

PIK3CA Mutation Analysis in Iranian Patients with Gastric Cancer

Mostafa Iranpour¹, Mahyar Nourian¹, Sana Saffari², Esmail Samizadeh³, Mahdieh Mirghafori⁴, Shahrokh Iravani^{1*} and Soudeh Ghafouri-Fard^{5*}

¹AJA Cancer Epidemiology Research and Treatment Center (AJA-CERTC), AJA University of Medical Sciences, Tehran, Iran; ²Department of Biology, Tehran North Branch, Islamic Azad University, Tehran, Iran; ³Department of Pathology, AJA University of Medical Sciences, Tehran, Iran; ⁴Department of Biology and Anatomical Sciences, Shahid Beheshti University of Medical Sciences, Tehran, Iran; ⁵Department of Medical Genetics, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Received 9 January 2018; revised 22 April 2018; accepted 25 April 2018

ABSTRACT

Background: Aberrant activation of phosphatidylinositol-3 kinases (PI3K)/AKT/mTOR (mammalian target of rapamycin) pathway is a critical event during gastric cancer progression. Selective function of AKT inhibitor AZD5363 in *PIK3CA* mutant gastric cancer necessitates the assessment of *PIK3CA* mutations in these patients. **Methods:** The study included 100 patients with gastric cancer who underwent surgical resection at Imam Reza Hospital, Tehran, Iran, between January 2009 and December 2016. Mutations in codon 1047 of *PIK3CA* were evaluated by tetra-primer ARMS-PCR and direct sequencing methods. **Results:** We detected p.H1047R and p.H1047L in 8 and 3 samples, respectively. Also, a significant association was found between *PIK3CA* mutations and lymphatic invasion. Kaplan-Meier analysis demonstrated no significant differences in overall survival between patients with and without mutations. **Conclusion:** Our study detected gain-of-function mutations in exon 20 of *PIK3CA* gene in 11% of gastric cancer patients. Future studies are needed to assess the mutation rate in other regions of this gene to find eligible patients for targeted therapies.

Keywords: Mutation, Phosphatidylinositol-3 kinases, Somach neoplasms

Corresponding Authors: Shahrokh Iravani and Soudeh Ghafouri-Fard
AJA Cancer Epidemiology Research and Treatment Center (AJA- CERTC), AJA University of Medical Sciences, Tehran, Iran;
Tel. & Fax: (+98-21) 43825064; E-mail: iravanishahrokh@yahoo.com

Department of Medical Genetics, Shahid Beheshti University of Medical Sciences, Tehran, Iran;
Tel. & Fax: (+98-21) 23872572; E-mail: s.ghafourifard@sbmu.ac.ir

INTRODUCTION

Gastric cancer is regarded as one of the most frequent causes of cancer death all over the world^[1]. Numerous genetic and environmental factors known to be implicated in the pathogenesis of this disease, as well as in patients' response to treatment modalities^[2]. Dysregulation of phosphatidylinositol-3 kinases (PI3K)/AKT/ mTOR (mammalian target of rapamycin) signaling pathway has been recognized as the causative agent of gastric

cancer pathogenesis^[3]. This pathway has a crucial role in the regulation of cell growth, metabolism, vesicular trafficking, cell apoptosis, metastasis, and response to chemotherapy^[4]. One of the most investigated components of this pathway is the *PIK3CA* gene encoding p110 α . Activating mutations in this gene have been detected in a variety of human malignancies, leading to the constitutive expression of p110 α and to the induction of downstream signaling proteins, including AKT/protein kinase B, mTOR, and ribosomal protein S6 kinase^[4]. Exons 20 and 9 of this

gene, which code for kinase and helical domains, respectively, are considered as mutation hotspots in cancer cells^[5, 6]. The advent of novel targeted therapies against PI3K pathway has further increased the significance of the assessment of *PIK3CA* gene alterations in cancers. Most notably, *PIK3CA* mutation has been identified as an essential determinant of response to AKT inhibitor AZD5363 in gastric cancer cell lines^[7]. Moreover, clinical evidence supports the essential role of *PIK3CA* mutations in the determination of response to ipatasertib and AZD5363 AKT inhibitors^[8]. Few studies have assessed the frequency of *PIK3CA* gene mutations in gastric cancer patients and their influence on patients' survival^[6]. However, based on the differences in etiologic factors of gastric cancer and disparities in patients' outcome in distinct ethnicities^[9], it is important to assess the rate of *PIK3CA* gene mutations in each region in association with patients' survival. Consequently, in the present study, we evaluated the mutation status of a single hotspot codon in *PIK3CA* gene in Iranian patients with gastric cancer.

MATERIALS AND METHODS

Patients

The present retrospective study included 100 gastric cancer patients who underwent surgical resection at Imam Reza Hospital, Tehran, Iran, between January 2009 and December 2016. The patients were followed-up for 12 months averagely to assess their overall survival. This parameter was delineated as the time from the date of the surgery to the date of demise. Staging was carried out based on the American Joint Committee on Cancer Staging Manual (7th edition)^[10]. The study was approved by the ethical committee of Medical School of Islamic Republic of Iran Army (AJA University). Informed consent was obtained from all participants.

Genomic DNA extraction

Paraffin-embedded tissue samples of surgically resected gastric cancers were assessed by a pathologist to spot cancer tissues. Genomic DNA was extracted from marked regions including tumor tissues without adjacent normal tissues using Exgene™ FFPE Tissue DNA kit (GeneAll, Korea).

PCR amplification of codon 1047 of *PIK3CA* gene

Primers for tetra-primer ARMS-PCR were designed by PRIMER1 online tool^[11]. Table 1 shows the nucleotide sequences of primers used in the current study. The PCR reaction was performed in a 25 µl total volume containing 100 ng of genomic DNA, 12.5 µl of Taq DNA Pol 2× Master Mix Red (Ampliqon, Denmark), and 0.5 µl of inner and outer primers. Then 10% of the samples were sequenced by using the ABI 3730xl DNA analyzer (Macrogen, Korea) in order to confirm the results of tetra-primer ARMS-PCR.

Statistical analysis

The results were statistically analyzed by SPSS software (version 16, Chicago, USA) using *t*-test and chi-square test (χ^2) for comparing means and proportional differences among the clinical features. Survival time was assessed by the Kaplan-Meier method and compared using the log-rank test.

RESULTS

PIK3CA codon 1047 mutations in gastric cancer patients

Among the 100 patients who had undergone gastric resection, 11 (11%) were recognized by tetra-primer ARMS-PCR and also direct sequencing to harbor *PIK3CA* codon 1047 mutations (Fig. 1). Eight patients were positive for c.3140A>G (p.H1047R) and three were positive for c.3140A>T (p.H1047L) mutations.

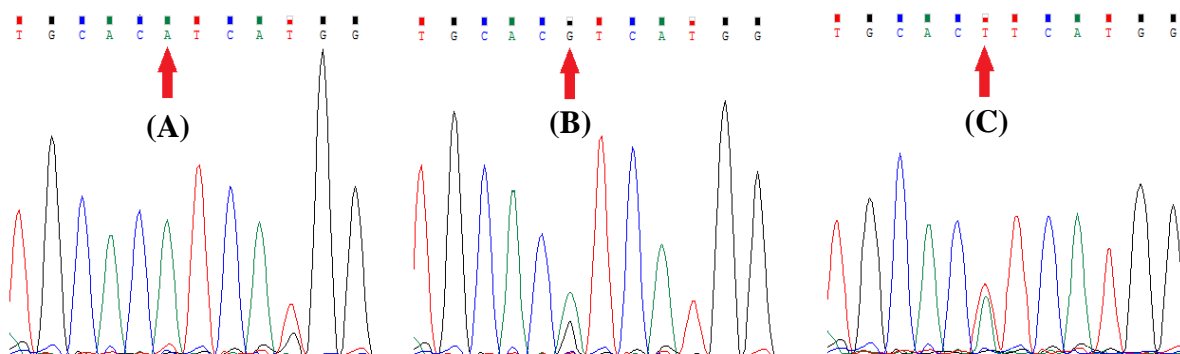


Fig. 1. The results of Sanger sequencing for confirmation of tetra-primer ARMS-PCR. The normal sequence (A), c.3140A>G (B), and c. 3140A>T (C) mutations.

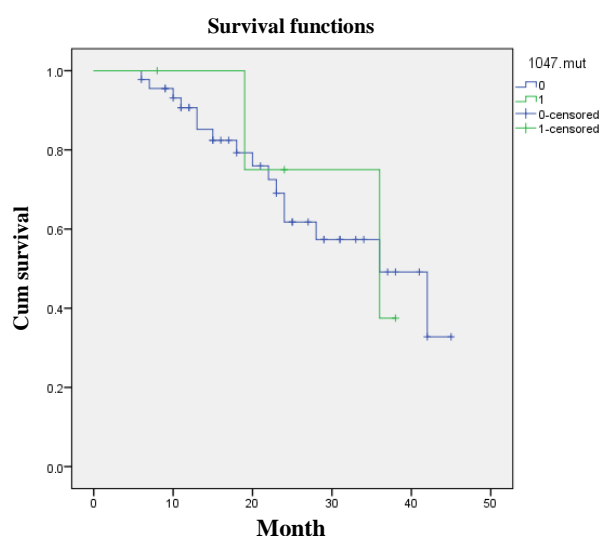


Fig. 2. Kaplan-Meier curves showing the relationship between codon 1047 mutational status and the overall survival of gastric cancer patients.

PIK3CA codon 1047 mutations and patients' clinical and demographic data

Table 2 shows the details of statistical analysis of the associations between mutation status and patients' demographic and clinical data. No significant association was found between mutation status and patients' age or gender. However, a significant correlation was detected between the presence of mutation and lymph node involvement ($p = 0.036$).

PIK3CA codon 1047 mutations and patients' survival

We evaluated the effect of *PIK3CA* codon 1047 mutations on patients' survival after the surgical resection of gastric cancer. The median follow-up time for patients was 20 months. Kaplan-Meier analysis showed no significant differences in overall survival (log rank $p = 0.55$) between the patients with and without codon 1047 mutations (Fig. 2).

DISCUSSION

In the present study, we demonstrated the presence of activating mutations in a single codon of *PIK3CA* gene in a significant number of Iranian patients with gastric cancer. Previous studies have detected mutations in other regions of this gene in gastric cancer samples^[5, 6]. Our detected mutation rate in this single codon was comparable with the total rate of detected mutations in two *PIK3CA* exons in a study conducted in Japanese patients^[6], implying the presence of higher mutation rate in entire gene in Iranian patients. Harada *et al.*^[6] have analyzed exons 9 and 20 of this gene in Japanese patients with gastric cancer. They reported a total mutation rate of 12% with p.H1047R and p.H1047L being seen in 4% and 0% of patients. Sukawa *et al.*^[12] have detected *PIK3CA* mutations in 8.7% of Japanese gastric cancer patients in their assessment of exons 1, 9, and 20 and reported p.H1047R and p.H1047L mutations in 2.6% and 0% of patients, respectively. Besides, Lee *et al.*^[13] have reported a total rate of 7.7% for exons 9 and 20 mutations in Korean gastric cancer patients. Meanwhile, Barbi *et al.*^[14] have reported a total mutation rate of 16% in mutation analysis of exons 9 and 20 in Italian gastric cancer patients with p.H1047R, being the most prevalent mutation. Our data are in accordance with the reported data in gastric cancer patients regarding the common occurrence of p.H1047R mutation. However, p.H1047L mutation has not been previously reported in gastric cancer.

Evaluation of biological and biochemical characteristics of p.H1047L, p.H1047Y, and p.H1047R mutations in the kinase domain of *PIK3CA* gene have shown that all of these mutations pack against the hinge region of the activation loop and trigger rapamycin-sensitive oncogenic transformation of chicken embryo fibroblasts through the induction of AKT and mTOR signaling^[15]. Consequently, no functional difference is predicted to occur in patients carrying any of these mentioned mutations.

Table 1. The nucleotide sequences of primers used in this study

Primers	Primer sequence	Product length (bp)
Inner	F: 5'-ATTTTCATGAAACAAATGAATGATGCACA-3' (A-allele)	149
	R1: 5'-CCATTTTTGTTGTCCAGCCACCATGAC-3' (G-allele)	198
	R2: 5'-CCATTTTTGTTGTCCAGCCACCATGAA-3' (T-allele)	198
Outer	F: 5'-CTATTCGACAGCATGCCAATCTCTTCAT-3' R: 5'-TTAACAGTGCAGTGTGGAATCCAGAGTG-3'	292

Table 2. Associations between mutation status and patients' demographic and clinical data

Characteristics	Mutation (%) (n = 11)	Wild-type (%) (n = 89)	p value
Age (years)	61.55 ± 11.50	62.87 ± 9.92	0.683
Gender			
Female	5 (5)	33 (33)	0.589
Male	6 (6)	56 (56)	
Lymph node involvement			
Positive	8 (8)	37 (37)	0.036
Negative	2 (2)	39 (39)	
Missing	1 (1)	13 (13)	
Familial history of cancer			
Positive	1 (1)	7 (7)	0.833
Negative	4 (4)	36 (36)	
Missing	6 (6)	46 (46)	

Previous studies have reported no associations between *PIK3CA* gene mutations and gastric patients' clinicopathologic features^[6,12,14]. Consistent with these investigations, we observed no significant association between *PIK3CA* gene mutations and patients' survival, which might be due to the occurrence of these mutations in the early stages of carcinogenesis that are shared between all patients. Therefore, other factors might override this event and change the disease course. In contrast, we found a significant correlation between the occurrence of codon 1047 mutations and lymph node involvement. As a result, we suggest the evaluation of other hotspots in *PIK3CA* gene to assess the clinical significance of such finding.

Assessment of somatic *PIK3CA* mutations has been suggested as an effective biomarker for recruitment of patients in clinical trials of AKT inhibitors^[8]. The high incidence of mutation in one codon of *PIK3CA* gene in our assessed population of gastric cancer patients warrants the evaluation of total coding region of this gene in Iranian patients to confirm the contribution of these mutations in gastric cancer and to develop personalized therapeutic regimens for patients.

ACKNOWLEDGEMENTS

The current study was supported by a grant from Medical School of Islamic Republic of Iran Army (AJA University; Grant number: 995757), Tehran, Iran.

CONFLICT OF INTEREST. None declared.

REFERENCES

1. Imani-Saber Z, Yousefi-Razin E, Javaheri M, Mirfakhraie R, Motaleb G, Ghafouri-Fard S.

Promyelocytic leukemia (PML) gene mutations may not contribute to gastric adenocarcinoma development. *Asian Pacific journal of cancer prevention* 2015; **16**(8): 3523-3525.

2. Imani-Saber Z, Ghafouri-Fard S. *In silico* interaction and docking studies indicate a new mechanism for PML dysfunction in gastric cancer and suggest imatinib as a drug to restore function. *Asian Pacific journal of cancer prevention* 2015; **16**(12): 5005-5006.
3. Singh SS, Yap WN, Arfuso F, Kar S, Wang C, Cai W, Dharmarajan AM, Sethi G, Kumar AP. Targeting the PI3K/Akt signaling pathway in gastric carcinoma: A reality for personalized medicine? *World journal of gastroenterology* 2015; **21**(43): 12261-12273.
4. Arcaro A, Guerreiro AS. The phosphoinositide 3-kinase pathway in human cancer: Genetic alterations and therapeutic implications. *Curr genomics* 2007; **8**(5): 271-306.
5. Samuels Y, Diaz LA, Jr., Schmidt-Kittler O, Cummins JM, Delong L, Cheong I, Rago C, Huso DL, Lengauer C, Kinzler KW, Vogelstein B, Velculescu VE. Mutant *PIK3CA* promotes cell growth and invasion of human cancer cells. *Cancer cell* 2005; **7**(6): 561-573.
6. Harada K, Baba Y, Shigaki H, Ishimoto T, Miyake K, Kosumi K, Tokunaga R, Izumi D, Ohuchi M, Nakamura K, Kiyozumi Y, Kurashige J, Iwatsuki M, Miyamoto Y, Sakamoto Y, Yoshida N, Watanabe M, Baba H. Prognostic and clinical impact of *PIK3CA* mutation in gastric cancer: pyrosequencing technology and literature review. *BMC cancer* 2016; **16**: 400.
7. Li J, Davies BR, Han S, Zhou M, Bai Y, Zhang J, Xu Y, Tang L, Wang H, Liu YJ, Yin X, Ji Q, Yu DH. The AKT inhibitor AZD5363 is selectively active in *PI3KCA* mutant gastric cancer, and sensitizes a patient-derived gastric cancer xenograft model with PTEN loss to Taxotere. *Journal of translational medicine* 2013; **11**: 241.
8. Nitulescu GM, Margina D, Juzenas P, Peng Q, Olaru OT, Saloustros E, Fenga C, Spandidos DA, Libra M, Tsatsakis AM. Akt inhibitors in cancer treatment: The long journey from drug discovery to clinical use (Review). *International journal of oncology* 2016; **48**(3): 869-885.

9. Merchant SJ, Li L, Kim J. Racial and ethnic disparities in gastric cancer outcomes: More important than surgical technique? *World journal of gastroenterology: World journal of gastroenterology* 2014; **20**(33): 11546-11551.
10. Washington K. 7th edition of the AJCC cancer staging manual: stomach. *Annals of surgical oncology* 2010; **17**(12): 3077-3079
11. Ye S, Dhillon S, Ke X, Collins AR, Day IN. An efficient procedure for genotyping single nucleotide polymorphisms. *Nucleic acids research* 2001; **29**(17): E88-E88.
12. Sukawa Y, Yamamoto H, Noshō K, Kunimoto H, Suzuki H, Adachi Y, Nakazawa M, Nobuoka T, Kawayama M, Mikami M, Matsuno T, Hasegawa T, Hirata K, Imai K, Shinomura Y. Alterations in the human epidermal growth factor receptor 2-phosphatidylinositol 3-kinase-v-Akt pathway in gastric cancer. *World journal of gastroenterology* 2012; **18**(45): 6577-6586.
13. Lee H, Hwang IS, Choi IJ, Kang YN, Park KU, Lee JH. Are PIK3CA mutation and amplification associated with clinicopathological characteristics of gastric cancer? *Asian pacific journal of cancer prevention*; **16**(11): 4493-4496.
14. Barbi S, Cataldo I, De Manzoni G, Bersani S, Lamba S, Mattuzzi S, Bardelli A, Scarpa A. The analysis of PIK3CA mutations in gastric carcinoma and metanalysis of literature suggest that exon-selectivity is a signature of cancer type. *Journal of experimental and clinical cancer research* 2010; **29**(1): 32.
15. Gymnopoulos M, Elsliger MA, Vogt PK. Rare cancer-specific mutations in PIK3CA show gain of function. *Proceedings of the national academy of sciences of the United States of America* 2007; **104**(13): 5569-5574.