* gene fragment was performed to produce the fusion gene for using as a megaprimer to produce the expression construct of LLO-PTS1 in pPSG-IBA35 vector. Lanes A and B, PCR product of the amplification of the *PTS1* and *LLO* genes; lane C, 1 kb DNA size marker; lane D, SOEing PCR product; lane E, pre-stained protein size marker; lane F, mutant PTS1 protein purified from *SHuffle T7* strain

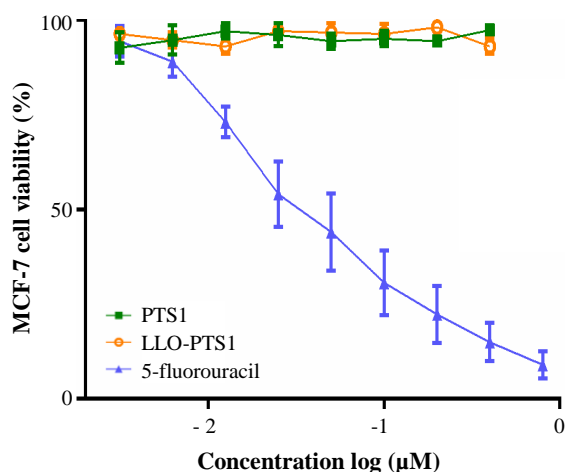


Fig. 6. MCF-7 cell viability in the presence of LLO-PTS1. MCF-7 breast cancer cells were cultured and exposed to increasing concentrations of purified PTS1 and LLO-PTS1 proteins. Independently increasing concentrations of 5-fluorouracil was also tested as the positive control.

in the *SHuffle* strain. Because the proteins produced in the present study were dialyzed in a buffer free of any oxidant and reducing agent after elution from the affinity column, it seems that the more oxidant environment of *SHuffle T7* cytoplasm can be responsible for the higher efficiency of disulfide bond formation in the protein obtained from this strain.

Chemically inactivated toxoid vaccines possess several adverse effects, thereby demanding for the use of alternative strategies to improve safety. Efficacy of the aP vaccines in generating a Th2, instead of Th1, immune response against pertussis antigens is also the matter of controversy^[26-29]. Subunit vaccines are attracting interest due to the lack of native toxin problems. Using new subunit vaccines has been discussed elsewhere^[30]. Higher stability of the subunit vaccines in the formulations prepared for use in populations in comparison with the live vaccines or even high-cost acellular vaccines is another advantage of these substances^[31].

Lee and colleagues^[32] have found that the fusion of the S1, S3, and filamentous haemagglutinin can be expressed as a single recombinant protein. In addition, the produced recombinant antigen was immunogenic in mice, but the antibody titers against the pertussis toxin subunits showed to be weak. In another study, recombinant *Mycobacterium bovis* BCG was utilized as a host for the production of PTS1 protein. This mode of administration provoked a good cellular immunity in a mouse model, but the level of humoral immunity against the pertussis toxin antigen, as the main virulence factor of pertussis, was low^[33]. Diphtheria, tetanus, and pertussis toxin molecule subunits have already been expressed in tobacco and

carrot in a soluble form. It has been indicated that these plant-derived proteins could elicit a specific antibody response in animal models^[34]. Although there are several advantages in large-scale protein production in plants over the conventional biofactories like the animal cell lines or yeast and bacteria cells, there are yet some challenges in the industrial extraction and purification steps. Recombinant pertussis toxin subunits have also been expressed and purified separately to form oligomers *in vitro*. They bear the potential to form *in vitro* assemblies, which are antigenic, and the antibodies they provoke have been displayed to have inhibitory activities against pertussis toxin^[35].

Herein, we have produced a new form of genetically inactivated PTS1 using a cloning strategy, which successfully produced two forms of inactivated PTS1 toxin (PTS1 and fusion LLO-PTS1 proteins) in *E. coli BL21 DE3* and *SHuffle T7* strains. However, the rate of the disulfide bond in *SHuffle T7* strain showed to be more efficient. The analysis of the safety of the proteins produced against a cell line model revealed that both recombinant proteins are safe and non-toxic for the human cell culture model. Our results demonstrated that the recombinant proteins PTS1 and LLO-PTS1 can be expressed in *SHuffle T7* strain cytoplasm and purified in a soluble form with disulfide bonds with high efficiency. Overall, the LLO-PTS1 fusion protein can be formulated as a new generation of the adjuvants to be used as a novel immune-stimulator in the next research studies of pertussis disease.

ACKNOWLEDGMENTS

This study is a part of the Ph.D. dissertation of Hossein Forghani and financially supported by Iran University of Medical Sciences, Tehran, Iran (grant no. 25818) and Shahid Sadoughi University of Medical Sciences, Yazd, Iran. The LLO-PTS1, a novel fusion protein, was registered as a patent in the <https://iripo.ssaa.ir/>.

CONFLICT OF INTEREST. None declared.

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