Equilibrium Isotherm, Kinetic Modeling, Optimization, and Characterization Studies of Cadmium Adsorption by Surface-Engineered Escherichia coli

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ABSTRACT

Background: Amongst the methods that remove heavy metals from environment, biosorption approaches have received increased attention because of their environmentally friendly and cost-effective feature, as well as their superior performances. Methods: In the present study, we investigated the ability of a surface-engineered Escherichia coli, carrying the cyanobacterial metallothionein on the cell surface, in the removal of Ca (II) from solution under different experimental conditions. The biosorption process was optimized using central composite design. In parallel, the kinetics of metal biosorption was studied, and the rate constants of different kinetic models were calculated. Results: Cadmium biosorption is followed by the second-order kinetics. Freundlich and Langmuir equations were used to analyze sorption data; characteristic parameters were determined for each adsorption isotherm. The biosorption process was optimized using the central composite design. The optimal cadmium sorption capacity (284.69 nmol/mg biomass) was obtained at 40°C (pH 8) and a biomass dosage of 10 mg. The influence of two elutants, EDTA and CaCl\textsubscript{2}, was also assessed on metal recovery. Approximately, 68.58% and 56.54% of the adsorbed cadmium were removed by EDTA and CaCl\textsubscript{2} during desorption, respectively. The Fourier transform infrared spectrophotometer (FTIR) analysis indicated that carboxyl, amino, phosphoryl, thiol, and hydroxyl are the main chemical groups involved in the cadmium bioadsorption process. Conclusion: Results from this study implied that chemical adsorption on the heterogeneous surface of E. coli E and optimization of adsorption parameters provides a highly efficient bioadsorbent. DOI: 10.18869/acadpub.ibj.21.6.380

Keywords: Adsorption, Kinetics, Response surface methodology, Fourier transform infrared spectrophotometer

INTRODUCTION

Cadmium contamination has serious potential implications for soil, water, and human health\textsuperscript{[1]}. Unlike organic pollutants, heavy metals are not degradable and may persist for relatively long periods of time in aquatic and terrestrial environments\textsuperscript{[2]}. Amongst the methods that remove heavy metals from environment, biosorption approaches have received increased attention because of their environmentally friendly and cost-effective feature, as well as their superior performances. Due to these properties, the biosorption is usually preferred to physical or chemical remediation technologies. Biosorption takes advantage...
of naturally occurring biomaterials for heavy metal removal\[3\]. Biomaterials such as bacteria, yeasts, algae, and some plants have been successfully applied for pollutant adsorption, and it has been shown that these materials demonstrate a high capacity in removing large amounts of heavy metals from wastewaters. Other benefits of using biomaterials include lower cost of investment, lower energy consumption, and high removal efficiency of the heavy metals\[4\]. However, natural biomaterials have neither a high selectivity nor a high capacity for the adsorption of heavy metals. Surface-engineered bacteria solve these problems by expressing the large amounts of selective sorbent motifs, such as metallothioneins (MTs), on their surfaces. MTs are low-molecular-weight (6–7 kDa), cysteine-rich, metal-binding proteins found in animals, higher plants, eukaryotic microorganisms, and some prokaryotes\[5\]. MTs have the ability to bind to both physiological (such as zinc, copper, and selenium) and xenobiotic (such as cadmium, mercury, silver, and arsenic) heavy metals through the thiol group of its cysteine residues, which represents nearly the 30% of its amino acidic residues\[6\].

The main advantages associated with the surface expression of metal adsorbents, in comparison to intracellular expression, include the elimination of the time-consuming and rate-limiting step of crossing the membrane, prevention of interference with redox pathways in the cytosol, the uptake of any heavy metal of interest by expressing its binding peptide on the surface of cell, and recycling of the biosorbents\[7,11\]. Various bacterial-based systems have been developed for heavy metal removal by using microorganisms over-expressing MTs on their surfaces\[9,12\]. Surface-engineered bacteria show enhanced adsorption of heavy metals and offer a promising strategy with respect to the development of bacterial-based biosorbents for the removal of heavy metal ions from wastewater. Cadmium is considered as one of the most toxic heavy metals, and in fact, cyanobacterial MTs such as SmtA have been shown to have high affinity for Cd (II), Zn (II), and Cu (II) ions\[13\]. In our previous research, the Lipoprotein-outer membrane protein A (Lpp′-OmpA) system was used for surface display of cyanobacterial MTs\[14\]. The ability of these MTs to be displayed on the bacterial surface has not been evaluated so far. The Lpp′-OmpA system consists of a signal sequence and the first nine N-terminal amino acids of the major E. coli Lpp′ joined to a transmembrane domain (residues 46 to 159) of the outer membrane protein of OmpA\[15\].

The purpose of this paper was to study the kinetics and equilibrium isotherm of the Cd (II) biosorption process. We optimized the uptake of Cd (II) by surface-engineered E. coli using the statistical design of experiments. In addition, the present study investigated the impact of desorption agents on the recovery of adsorbed Cd (II). Surface functional groups of bacterial cells involved in cadmium adsorption were also determined using Fourier transform infrared spectrophotometer analysis.

**MATERIALS AND METHODS**

**Microorganism and media**

The recombinant E. coli strain E, engineered with a cyanobacterial MT, SmtA, using the pET26b-Lpp′-OmpA expression vector was obtained from National Institute of Genetic Engineering and Biotechnology (Tehran, Iran)\[14\]. Luria Bertani (LB) was used as the growth medium and supplemented with kanamycin sulfate to a final concentration of 50 mg/mL. Isopropyl β-D-1-thiogalactopyranoside, as an inducer, was added to the culture medium when the cells reached an optical density of 0.6 at 600 nm. After induction period, the culture was incubated at 25°C for 5 h. The cells were subsequently harvested by centrifugation at 4000 xg and freeze dried until further use.

**Preparation of metal solutions**

Metal solutions were prepared by diluting a 1000 mg/L stock solution of Cd (NO₃)₂.4H₂O, with 0.1 M trisaminomethane-hydrochloric acid (Tris-HCl) to obtain concentrations between 10-110 mg/L. For each solution, the initial Cd (II) concentration and the concentration in the samples following the biosorption treatment process were determined using a flame atomic absorption spectrometer (Perkin Elmer Analyst, USA).

**Cadmium adsorption studies**

Batch adsorption experiments were conducted to study kinetic models, equilibrium isotherms, and the effect of different variables on cadmium adsorption, consisting of pH, temperature, and mass dosage. Each experiment was carried out in 100-mL Erlenmeyer flasks containing 10 mL of Cd (II) solution by shaking at 100 rpm. Then biomass was separated by centrifugation at 4000 xg and filtered through a Whatman filter paper with a pore size of 25 μm. Filtered samples were then analyzed for residual Cd (II) ion concentration using an analyst 700 atomic adsorption spectrometer (Perkin Elmer Analyst 700, USA). A control experiment was also carried out using the same solution and equipment, but in the absence of the biosorbent, E. coli E. Solute uptake by the recombinant E. coli E strain can be calculated from the
differences between the initial and final quantities of the solute contained in the supernatant as follows:

\[ Q = V \left( C_0 - C_f \right) / M \]  

(1)

where \( Q \) is the solute uptake (mg/g); \( C_0 \) and \( C_f \), the initial and equilibrium solute concentrations in solution (mg/L), respectively; \( V \), solution volume (L); \( M \), the mass of the biosorbent (g)\(^{16}\).

**Biosorption kinetics**

The sorption kinetics data provide valuable insights into the reaction pathways, the mechanism of the sorption reaction, and solute uptake\(^{16}\). The pseudo-first-order and pseudo-second-order biosorption models were applied to describe the kinetics of biosorption. The initial Cd (II) concentration was 20 mg/L in Tris-HCl buffer, pH 6.5. The sorption time varied between 5 and 100 min, and temperature was set at 30°C. At different times, each flask was removed from the shaker, and the biomass was centrifuged as mentioned above and then filtered. Finally, the solutions were analyzed to measure the residual Cd (II) concentration. The pseudo-first order model points that the rate of adsorption sites occupation is proportional to the number of unoccupied sites\(^{17}\). The linear equation for this model is:

\[ \log (q_e - q_t) = \log q_e - (K_r / 2.303) t \]  

(2)

where \( q_e \) and \( q_t \) are the amounts of metal ions adsorbed at equilibrium and at any time \( t \), respectively (nmol/mg) onto the biosorbent surface, and \( K_r \) is the rate constant of the first-order biosorption\(^{17}\). In the pseudo-second order model, it is assumed that the rate of the occupation of adsorption sites is proportional to the square of the number of unoccupied sites\(^{17}\). Linear equation for this model is:

\[ t/q_e = (1/K_s q_{aw}) + (1/q_{aw}) t \]  

(3)

where \( q_{aw} \) and \( q_e \) are the amounts of metal ions adsorbed on the biosorbent at equilibrium and at any time \( t \), respectively (nmol/mg), and \( K_s \) is the rate constant of second-order biosorption (mg/nmol min\(^{-1}\))\(^{17}\). The linear regression curve fitting procedure was performed with Microsoft Excel (version 7). The goodness of fit of the data to the model was evaluated by the coefficient of determination, \( R^2 \), by least-squares method\(^{17}\).

**Biosorption isotherms**

Equilibrium adsorption isotherms are usually used to determine the capacity, surface properties, and affinity of an adsorbent. Among all theoretical models, the Langmuir and Freundlich equilibrium models, the most widely used sorption isotherms, were chosen for the estimation of the adsorption capacity of \( E. coli \) E.

The linear Langmuir equation is written as follows\(^{18}\):

\[ C_e/q_e = (C_e/q_{max}) + (1/K_L q_{max}) \]  

(4)

where \( q_e \) is the equilibrium biosorption capacity of biomass in nmol Cd (II)/mg of biomass, \( C_e \) is the equilibrium concentration of Cd (II) ion in nmol/L, \( q_{max} \) is the maximum amount of metal sorbed in nmol Cd (II)/mg of biomass, and \( K_L \) is the constant that is referred to the bonding energy of sorption in nmol/L. Langmuir isotherm refers to homogeneous adsorption, in which each molecule possesses constant enthalpies and sorption activation energy (all sites possess equal affinity for the adsorbate), with no transmigration of the adsorbate in the plane of the surface\(^{19}\). The linear Freundlich equation is written as follows\(^{18}\):

\[ \log q_e = \log K_F + 1/n \log C_e \]  

(5)

where \( q_e \) is the equilibrium biosorption capacity of the biomass in nmol Cd (II)/mg biomass, \( C_e \) is the equilibrium concentration of Cd (II) ion in nmol/L, and \( K_F \) and \( 1/n \) are constants related to the sorption capacity and intensity, respectively. Freundlich isotherm is the earliest known relationship describing the non-ideal and reversible adsorption, not restricted to the formation of monolayer. This empirical model can be applied to multilayer adsorption, with non-uniform distribution of adsorption heat and affinities over the heterogeneous surface\(^{19}\).

Isotherm experiments were carried out at 30°C, using 10 mg dried biomass/10 mL of varying initial Cd (II) concentrations in the range of 10–110 mg Cd (II)/L of Tris-HCl buffer (pH 6.5), with constant shaking at 100 rpm and using an equilibrium time of 1 h. The linear regression curve fitting procedure was performed as mentioned above.

**Response surface methodology (RSM)**

One-factor-at-a-time, the classical method of experimental designs, involves changing one independent variable while maintaining all others at a fixed level. This method does not include the interactive effects among the variables. Experimental factorial designs can overcome this problem\(^{20}\). The RSM based on full or factorial design is a powerful tool for optimization, which have been employed extensively to optimize the biosorption of heavy metals\(^{21-23}\).

In order to obtain the optimum conditions for Cd (II) adsorption, three variables, including pH, temperature, and biomass dosage of solution were selected for the study. The range for these factors was chosen based on preliminary screening experiments according to cell
viability on LB agar. For this purpose, cell suspensions of the same weight were adjusted to different pH values using the following buffers (0.1 M): glycine-HCl (pH 3), sodium acetate (pH range of 4-5), Tris-HCl (pH range of 6-8), and glycine-NaOH (pH range of 9-10) and incubated at 30°C for 1 h with shaking at 100 rpm. In another test, cell suspensions of the same weight were incubated at different temperatures (20, 30, 40, 50, 60, 70, and 80°C) for 1 h at pH 7 with shaking at 100 rpm. Subsequently, equal volumes of the cell suspensions were spread onto LB agar plates (pH 7) and incubated at 37 °C overnight. Colony forming units were counted and compared to the condition with maximum colony forming units, at pH 7 and 30°C.

The central composite design (CCD) based on RSM was used to optimize the above mentioned factors. There are three types of CCD, which depend on where the axial points are placed. In the face-centered CCD type, the axial points are at the center of each face of the factorial space, so α=±1. The high and low values of the factors are coded as +1 and -1, respectively. The mean value of the factors was assigned to 0 as the central point. The experimental design for the three mentioned factors contained a total of 17 experiments, representing six axial points on cubic surfaces and 2 factorial points on vertices, as well as the central point with 3 replications (Table 1).

The optimum values of the selected variables were obtained by solving the regression equation and also by analyzing the response surface plots using the Design Expert software (version 7.0.0; Stat-Ease, Inc., USA). The initial Cd (II) concentration was 40 mg/L, and each experiment was carried out in duplicate, with the results being reported as mean values±standard deviation.

**FTIR analysis**

The FTIR study was intended to provide a deeper insight into the interaction between the surface functional groups of the biosorbent and the cadmium ions. The biomass was first dried prior and after adsorption using a lyophilizer (Denmark) and was grounded into fine particles using mortae and pestle. Each sample was then mixed with potassium bromide (1 mg in 100 mg of KBr), compressed into a 0.25-mm thickness disk and stabilized under controlled relative humidity before acquiring the spectrum. The FTIR spectrophotometer (Bruker-Vector22) used to record spectra was a Shimadzu IRPrestige in the wave number range of 400 to 4,000 cm⁻¹.

**Desorption**

Metals desorption is an important process in regeneration of the used adsorbents for their repeated use in water purification systems. It is also necessary to recover the precious metals. In this study, the binding sites of the biosorbent were first loaded with metal ions and desorption was then carried out to recover the adsorbed Cd (II). The cells were collected after adsorption under optimum conditions and washed with 0.1 M Tris-HCl. They were then incubated on ice for 15 min with 5 mM EDTA in 0.1 M Tris-HCl (pH 8.0) to remove the surface-bound metals. Alternatively, following adsorption and washing, the cells were incubated with 0.1 M CaCl₂ in 0.1 M Tris-HCl (pH 8.0) at 40°C for 1 h. A control sample was also incubated in

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**Table 1. Experimental design based on the central composite design used in this study**

<table>
<thead>
<tr>
<th>Trials</th>
<th>Point type</th>
<th>pH</th>
<th>Temperature (°C)</th>
<th>Biomass dosage (mg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Factorial</td>
<td>5.0</td>
<td>20</td>
<td>10</td>
</tr>
<tr>
<td>2</td>
<td>Factorial</td>
<td>8.0</td>
<td>20</td>
<td>10</td>
</tr>
<tr>
<td>3</td>
<td>Factorial</td>
<td>5.0</td>
<td>40</td>
<td>10</td>
</tr>
<tr>
<td>4</td>
<td>Factorial</td>
<td>8.0</td>
<td>40</td>
<td>10</td>
</tr>
<tr>
<td>5</td>
<td>Factorial</td>
<td>5.0</td>
<td>20</td>
<td>30</td>
</tr>
<tr>
<td>6</td>
<td>Factorial</td>
<td>8.0</td>
<td>20</td>
<td>30</td>
</tr>
<tr>
<td>7</td>
<td>Factorial</td>
<td>5.0</td>
<td>40</td>
<td>30</td>
</tr>
<tr>
<td>8</td>
<td>Factorial</td>
<td>8.0</td>
<td>40</td>
<td>30</td>
</tr>
<tr>
<td>9</td>
<td>Axial</td>
<td>5.0</td>
<td>30</td>
<td>20</td>
</tr>
<tr>
<td>10</td>
<td>Axial</td>
<td>8.0</td>
<td>30</td>
<td>20</td>
</tr>
<tr>
<td>11</td>
<td>Axial</td>
<td>6.5</td>
<td>20</td>
<td>20</td>
</tr>
<tr>
<td>12</td>
<td>Axial</td>
<td>6.5</td>
<td>40</td>
<td>20</td>
</tr>
<tr>
<td>13</td>
<td>Axial</td>
<td>6.5</td>
<td>30</td>
<td>10</td>
</tr>
<tr>
<td>14</td>
<td>Axial</td>
<td>6.5</td>
<td>30</td>
<td>30</td>
</tr>
<tr>
<td>15</td>
<td>Central</td>
<td>6.5</td>
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<td>20</td>
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<tr>
<td>16</td>
<td>Central</td>
<td>6.5</td>
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<td>20</td>
</tr>
<tr>
<td>17</td>
<td>Central</td>
<td>6.5</td>
<td>30</td>
<td>20</td>
</tr>
</tbody>
</table>
Tris-HCl (pH 8.0) at 40°C for 1 h. The supernatants resulting from these treatments were then subjected to atomic absorption analysis.

**RESULTS**

**Kinetic studies**

Figure 1 represents the effects of contact times (5 to 100 min) on the biosorption of Cd (II) by E. coli E. A fast rate of Cd (II) adsorption was observed in the first 5 min. The maximum removal of Cd (II) occurred after 40 min, when the uptake of Cd (II) was approximately 74.72%.

Biosorption kinetics was studied to understand the adsorption dynamics of Cd (II) onto the E. coli E surface. The models obtained in this study allowed to estimate the amount of Cd (II) adsorbed at the time of processing. Accordingly, two types of kinetic models were applied, the pseudo-first-order and pseudo-second-order.

The pseudo-first-order kinetic model is the plot of Log (qe–qf) versus time. Table 2 shows the values of the biosorption rate constant, K1, calculated qe, and experimental qe. The coefficient of determination (R²=0.75) showed that linear regression did not fit the experimental data (Table 2) and could not predict qe accurately. Therefore, the pseudo-second-order kinetic model was used to analyze the biosorption kinetics of Cd (II). The pseudo-second-order kinetic model is the plot of t/qf versus time. As shown in Table 2, according to the R² of 0.99, the pseudo-second-order model could satisfactorily fit the experimental data, where the calculated qe was in acceptable agreement with the experimental data.

**Isotherm**

The biosorption capacity of E. coli E was increased with the initial concentration of Cd (II) ions in solution (Fig. 2), demonstrating the potential of E. coli E, as a biosorbent, to treat wastewater containing high concentrations of metal ions.

The biosorption process will stop only when equilibrium is reached between the amount of metal ions adsorbed onto the biosorbent and that of solution. The isotherm models were thus investigated at the time of equilibrium. The parameters of equilibrium sorption were used to investigate the surface properties and affinity of the sorbent[24]. The Langmuir and Freundlich isotherm models were employed for this purpose.

**Table 2.** The biosorption rate constants and the qe values from the pseudo-first-order and pseudo-second-order kinetics for the biosorption of Cd (II) by E. coli E

<table>
<thead>
<tr>
<th>Metal ion</th>
<th>Expt. Q</th>
<th>Pseudo-first-order kinetics</th>
<th>Pseudo-second-order kinetics</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>R²</td>
<td>Cal. Qe</td>
</tr>
<tr>
<td>Cd (II)</td>
<td>133 ± 3.53</td>
<td>0.75</td>
<td>16.34</td>
</tr>
</tbody>
</table>

Expt., experimental data of adsorbed metal; Cal., data of adsorbed metal calculated from the model

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Fig. 1. Effect of contact time on the biosorption capacity of Cd (II) (qₐ) by E. coli E. [Cd (II)] concentration, 20 mg/L; biomass dosage, 1 mg/mL Tris-HCl, pH 6.5, 30°C, 100 rpm.
The Langmuir adsorption model was applied for the experimental data obtained. A graph of $C_i/q_e$ versus $C_i$ is a linear plot of the Langmuir model. The value of the biosorption capacity, $q_{\text{max}}$, and the Langmuir constant, $K_L$, were obtained from linear regression. The value of $R^2$ for Cd (II) was found to be 0.97. Although the Langmuir model fits the experimental data for Cd (II), it was preferred to study the model fitting for the Freundlich model according to the $R^2$ value of 0.97. The linear Freundlich plot can be obtained by the Log $q_e$ (nmol/mg) versus Log $C_i$ (nmol/mL). According to $R^2$ value of 0.98, the Freundlich isotherm was in good agreement with the experimental data in this study. The value of the $1/n$ was obtained as 0.32, indicating there is a chemical interaction between Cd (II) and E. coli E. The adsorption constants estimated from the Freundlich and Langmuir isotherms are summarized in Table 3.

### Optimization of the biosorption process

The range of variables used in the optimization process was determined according to screening of variables based on cell viability. The t-test indicated a significant difference between pH values 4 and 7, as well as 9 and 7; however, no significant differences were found between pH values 5 and 7, 6 and 7, and 8 and 7 ($P>0.05$). Also, it demonstrated a significant difference between the temperatures 50, 60, 70, 80, and 30°C, but no significant difference was detected between temperatures 20, 30, and 40°C ($P>0.05$).

Consequently, the range of variables chosen to analyze the optimum conditions provided maximum biosorption efficiency using the face-centered CCD, as mentioned in Table 1. The Cd (II) uptake rate ($q_e$) was measured as the response, and then the results were compared with the predicted values (Table 4). Analysis of variance (ANOVA) showed that there is a statistical significance in the quadratic model. The $F$ and $P$ values of the regression model were 91.42 and <0.0001, respectively, implying the significance of the model. $R^2$ was 0.98, which indicates that only 0.28% of the total variable could not be explained by the model. The lack of fit value of 0.05 implied that the lack of fit is not significant relative to the pure error. Both parameters showed that the model could well fit the experimental data. The value of the adjusted $R^2$ of 0.96 was in reasonable agreement with the predicted $R^2$ of 0.93. In addition, a relatively low value of the coefficient of variation (CV=6.46) indicates the repeatability of the experiments.

Equation indicates the final mathematical model corresponding to the coded factors after eliminating the insignificant terms ($P>0.05$), as determined by the Design-Expert software. (6):

$$Y=146.16+19.18 A+8.70 B-63.58 C-12.73 AC+21.58 C^2$$

(6)

### Table 3. Langmuir and Freundlich constants for Cd (II) biosorption by E. coli E

<table>
<thead>
<tr>
<th></th>
<th>Langmuir</th>
<th></th>
<th>Freundlich</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>$K_L$</td>
<td>0.01</td>
<td>277.77</td>
<td>0.97</td>
<td>30.535</td>
</tr>
</tbody>
</table>

**Fig. 2.** Biosorption of Cd (II) at various initial concentrations (pH 6.5, 1 mg/mL biomass, 30°C).
The effect of temperature and pH on biosorption was investigated. Our results indicated that metal uptake was decreased when the amount of biomass was increased from 1 to 3 mg/mL, and the pH was decreased from 8 to 5. Metal uptake, however, was at a maximum when the biomass concentration and pH value reached 1 mg/mL and of 8, respectively. Therefore, increased biomass dosages and decreased pH values simultaneously led to a reduction in the Cd (II) ion uptake.

**Interactive effect of temperature and pH on biosorption**

Figure 3B represents the effect of temperature and pH in the solution on Cd (II) uptake. Our results demonstrated the decreased metal uptake after the temperature and pH were lowered from 40 to 20°C and 8 to 5, respectively. The lowest uptake was observed at a temperature of 20°C and pH value of 5.

**Interactive effect of biomass dosage and temperature on biosorption**

The effect of biomass dosage and temperature on Cd (II) uptake is shown in Figure 3C. It was observed that the metal uptake was decreased when the amount of biomass was increased from 1 to 3 mg/mL, and the temperature was lowered from 40 to 20°C, demonstrating the highest reduction in metal uptake at the biomass value of 1 mg/mL and a temperature of 40°C.

Results from ANOVA showed that temperature has a significant effect on adsorption (P=0.0001). However, the F value of this factor (7.20) implied that this variable is not highly significant when compared with the biomass dosage (F=384.36). Hence, unlike the biomass dosage, a color range (blue to red) associated with temperature was not present in the response surface plot (Fig. 3C). A study, carried out by Hassan et al.\(^{25}\) in 2009 showed that temperatures in a range from 25 to 55°C have no remarkable effects on the copper biosorption by the brown seaweed *Sargassum* sp.

**Effect of the optimization of different variables**

According to the results shown in Table 4, optimization of different variables led to increased cadmium adsorption from 94.84±2.83 to 284.70±3.54 nmol/mg. This result was compared with the other biosorbents retrieved from selected literatures (Table 5).

**FTIR analysis results**

FTIR analysis allows the identification of the functional groups involved in cadmium bioadsorption by shifting the changes in signal intensity compared to the control. FTIR analysis was carried out on our developed bioadsorbent (*E. coli* strain E) in the absence and presence of cadmium to determine the differences that are due to interaction of the metal ions with the surface functional groups (Fig. 4). Our results showed that cadmium ions bind to the functional groups, including carboxyl, amino, phosphoryl, thiol, and hydroxyl groups, which results in the shift and changes in the spectra of the bioadsorbent before and after cadmium adsorption.

The results attributable to the presence of the specific bands is summerized in Table 6. As shown in Figure 4, a weak –SH stretching was observed in the range of 2500-2600 cm\(^{-1}\). It has long been known that the SH stretch(s) mode, v (SH), is generally found in the range of 2500-2600 cm\(^{-1}\), being weak in the IR\(^{26}\). The peaks at 2937.2, 1433, 1373 and 922 cm\(^{-1}\) might be due to CH\(_2\) stretching and bending vibrations, while the peaks at 922 and 913 cm\(^{-1}\) might be because of P-OR stretching. These groups could be attributed to polysaccharides, phospholipids, or ribose phosphate chain pyrophosphate, which are the main components of the bacterial cell wall\(^{27}\). The band observed at 1543.5 cm\(^{-1}\) was assigned to amide of proteins and peptides.
Desorption of Cd (II)
Because of the sensitivity of *E. coli* E to such conditions, no alkaline and acidic elutants were used for Cd (II) desorption. The recovery experiments showed that approximately 68.58% and 56.54% of the Cd (II) ions were recovered when 5 mM EDTA and 100 mM CaCl₂ were used, respectively.

**DISCUSSION**

In this study, the bioadsorbent developed in our previous study was evaluated by kinetic and isotherm studies, followed by optimization of adsorption conditions. Study on contact times showed that cadmium adsorption was initially rapid, presumably due to more active-binding sites present on the adsorbent in the beginning. However, the rate of cadmium adsorption became slower after some time, probably because of a lesser number of active-binding sites available on the adsorbent[28].

The first-order kinetic process was used for reversible reactions, in which equilibrium was established between liquid and solid phases. However, the pseudo-second order kinetic model assumes the chemical adsorption, as a probable rate-limiting step[29]. Bacterial cell walls contain several functional groups, including carboxyl, phosphonate, amine, and hydroxyl groups[30]. It is known that the functional groups present on the negatively-charged bacterial cell wall participate in the binding of metal cations, such as Cd (II)[30]. In addition, the cyanobacterial MT, SmtA, used in this adsorbent as a metal-binding component of this bioadsorbent contains a Zn₄Cys₉His₂ cluster structure that can replace Cd (II) ions in the coordination bonds[31,32]. This result might be the main reason why the Cd (II) biosorption reaction by *E. coli* E has been well-modeled by the pseudo-second order kinetics. A number of studies have shown that the pseudo-second order mechanism is a better model for explaining the kinetics of divalent metal sorption onto heterogeneous sorbents[28,33].

According to Giles et al.[34], the plateau or the beginning of the linear portion above the "knee" must show "first-degree saturation" of the surface (Fig. 2); this observation conforms the conditions that all

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Fig. 3. Response surface plot of different factors interactions. (A) The interactive effect of the biomass dosage and pH at constant temperature 30ºC on metal uptake (q); (B) the effect of temperature and pH at constant biomass dosage of 2 mg/mL on the metal uptake (q); (C) the effect of temperature and biomass dosage at constant pH of 6.5 on metal uptake (q).
Table 5. Comparison between cadmium adsorption of selected literatures and this work

<table>
<thead>
<tr>
<th>Biosorbent</th>
<th>Operating condition</th>
<th>pH</th>
<th>Temp (°C)</th>
<th>Biomass (g/L)</th>
<th>Time (h)</th>
<th>Amount adsorbed (nmol/mg)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bacillus circulans</td>
<td></td>
<td>7</td>
<td>20</td>
<td>0.5</td>
<td>2</td>
<td>235.764</td>
<td>[37]</td>
</tr>
<tr>
<td>Pseudomonas putida</td>
<td></td>
<td>6</td>
<td>30</td>
<td>Not available</td>
<td>1</td>
<td>71</td>
<td>[38]</td>
</tr>
<tr>
<td>Pseudomonas stutzeri</td>
<td></td>
<td>5</td>
<td>30</td>
<td>1</td>
<td>30 min</td>
<td>387</td>
<td>[25]</td>
</tr>
<tr>
<td>KCCM 34719</td>
<td></td>
<td>5</td>
<td>28+2</td>
<td>1</td>
<td>&gt;5</td>
<td>284.252</td>
<td>[27]</td>
</tr>
<tr>
<td>Bacillus cereus RC-1</td>
<td></td>
<td>Not</td>
<td>37</td>
<td>3.33</td>
<td>2</td>
<td>510</td>
<td></td>
</tr>
<tr>
<td>Engineered E.coli</td>
<td></td>
<td>8</td>
<td>40</td>
<td>0.01</td>
<td>1</td>
<td>284.697</td>
<td>this study</td>
</tr>
<tr>
<td>Engineered E.coli</td>
<td></td>
<td>8</td>
<td>40</td>
<td>0.01</td>
<td>1</td>
<td>284.697</td>
<td>this study</td>
</tr>
</tbody>
</table>

possible sites in the original surface are filled, and further adsorption would take place only on the new surfaces. For convenience, this degree of coverage may be called the formation of a complete "monolayer," but this does not necessarily mean that it is a close-packed layer of single molecules or ions, as in a compressed monolayer on water[34]. Based on the R² value, the experimental data was in good agreement with the Freundlich isotherm. It can be due to the presence of different functional groups on the E. coli E surface, especially, SmtA that produces a heterogeneous biosorbent. The Freundlich isotherm can fit various experimental adsorption data and has been found to be in line with data from the highly heterogeneous sorbent systems. For example, Gong et al. [35] have reported that the process of lead biosorption by Spirulina maxima, a filamentous cyanobacterium that can be used as a food supplement, follows the Freundlich isotherm model.

In optimization process, the interactive effect of biomass dosage and pH showed that increased biomass dosages and decreased pH values simultaneously lead to a reduction in Cd (II) ion uptake. This phenomenon can be attributed to cadmium precipitation at higher pH[36-38] or the reconstitution of SmtA molecules with cadmium ions, at higher pH values[32]. Masoudzadeh et al.[39] have stated that it might also be due to the fact that the available solute was insufficient to completely cover the available exchangeable sites on the cell surface, thereby resulting in low solute uptake at high biomass concentration. Similar observations have been made in study on Cd (II) biosorption using pretreated Saccharomyces cerevisiae biomass[22]. Meanwhile, biomass aggregation could interfere with the surface metal-binding sites. At high biomass dosages, the available binding sites are insufficient as they could be masked due to the surface protein interactions and limited metal accessibility[40]. It is likely that protons will then combine with the metal ions, thereby decreasing the interaction of the metal ions with the cell components[41].

The study of interaction between temperature and pH ranges on biosorption showed Cd (II) adsorption at a higher pH value. This finding can be related to the reaction between the binding sites of the sorbent and the metal, leading to the breaking of hydrogen bonds and the release of hydrogen ions, which are ultimately substituted by the metal ions[42,43]. In the process of
Table 6. The FTIR Spectral Characteristics of Biosorbent before and after Biosorption of Cd

<table>
<thead>
<tr>
<th>Wavelength range (cm⁻¹)</th>
<th>Biosorbent after biosorption</th>
<th>Assignment</th>
<th>Functional class</th>
</tr>
</thead>
<tbody>
<tr>
<td>3550-3600 3200-3550</td>
<td>3562.4</td>
<td>O–H</td>
<td>Oxime (=NOH)</td>
</tr>
<tr>
<td>3200-3550</td>
<td>3338.1</td>
<td>O–H or N–H</td>
<td>Alcohols, phenols or secondary amines</td>
</tr>
<tr>
<td>2500-3300</td>
<td>3389.5</td>
<td>O–O–H or N–H</td>
<td>Alcohols, phenols or secondary amines</td>
</tr>
<tr>
<td>2600-2550</td>
<td>2937.2</td>
<td>C–H or O–H</td>
<td>Alkanes(methylene) or carboxylic acids</td>
</tr>
<tr>
<td>Very weak</td>
<td></td>
<td>S–H</td>
<td>Thiols</td>
</tr>
<tr>
<td>1630-1680</td>
<td>1655.3</td>
<td>C=C or C=N</td>
<td>Alkenes or oxime (=NOH)</td>
</tr>
<tr>
<td>1500-1560</td>
<td>1543.5</td>
<td>N=O or N–H</td>
<td>Nitro compounds or amides</td>
</tr>
<tr>
<td>1350-1470</td>
<td>1433.0</td>
<td>C–H or O–H</td>
<td>Alkanes and aldehyde or carboxylic acids</td>
</tr>
<tr>
<td>1330-1430</td>
<td>1373.1</td>
<td>C–H or O–H</td>
<td>Alkanes or alcohols, phenols</td>
</tr>
<tr>
<td>1325± 25</td>
<td>1346.7</td>
<td>S=O</td>
<td>Sulfonic acid or sulfone</td>
</tr>
<tr>
<td>1210-1320</td>
<td>1279.1</td>
<td>C–O or N–O</td>
<td>Carboxylic acids or aromatic amine oxide</td>
</tr>
<tr>
<td>1210-1320</td>
<td>1238.4</td>
<td>C–O or P=O</td>
<td>Carboxylic acids or phosphonate and phosphoramides</td>
</tr>
<tr>
<td>1100-1200</td>
<td>1125.2</td>
<td>P=O</td>
<td>Phosphine oxide and phosphate</td>
</tr>
<tr>
<td>1050-1200</td>
<td>1068.0</td>
<td>C=S</td>
<td>Thiocarboxyl</td>
</tr>
<tr>
<td>880-1050</td>
<td>992.6</td>
<td>P–OR or ±C–H &amp; ±CH₂</td>
<td>Esters or alkenes</td>
</tr>
<tr>
<td>900-1050</td>
<td>913.2</td>
<td>P–OR (P–O–C)</td>
<td>(Phosphite) esters</td>
</tr>
<tr>
<td>600-900</td>
<td>864.2</td>
<td>C–H</td>
<td>Alkenes or amines</td>
</tr>
<tr>
<td>600-900</td>
<td>683.6</td>
<td>C–H or O–H</td>
<td>cis-RCH=CHR alkenes and amines or or NH₂ and N-H alcohols, phenols or amines</td>
</tr>
<tr>
<td>500-700</td>
<td>583.4</td>
<td>C–Br and C–Cl</td>
<td>Bromoalkanes and chloroalkanes</td>
</tr>
<tr>
<td>500-600</td>
<td>551.5</td>
<td>C–Br and C–Cl</td>
<td>Bromoalkanes and chloroalkanes</td>
</tr>
</tbody>
</table>

physicosorption, weak adsorption interactions between surface and the metal ions was decreased with increased temperature\(^{[44]}\). However, based on the pseudo-second-order kinetic model, the interaction between E. coli E and Cd (II) ions can be ascribed to chemosorption. Therefore, temperature has a positive effect on the metal uptake capacity of E. coli E over the tested temperature ranges of 20-40°C. Indeed, the results from this study, regarding the effect of temperature on Cd\(^{2+}\) adsorption, are consistent with those observed by Dang et al.\(^{[45]}\). On the other hand, the study of interaction between effects of biomass dosage and temperature on biosorption showed that although raising the biosorbtion dosage caused an increase in the biosorbent surface area and the availability of more adsorption sites, the uptake capacity \((q_u)\) was decreased under such a condition. This result can be attributed to the unsaturated sites during the adsorption reaction, whereas the number of the sites available for the adsorption increases by raising the adsorbent dosage.

FTIR analysis was carried out to determine the functional groups involved in the cadmium biosorption. The significant changes in the appearance of new peak positions at specific wavelengths suggested that different compounds such as polysaccharides, phospholipids, proteins, and peptides were involved in the cadmium adsorption on the cell surface. A weak –SH stretching on FTIR spectra could be assigned to the molecules including thiol groups present in the cystein residues in MT on the surface of our developed biosorbent. According to the Hard-Soft Acid-Base theory, sulfur is a relatively soft (polarizable) atom. This also explains the tendency of thiol groups to bind to the soft elements/ions such as mercury, lead, or cadmium.

Our experiment regarding cadmium desorption using EDTA and CaCl\(_2\) showed that this kind of adsorption could be due to electrostatic and coordination interactions occurring between Cd (II) ions and the E. coli E cell surface. The presence of competitor ions, such as calcium, at high concentrations, can release the Cd (II) ions involved in the electrostatic interactions. In addition, because of the affinity of heavy metals for EDTA, as a chelating agent, it could release the Cd (II) ions more efficiently than CaCl\(_2\). This result is consistent with that of Hua et al.\(^{[46]}\) who indicated that the presence of EDTA generally decreased the adsorption of Cd to biofilm in natural waters.

Our results demonstrated that the presence of the different functional groups and MT present on the surface of the E. coli E caused an increased chemical adsorption on heterogenous surface. Furthermore, the surface adsorption was remarkably increased while the critical adsorption parameters were optimized. It can be concluded that surface engineering of the
surface proteins involving metal adsorption as well as optimization of operational parameters have a great impact on the efficiency of biosorption process.

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CONFLICT OF INTEREST. None declared.

REFERENCES


