Contribution of Streptokinase-Domains from Groups G and A (SK2a) Streptococci in Amidolytic /Proteolytic Activities and Fibrin-Dependent Plasminogen Activation: A Domain-Exchange Study

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ABSTRACT

Background: Streptokinase (SK), a heterogeneous plasminogen (PG) activator (PA) protein from groups A, C, and G streptococci (GAS, GCS, GGS, respectively) contains three structural domains (SKα, SKβ, and SKγ). Based on the variable region of SKβ, GAS-SKs (ska) are clustered as SK1 and SK2 (including SK2a/ SK2b), which show low and high fibrinogen (FG)-dependent PG activation properties, respectively. Despite being co-clustered as SK2a, GCS/GGS-SKs (skcg) variants display properties similar to SK1. Herein, by SKβ exchange between GGS (G88) and GAS-SK2a (STAB902) variants, the potential roles of SK domains in amidolytic/proteolytic activity and FG-bound-PG activation are represented. Methods: Two parental SKG88 and SKSTAB902 genes were cloned into the Ndel/Xhol site of pET26b expression vector. The two chimeric SKβ-exchanged constructs (SKC1: αG88βSTABγG88 and SKC2; αSTABβG88γSTAB) were constructed by BstEII/BsiWI digestion/cross-ligation in parental plasmids. SKs were expressed in E. coli and purified by nickel-nitriloacetic acid chromatography. PA potencies of SKs were measured by colorimetric assay. Results: SDS-PAGE and Western-blot analyses confirmed the proper expression of 47-kDa SKs. Analyses indicated that the catalytic efficiency (Kcat/Km) for amidolytic and proteolytic activity were less and moderately dependent on SKβ, respectively. The increase of FG-bound-PG activation for SKSTAB902/SKC1 containing SK2aβ was around six times, whereas for SKG88/SKC2 containing skcgβ, it was four times. Conclusion: Although SKβ has noticeable contribution in FG-bound-PG activation activity, it had minor contribution in fibrin-independent, amidolytic activity. These data might be of interest for engineering fibrin-specific versions of SK.

Keywords: Plasminogen, Streptokinase, Thrombolytic therapy

INTRODUCTION

Conversion of inactive plasminogen (PG) into active protease plasmin (PN) in blood is the unique mechanism of all PG activators (PA) such as tissue-PA (tPA) and streptokinase (SK) that are used as fibrinolytic drugs for the treatment of life-threatening thrombotic disorders like infarction and stroke[1-3]. One of the limiting factors of fibrinolytic drugs is their specificity toward fibrin clots, which is important for the bleeding risk of the patients as a side effect of the treatment[4]. In this regard, PAs are categorized into two main groups, fibrin-specific and fibrin-independent agents[5]. Fibrin-specific PAs, like tPA, target fibrin-bound PG and act specifically onto thrombus, whereas fibrin-independent ones such as SK...
have tendency toward circulatory PG\textsuperscript{[4,6]}. In fact, SK is not a proteolytic enzyme by itself, and its activity relies on several protein-protein interactions. At first, it binds to PG and forms a binary 1:1 complex (activator complex) inducing conformational changes in the molecule that results in the generation of an amidolytically activated SK-PG* complex capable of converting free PG as the substrate to PN (pathway I, conformational activation pathway that is fibrin-independent). Finally, the PG within the activator complex is converted to PN. SK can also bind to PN directly with higher affinity, compared with PG, to form the SK-PN complex, which then it converts other PG substrates to PN (pathway II, direct proteolytic activation pathway)\textsuperscript{[7,8]}

SK is secreted by β-hemolytic Streptococci of the Lancefield groups A (GAS), C (GCS), and G (GGS). SK, as a virulence factor in the streptococcal pathogenesis (especially for GAS) isolated from a moderately virulent GCS (ATCC H46A), was traditionally used as a fibrinolytic drug for decades\textsuperscript{[9,10]}. SK is a 414-residue protein containing three domains: SKα (aa1-144), SKβ (aa145-287), and SKγ (aa 288-414). SK isolated from different groups of streptococci or even within the isolates of the same groups shows a high degree of heterogeneity at gene and protein levels and results in variations in functional characteristic of SKs such as PA potencies and fibrin-specific activity\textsuperscript{[9,11,12]}. Identification of SK heterogeneity and its structure-dependent characteristics encouraged the identification of the functional regions in the SK domains for engineering more efficient SKs as a thrombolytic drug, especially for enhanced fibrin-dependent activity\textsuperscript{[10,13,14]}. SK is the most cost-efficient drug of choice for thrombolytic therapy, particularly in developing countries, and improvement of its therapeutic properties is of high demand\textsuperscript{[4,10,13,14]}

Studies have indicated that the heterogeneity is present in all domains, but the main source of SK variation is mainly in residues 147–218 of β-domain, known as variable one region (SKβ-V1)\textsuperscript{[11]}. According to the phylogenetic analysis of nucleotide sequences of SKβ-V1, GAS-ska alleles are categorized into two major clusters with different functional features, cluster1 (SK1) and cluster2 (SK2), in which cluster2 is further subdivided into two subclusters, SK2a and SK2b\textsuperscript{[15,16]}. Of note, SKs from GCS/GGS (skcg), which exhibit high PA activity in solution (similar to SK1), are categorized into SK2a cluster of the phylogenetic tree, indicating the high similarity in the SKβ-V1 of skcg and cluster2-ska (SK2a) alleles\textsuperscript{[15,16]}. Prior studies on functional properties of SK1 and SK2b have demonstrated that SK2b could activate PG effectively when fibrinogen (FG) is present, whereas despite exhibiting higher PA potency, SK1 does not require FG for efficient PG activation\textsuperscript{[9,16,17]}. In addition, domain-exchange studies between SK1 and SK2b exhibited the major contribution of SK β-domain (SKβ) versus minor role of other domains (α/γ) in determining the PA potency\textsuperscript{[17,18]}. In a recent comparative study, evaluation of the fibrin-dependent activity of recombinant SK form SK2a and SKC-H46A (the commercial source of therapeutic SK) has demonstrated that in the presence of FG, the PA activity of SK2a enhances several folds compared to that of the SKC\textsuperscript{[19]}. However, in none of these prior studies, the fibrin-dependent activity or SK kinetics was/were addressed by domain-exchange strategies, especially in case of SK2a or skcg alleles.

We have recently reported the isolation of SK (skg) with high PA activity from a GGS (SKG88)\textsuperscript{[20]}. In the present study, using SKG88 (with high PA activity) and a well-known SK2a variants from GAS (SK\textsubscript{STAB902}) with low PA activity\textsuperscript{[21]} - we evaluated the contribution of SK domains in kinetics and FG-bound-PG activation via “Molecular (SKβ) domain-exchange strategy” between SK genes of these two groups of streptococci.

**MATERIALS AND METHODS**

**Bacterial variants**

GAS (STAB902) and GGS (G88) with accession numbers CP007041.1 and HM390000.1, respectively were selected as the sources of SK for β-domain exchange. Based on DNA sequences of the variable region of SKβ, SK\textsubscript{STAB902} and SK\textsubscript{GAS} have been reported as SK2a and SK2a co-clustered-skcg alleles, respectively\textsuperscript{[15,20,21]}.

**Isolation of the streptokinase genes and plasmid construction**

In the first step, to construct the recombinant parent SKs, the genomic DNA was isolated by DNA extraction kit (Qiagen, USA). The coding region of sk gene (lacking the signal peptide sequence) was amplified by PCR using primers with inserted restriction sites for direct cloning into pET26b vector (forward primer: NdeI-SKF: 5'-GA CGAGA CATATG ATTGTGGACCTGAGTG-3'; reverse primer: XhoI-SKF 5’-GACACTCAGT ITGTCGTAGGTTATC AG-3'; the sequences corresponding to restriction sites are underlined). Thermal program was set as 30 cycles of 95 °C for 1 min, 56 °C for 1 min, and 72 °C for 3 min, which was followed by a final extension at 72 °C for 10 min. The resulting amplified fragments were digested with NdeI and XhoI and cloned into the same
sites of pET26b expression vector downstream of T7 promoter and in tandem with the fused C-terminus 6xHis-tag to yield two parent molecules, pET26b-SK<sub>G88</sub> and pET26b-SK<sub>STAB902</sub>. (Fig. 1A). In the second step, to construct the chimeric molecules, the β-domain of parent SKs was exchanged. In this context, chimeric SKs (SK<sub>C1</sub> and SK<sub>C2</sub>) were constructed by BstEI/BsiWI digestion of the cloned genes in the parental constructs, from nucleotides 375 to 699 of sk (the variable region of β-domain was composed of 109 residues) and cross-ligation of the resulting fragments (Fig. 1B). E. coli DH5α cells were used for the propagation of plasmids. All cloning steps were performed according to standard procedures[22].

**Protein expression**

E. coli Rosetta strain (Novagen, USA) was used as an expression host for pET26b plasmids according to the manufacturer’s protocol. Briefly, after the transformation of cells with the recombinant plasmids using the standard CaCl<sub>2</sub> method, expression of protein was induced at OD<sub>600</sub> of 0.5–0.6 by isopropyl-β-D-thio-galactoside (IPTG) to a final concentration of 1 mM. Cells were harvested by centrifugation after three hours of incubation at 37 °C and stored at -20 °C for purification steps[22].

**Protein purification**

The expressed SK proteins were purified under native conditions using nickel-nitriloacetic acid (Ni-NTA) affinity chromatography according to manufacturer’s protocol[23]. Briefly, the cell pellets were resuspended in a binding buffer (50 mM of NaH<sub>2</sub>PO<sub>4</sub>, 300 mM of NaCl, and 10 mM of imidazole) with 0.5 mg/ml lysozyme at 2–5 ml per gram wet weight. Following incubation on ice for 30 min, the cells were disrupted by sonication, and supernatant was collected after centrifugation at 10,000 xg at 4 °C for 20-30 min. After the addition of 1 ml Ni-NTA resin to the clear lysate, the mixture was shaken at 4 °C for 60 minutes, loaded on column and washed four times with 4 ml of wash buffer (50 mM of NaH<sub>2</sub>PO<sub>4</sub>, 300 mM of NaCl, and 20 mM of imidazole) and four times with 0.5 ml of elution buffer (50 mM of NaH<sub>2</sub>PO<sub>4</sub>, 300 mM of NaCl, and 250 mM of imidazole).

![Fig. 1.](image-url)

**Fig. 1.** (A) Schematic illustration for the insertion of sk genes into pET26b. Fragments corresponding to sk genes were digested with NdeI and XhoI and ligated with the vector pET26b. ATG, start translation codon derived from vector; 6x His-tag, the tag derived from the vector. (B) Construction of chimeric SKs (SK<sub>C</sub>) by β-domain exchange (exchange of residues 125 through 233 between parental SKs). The unique restriction sites used for sequence exchange are indicated on parental SKs. SK<sub>C1</sub> and SK<sub>C2</sub> are made by β-domain exchanges between SK<sub>G88</sub> and SK<sub>STAB902</sub>-
SDS-PAGE and Western blot analysis

The purity of purified SKs was analyzed on a 12% (w/v) SDS-PAGE gel, and the concentrations were determined by standard Bradford assay and optical density at 280 nm (OD280). For Western blotting, proteins were transferred to nitrocellulose membrane, and the membrane was blocked by 5% BSA. Mouse anti-His monoclonal antibody (Qiagen) was used as the primary antibody, and HRP-labeled goat anti-mouse IgG (Qiagen) as the secondary (tracking) antibody. The bound antibodies were detected using 3,3-diaminobenzidine (Qiagen).

Determination of SK activity

The chromogenic assay is known as an approved internationally standard assay for SK activity (Third International Standard for Streptokinase, National Institute of Biological Standards and Controls, NIBSC, 2004, UK). SK activity was determined by using chromogenic substrate, a synthetic tripeptide H-D-valyl-L-leucyl-L-lysine-p-nitroanilide dihydrochloride (S-2251; Sigma, USA). Purified SK proteins (100 nM) were added to a microtiter plate containing 1 mM of S-2251 and 1 µM of PG (Sigma) at 37 °C in a total volume of 100 µl of the assay buffer (50 mM of Tris-HCl, pH 7.4). Then hydrolysis of S-2251 was measured at 405 nm every 5 min for 60 min in a microplate reader (Synergy 4, UK). To determine fibrin-dependent activity, FG (1 µM) was mixed in a 1:1 stoichiometric ratio with PG (1 µM) and preincubated at 37 °C for 15 min. SKs (100 nM) were added to the mixture, and the change in absorbance at 405 nm was measured at 37 °C after adding S-2251 (final concentration 1 mM). OD at 405 nm was plotted against time and activity rate (slope) was determined from linear portion of the curve. Serial dilutions of Streptase® (CSL, Behring, Germany), a commercially available standard SK, were used to prepare the standard calibration curve based on Hydrolysis of S-2251 by PG, as described before. All reactions were performed in triplicate.

Determination of kinetic constants

For analyzing amidolytic parameters, stoichiometric concentrations of PG and SK (5.5 µM SK and 5 µM PG) were mixed in a 96-well microplate containing the assay buffer (50 mM Tris/HCl, pH 7.5) and incubated at 37 °C for 5 min to construct the SK-PG activator complex. A suitable aliquot of the complex (final concentration, 100 nM) was transferred to the assay buffer along with various concentrations of S2251 (0.1–1 mM) in a total volume of 100 µl. To determine the kinetic parameters for PG activation (proteolytic kinetics), 100 nM of SKs was added to the assay buffer containing 0.1 mM of S2251 and varying concentrations of PG (0.3 to 5.0 µM). The reactions were carried out in a microplate reader at 37 °C. The change in absorbance at 405 nm was monitored for 30 min, and the initial reaction rates were obtained from plotting A405/min. The data were plotted as velocity over substrate concentration, and kinetic parameters of PG activation were determined from Michaelis-Menten (V vs. S) kinetic and inverse (1/V vs. 1/S) Lineweaver-Burk plot using GraphPad Prism 8 (GraphPad Software, San Diego, USA).

Statistical analyses

Differences of SK activities and kinetic parameters among SK variants were determined using unpaired, two-tailed Student’s t-test with 95% confidence intervals. Statistical analysis was carried out using SPSS software version 22.0 (SPSS, Inc., Chicago, IL). All linear regressions were performed applying GraphPad Prism 8, and p values less than 0.05 were considered statistically significant.

RESULTS

Cloning, expression, and purification of the SK

Using the skf and skr primers and genomic DNA as emplate, PCR reactions resulted in a single band of the expected length (1250 bp) of sk gene (Fig. 2A). Cloning steps for the insertion of sk gene in pET26b vector is illustrated in Figure 1A. Chimeric SKs (SKC1 and SKC2) were constructed by the exchange of DNA fragments encoding the 125-233 residue fragments between two parent molecules (Fig. 1B). Restriction enzyme analysis of the recombinant vectors harboring sk genes (Fig. 2B, 2C, and 2D) and nucleotide sequence analysis (not shown) confirmed the accuracy of cloning procedures. The recombinant parent and chimeric proteins were verified by SDS-PAGE (Fig. 3A and 3B) and Western blot analyses and the results (Fig. 3D) indicated the presence of the full length protein with expected molecular weight of 47 kDa. Induction of protein expressions in large-scale cultures (50 ml) and purification of His-tagged SK proteins using Ni-NTA affinity chromatography finally provided us with approximately 5 mg of full length proteins with a purity of more than 90% for each protein that was shown by SDS-PAGE (Fig. 3C).

The steady-state kinetic constants for amidolytic activity

To evaluate the contribution of SKβ on non-proteolytic formation of SK/PG* complex, the amidolytic activity of SK/PG* activator complex was...
studied. The hydrolysis of substrate S2251 by SK/PG* complex is a measure of non-proteolytic activity. The kinetic parameters, including substrate affinity (Km), catalytic activity (kcat), and the constant of catalytic efficiency (Kcat/Km; efficiency of the PG conversion into PN) were measured (Table 1). As shown in Table 1, β-domain exchange did not influence the catalytic efficiencies of resultant chimeras significantly; Kcat of SKC1 (G88-STAB-G88) decreased only 10% (from 0.41 mM of parent SKG88 to 0.36 mM); furthermore, the Kcat attenuated 10% compared to that of the parent SKG88 (from 83.33 min⁻¹ of parent SKG88 to 74.93 min⁻¹), which resulted in almost equal catalytic efficiency (204.99 vs. 205.84 min⁻¹/mM). Likewise, while the Km of SKC2 (STAB-G88-STAB) increased 9% (0.39 vs. 0.36 mM) and the Kcat enhanced 20% relative to the parent molecule, SKSTAB902 (from 30.93 min⁻¹ to 36.96 min⁻¹) resulted in only 10% raise of catalytic efficiency (94.58 vs. 86.09 min⁻¹/mM; p < 0.05; Table 1).

![Figure 2](image1.png)

**Fig. 2.** Agarose gel (1%) of amplified sk genes and restriction analysis of recombinant pET26b-SKs. (A) The coding region of sk gene (lacking the signal peptide sequence) was amplified by PCR using primers with inserted restriction sites for direct cloning into pET26b vector. PCR reactions resulted in a single band of the expected length (1250 bp) of sk gene. Lane 1, DNA marker. Lanes 2 and 3, PCR products of skg88 and skstab902 genes from genomic DNA. (B) Restriction enzyme analysis of the recombinant vector pET26b-SKG88, pET26b-SKC1, and pET26b-SKC2 by NdeI-Xhol yielded 5250 and 1250 bp fragments corresponding to vector and PCR fragments (Lanes 1, 2, and 3, respectively); Lane 4, DNA marker. (C) Digested pET26b-SKG88 and pET26b-SKC1 by BstEII produced two bands with the size of 1400 and 5100 bp (Lanes 1 and 2, respectively); lane 3, DNA marker. (D) The size and pattern of DNA markers. The corresponding bands are indicated by arrows, and the sizes of the bands of DNA marker are illustrated on the right. DNA marker (1 kb; Thermo Fisher Scientific SM0311, USA).

![Figure 3](image2.png)

**Fig. 3.** Analysis of expressed proteins by SDS-PAGE and Western blotting. (A) SDS-PAGE (12%) of total protein extracted from E. coli Rosetta/pSKG88 cells. Lane 1 corresponds to uninduced bacterial cells; lane 2, protein marker 10-180 kDa; lane 3-6, total protein extracted from IPTG (1 mM) induced E. coli Rosetta/pSKG88, pSKSTAB902, and pSKC1-2 cells, respectively. (B) The size and pattern of the bands of protein marker 10-180 kDa. (C) Analysis of purified proteins by SDS-PAGE (12%). Lane 1, molecular weight marker; lane 2-5, purified SKG88, SKSTAB902, SKC1, and SKC2 proteins, respectively. (D) Western-blot analysis of SKC1 and SKC2 proteins. Lanes 1 and 3, crude lysis of E. coli Rosetta cells expressing SKC1 and SKC2, respectively after induction by IPTG (1 mM); lane 2, crude lysis of E. coli Rosetta cells before induction (no band was observed); lane 4, molecular weight marker. The arrows indicate the band of 47 kDa SK.
TABLE 1. The amidolytic and proteolytic kinetic parameters of SK variants

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<th>SK variants</th>
<th>Amidolytic constants</th>
<th>Proteolytic constants</th>
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<tr>
<td></td>
<td>Km (mM)</td>
<td>Kcat (min⁻¹)</td>
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<tr>
<td>SKG8</td>
<td>0.41 ± 0.006</td>
<td>83.33 ± 2.82</td>
</tr>
<tr>
<td>SKSTAB902</td>
<td>0.36 ± 0.007</td>
<td>30.93 ± 0.78</td>
</tr>
<tr>
<td>SKC1</td>
<td>0.46 ± 0.026</td>
<td>74.93 ± 3.58</td>
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<tr>
<td>SKC2</td>
<td>0.39 ± 0.063</td>
<td>36.96 ± 3.75</td>
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*The amidolytic kinetic parameters of SK variants were determined by pre-complexing equimolar ratios of SK and PG.

The steady-state kinetic constants for proteolytic activity

The proteolytic activity of SK variants after equimolar SK.PG complex formation was assayed against a concentration range of substrate PG. As shown in Table 1, the Km of SKC1 raised 1.5fold (1.09 vs. 0.77 µM), and the Kcat raised 13% (22.59 vs. 19.82 min⁻¹) compared with that of SKG8 that led to 20% lower catalytic efficiency (from 25.67 of parent SKG8 decreased to 20.70 min⁻¹/µM). The proteolytic constants of SKC2 were also altered relative to the parent SKSTAB902. The catalytic efficiency of SKC2 increased 20% (3.12 vs. 2.56 min⁻¹/µM), since the Kcat decreased 10% (3.73 vs. 4.15 µM) and the Kcat raised 10% (11.68 vs. 10.64 min⁻¹) compared with those of SKSTAB902 (p < 0.05; Table 1). These alterations were in accordance with the change of specific activities and imply that β-domain exchange influences the conformational and functional changes of SK yielding alteration of the kinetic constants of proteolytic pathway and conversion of PG substrate to PN, which led to different activities of SK variants.

### DISCUSSION

The rationale for performing the present study was to gain insights into the degree of fibrin dependency and kinetic differences of SKG8 from group G streptococci (skcg allele) compared with the SKSTAB902 from cluster 2a, group A streptococci, and the role of β-domain in these characteristics. To the best of our knowledge, there is no prior study on the role of β-domain in fibrin-dependent mode of action or SK kinetics by domain-exchange strategies, especially for SK2a or skcg alleles. Our results confirmed the higher increase in SK activities of SKSTAB902 than skcg allele in the presence of FG and indicated the major contribution of β-domain in conferring this feature, which might lead
to designing fibrin-specific generations of SK. As shown in Figures 2A and 3, results of PCR and SDS-PAGE/ Western blot analyses indicated the expected length (1250 bp) of sk gene and SK protein (47 kDa), which is in agreement with prior molecular isolation and expression studies for SK (reviewed in [10]).

According to kinetic results (Table 1), the β-domain exchange did not cover any significant alteration in the amidolytic catalytic efficiency (Kcat/Km) of SK.PG* activator complexes of the chimeric SKs (SKC1 and SKC2) compared to their parental SKs. Indeed, these results revealed a major role for α/γ-domains in determination of amidolytic activity, which is consistent with several prior reports on the potential role of α/γ-domains in interaction with PG and formation of the SK.PG* activator complexes [25–29]. In this context, the critical role of Ile1 in α-domain of SK for the formation of SK.PG* activator complex through establishing a salt bridge with Asp741 of PG, which is essential for the induction of an active site in PG is emphasized [27]. In addition, a surface-exposed loop in residues 88-97 of α-domain has been also reported. This loop not only is involved in SK.PG* activator complex formation but also interacts with the catalytic domain of PG (microplasmin). Thus, this behavior might have a crucial role in catalytic turnover of the substrate PG while minimally affecting enzyme-substrate affinity [35,36]. Potentially, the key role of Arg324, Asp325, Lys332, and Lys334 as well as residues between 314-342 in the γ-domain for the amidolytic activity of the SK.PG* activator complex has been proposed [28,29].

The Kcat/Km values of proteolytic activity (as an indicator for the efficiency of conversion of substrate PG into PN) for chimeric SKC1 and SKC2 compared to parental SKs showed more significant alterations compared to the amidolytic efficiency (Table 1). Of note, the Km values, which show the substrate affinity, were affected more significantly compared to the Kcat, indicating the catalytic turnover (which implies the important role of the exchanged segment in affinity of the activator complex to substrate PG). Our finding is in accordance with several studies addressing the importance of SKβ in the proteolytic activity of SK. Indeed, by bridging the SKα and SKγ, SKβ mediates high affinity interaction between SK and PG [20,30–32]. This domain seems to involve in high-affinity interactions between SK and PG substrate, as well as strong binding of PG substrate to the proteolytic complex and efficient conversion of PG substrate to PN [31]. Our results are in line with reports on the critical role of loop 170 of SKβ in mediating catalytic turnover of the substrate PG (indicated by Kcat of the proteolytic pathway) [8,25]. In further support of our results, the role of loop 250 in SKβ for PG recognition by active SK.PG* complex and PG docking has also been suggested [22]. It should be noted, however, that several residues in other domains of SK, like residues 314–347 and 285–414 of SKγ, have been proposed for contribution in the processing of PG by the SK-PN complex and catalytic activation of PG [28].

In general, the presence of FG enhanced the activity of all SKs, but its effect was around two times higher for SK2αβ containing SKSTAB902/SKC1 than skcgβ containing SKG88/SKC2 proteins (6.1/5.7 vs. 3.5/4.5-fold enhancement of activity, respectively; Fig. 4 and Table 2). These results accord with prior reports on generally positive effect of the binding of FG to PG to change its conformation in favor of enhanced activation of PG [16,19,33], which has been higher for GAS-SK2α variants compared to skc-SK from GCS [19]. In further agreement, it has also been reported that the PA activity of SK1 and SK2b variants enhances 1.2-1.8 and 10-18-fold, respectively in the presence of FG [33]. However, in these prior studies, the role of SK domains or special residues on this property was not elucidated.

Identification of the SK domains involved in FG-bound-PG activation might help to improve the fibrin-specific characteristics of SK for therapeutic purposes [15]. Consistent with a recent study, SKG88, as represented in Figure 4, showed high intrinsic FG-bound-PG activation [20], which was about fourfold higher than SKSTAB902. Interestingly, this high FG-bound-PG activation further enhanced in SKC1, while in the absence of FG, PA potency of SKC1 was still lower than the parental SKG88. Of note, these characteristics of SKC1 might be of interest for development of a fibrin-specific version of SK for targeted fibrinolysis [15]. Collectively, these observations might indicate both the contribution of

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<th>Table 2. PA activity in the absence and presence of fibrinogen (FG)</th>
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<td><strong>The parent/ chimeric SK</strong></td>
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<td>SKG88</td>
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<td>SKSTAB902</td>
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<td>SKC1</td>
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SK2αβ of SKcTAIB9023 and SKαγ of scek (SKGRS) in PA properties of SKcC1. Almost the same justification might be considered for SKcC2 compared to the parental SKs, which implies the collaborative contributions of β- and αγ-domains in FG-bound-PG activation and agrees with proposed negative role of the first 59 residues of SKα in fibrin-dependent mode of the SK action [34].

Taken together, by molecular exchanging SKβ-domains between groups G and A (SK2α) streptococci and recombinant expression of the two chimeric and two parental SKs, we could assess and compare the kinetics and FG-bound-PG activation of the four SKs to gain insights into the role of SKβ and SKαγ in these functional characteristics. To our best of knowledge, this is the first report on domain exchange study between groups G and A streptococci. Our results indicated the minor role of SKβ compared to SKαγ in fibrin-independent amidolytic activity, while the reverse was demonstrated for fibrin-dependent proteolytic activity and FG-bound-PG activation. The obtained data might be of interest for engineering and development of a fibrin-specific version of SK for targeted fibrinolysis and thrombolytic therapy.

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CONFLICT OF INTEREST. None declared.

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