FULL LENGTH

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Comparison of Biofilm Formation between Methicillin-Resistant and Methicillin-Susceptible Isolates of *Staphylococcus aureus*

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ABSTRACT

**Background:** The aim of this study was to compare the biofilm formation and the prevalence of biofilm-associated genes between the isolates of methicillin-resistant (MRSA) and methicillin-susceptible (MSSA) *Staphylococcus aureus*. **Methods:** In total, 209 *S. aureus* isolates were collected. The antibiotic susceptibility test was conducted using nine antibiotics according to the guidelines of Clinical and Laboratory Standards Institute. Phenotypic biofilm formation was performed with microtiter plate assay. The polymerase chain reaction was employed to detect icaA, icaD, icaB, icaC, clfA, clfB, fnbA, fnbB, fib, cna, eno, ebpS, ebpB, mecA, and SCCmec types as well as agr group genes with specific primers. **Results:** Sixty-four (30.62%) isolates were resistant to methicillin, and 54 (83%) MRSA harbored SCCmec III. Furthermore, 122 (58.3%) isolates belonged to agr group I. Twenty-six (36.1%) MRSA and 42 (28.9%) MSSA isolates were strong biofilm producers (no significant difference). The prevalence of icaA, icaD, icaB, and icaC genes in MSSA isolates was 71, 41, 76, and 72%, respectively. The frequency of clfA, clfB, fnbA, fnbB, fib, cna, eno, ebpB, and ebpS in MSSA was 100, 100, 56, 46, 74, 54, 78, 11, and 1%, respectively. However, in MRSA isolates, the frequency was 97, 97, 64, 51, 76, 56, 79, and 12% with no track of ebpS, respectively. **Conclusion:** Statistical difference between MSSA and MRSA regarding biofilm formation and the frequency of all biofilm-encoding genes was not significant. The majority of the *S. aureus* isolates harbored clfA, clfB, eno, fib, icaA, and icaD genes. DOI: 10.7508/ibj.2016.03.007

**Keywords:** Biofilm, Methicillin-resistant *Staphylococcus aureus*, Methicillin-susceptible *Staphylococcus aureus*

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INTRODUCTION

*Staphylococcus aureus* is one of the most nosocomial pathogens. Methicillin-resistant *S. aureus* (MRSA) strains, which have been developed for four decades, resist a wide spectrum of antibiotics. Biofilm formation in *S. aureus* isolates occurs through a polysaccharide intercellular adhesion (PIA) and also through microbial surface components recognizing adhesive matrix molecules (MSCRAMMs)⁴⁻⁶. These structures mediate the *S. aureus* initial attachment to both host tissues and biomaterials.⁵ Biofilm formation interferes with bacterial recognition and killing mechanisms of the innate immune system⁶. MSCRAMMs play a key role in initiation of endovascular, bone and joint and prosthetic device infections⁶. Various *S. aureus* strains may not have a similar profile in the prevalent constellations of MSCRAMMs and also can make the individuals predispose to certain kinds of infections through binding to molecules such as collagen, fibronectin and fibrinogen⁷,8. *S. aureus* can express up to 20 different adhesive MSCRAMMs that are covalently anchored by sortase to peptidoglycan via the C-terminal LPXTG motif⁶. These adhesion proteins include the ClfA and ClfB (clumping factors A and B),
FnbA and FnbB (fibronectin-binding proteins A and B), Fib (fibrinogen-binding protein), Cna (collagen-binding protein), Eno (laminin-binding protein), Ebp (elastin-binding protein), and Bbp (bone sialoprotein-binding protein). The initial site of attachment is in the moist squamous epithelium of the anterior nares of the host[9]. ClfA is the major staphylococcal fibrinogen binding-protein and is responsible for the observed clumping of S. aureus in blood plasma and culminating in arthritis and endocarditis[10]. ClfA also binds to platelet αIIbβ3 integrin[7]. On the other hand, ClfB binds to human cytokeratin 10 and to fibrinogen, as a bi-functional protein and thus mediates the nasal colonization and acts as a key virulence factor, which leads to metastatic infection and/or development of sepsis[7]. Both ClfA and ClfB interact with and inhibit complement C3. FnbA and FnbB are also involved in bacterial invasion of the endothelial cells in vivo and in vitro through RGD motif and then mediate the induction of cell signaling and reorganization of the actin cytoskeleton[10]. The FnBPA adhesions interact with receptors on endothelial cells and result in cardiovascular disease and cardiac device infections via platelet activation, a key step in thrombus formation and attachment to implanted prosthetic materials, respectively[7]. The cna gene encodes Cna protein and mediates the adhesion of S. aureus to collagenous tissues and cartilage[11]. It has been reported that the acquisition of antibiotic resistance does not change the capacity of biofilm formation in MRSA strains[8]. The aim of this study was to compare the prevalence of biofilm-related genes and their ability in biofilm formation between MRSA and MSSA isolates.

MATERIALS AND METHODS

Bacterial isolates

A total of 209 S. aureus clinical isolates were collected from patients in different hospitals from July 2012 to January 2013. Figure 1 shows the major sites of infection, including trachea, blood, wound, bronchus, sputum, and soft tissue. The isolates were identified using biochemical tests, such as mannitol fermentation on mannitol salt agar (Merck, Germany) medium, slide and tube coagulate tests, DNase production and colony morphology on blood agar medium.

Antibiotic susceptibility test

Antimicrobial susceptibility test (AST) was performed according to the guidelines of Clinical and Laboratory Standards Institute[12]. The S. aureus strain of ATCC25923 was prepared to control the quality of the antibiotic susceptibility test. Different disks were used in AST, including oxacillin (1 µg), erythromycin (15 µg), clindamycin (2 µg), vancomycin (2 µg), linezolid (30 µg), tetracycline (30 µg), trimethoprim-sulfamethoxazole (25 µg), gentamicin (10 µg), amoxicillin (10 µg) and ciprofloxacin (5 µg) (all purchased from MAST, UK).

Microtiter tissue plate (MTP) method

The MTP method was conducted as previously described[10]. In brief, the wells of a microtiter plate were inoculated with 180 µl trypticase soy broth supplemented with 1% glucose. Each bacterial culture (20 µl) with a turbidity equivalent to an 0.5 McFarland standard was added to each well of polystyrene, 96-well, sterile, flat-bottomed tissue culture plates. After 24-h incubation at 35°C, the wells were decanted and washed three times with sterile saline phosphate buffer. Next, methanol (for 20 min), and safranin 0.1% (for 15 min) were added to the wells. The stained wells were washed and left to ambient temperature to be dried. The safranin dye bound to the adherent cells was dissolved in 1 mL 95% ethanol per well, and the optical densities (ODs) of the plates were observed at 490 nm (A490) by using a microtiter-plate reader. Each assay was performed in triplicate. As a negative control, trypticase soy broth medium was used to determine background OD. OD cut-off was then determined as an average OD of negative control + 3x standard deviation (SD) of negative control. The OD cut-off value was separately calculated for each microtiter plate. Biofilm formation by isolates was calculated and categorized according to the absorbance of the safranin-stained attached cells (Table 1)[10].

Table 1. Classification of biofilm formation abilities by MTP assay[10]

<table>
<thead>
<tr>
<th>Cut-off value calculation</th>
<th>Biofilm formation abilities</th>
</tr>
</thead>
<tbody>
<tr>
<td>OD&gt;4×ODc</td>
<td>Strong</td>
</tr>
<tr>
<td>2×ODc&lt;OD≤4×ODc</td>
<td>Moderate</td>
</tr>
<tr>
<td>OD=ODc</td>
<td>Weak</td>
</tr>
<tr>
<td>OD≤2×ODc</td>
<td>Negative</td>
</tr>
</tbody>
</table>

Fig. 1. Different sampling sites for collection of the isolates.

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**RESULTS**

**Antibiotic susceptibility testing**

All the isolates were susceptible to vancomycin and linezolid. The antibiotic susceptibility pattern of the isolates for amoxicillin, erythromycin, tetracycline, gentamicin, clindamycin, ciprofloxacin, and trimethoprim-sulfamethoxazole was as follows: 90% (n=189), 29% (n=61), 25% (n=52), 19% (n=40), 19% (n=40), 15.3% (n=32), 11% (n=23), respectively.

**Detection of methicillin-resistant Staphylococcus aureus strains**

The phenotypic test indicated that 64 (30.62%) isolates were resistant to methicillin. The mecA gene was also detected in these isolates with specific primers. The MRSA isolates were significantly more resistant than MSSA isolates to the majority of the antibiotics (P≤0.05, Table 2), except for vancomycin and linezolid (Fig. 2).

**Phenotypic biofilm production**

The MTP assay demonstrated that 14 (21.8%) MRSA and 42 (28.9%) MSSA isolates were strong biofilm producers (no significant difference). Furthermore, approximately 50% of the total isolates showed a moderate level of biofilm formation.

**SCCmec types**

The majority of the MRSA isolates (n=54, 84%) harbored SCCmec III. However, 30 isolates with SCCmec type III were only susceptible to vancomycin and linezolid. The SCCmec types V and I were detected in 9% (n=6) and 6% (n=4) of the isolates, respectively.

**agr genes**

In total, 122 (58.3%) isolates belonged to agr group I, followed by agr group II (n=46, 22%), agr group IV (n=27, 13%) and agr group III (n=9, 4%). There was no relationship between agr specific groups and clinical signs (P=0.21).

**Frequency of icaA, icaD, icaB and icaC genes**

The ica genes were identified by PCR method. The size of PCR product for icaA, icaD, icaB, and icaC genes were 188, 198, 900, and 1100 bp, respectively. The frequency of the icaA, icaD, icaB, and icaC genes in MSSA isolates was 71%, 54%, 69%, and 71%, respectively. In the MRSA isolates, the frequency of the icaA, icaD, icaB, and icaC genes was 75%, 52%, 65%, and 65%, respectively.

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**Table 2.** Comparison between MSSA and MRSA isolates regarding antibiotic resistance percentage and the prevalence of agr group I

<table>
<thead>
<tr>
<th>Isolates</th>
<th>AMX</th>
<th>CD</th>
<th>E</th>
<th>T</th>
<th>SXT</th>
<th>GM</th>
<th>CIP</th>
<th>agr I</th>
</tr>
</thead>
<tbody>
<tr>
<td>MSSA</td>
<td>86.66</td>
<td>6.66</td>
<td>11.11</td>
<td>31.11</td>
<td>4.44</td>
<td>4.44</td>
<td>11.11</td>
<td>56%</td>
</tr>
<tr>
<td>MRSA</td>
<td>90.00</td>
<td>56.66</td>
<td>77.00</td>
<td>66.66</td>
<td>23.23</td>
<td>56.66</td>
<td>76.66</td>
<td>58%</td>
</tr>
<tr>
<td>P</td>
<td>0.57</td>
<td>0.001</td>
<td>0.001</td>
<td>0.03</td>
<td>0.02</td>
<td>0.001</td>
<td>0.001</td>
<td>0.52</td>
</tr>
</tbody>
</table>

AMX, amoxicillin; CD, clindamycin; E, erythromycin; T, tetracycline; SXT, trimethoprim-sulfamethoxazole; GM, gentamicin; CIP, ciprofloxacin

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**Fig. 2.** The antibiotic resistance pattern of MRSA and MSSA isolates. AMX, amoxicillin; T, tetracycline; E, erythromycin; CD, clindamycin, GM, gentamicin; SXT, trimethoprim-sulfamethoxazole; CIP, ciprofloxacin; LZD, linezolid; VAN, vancomycin.
frequency of these genes was 76%, 69%, 64%, and 74%, respectively. There was no significant difference between MRSA and MSSA regarding the presence of these genes. Also, there was no relation between icaA, icaD, icaB, icaC genes and agr groups or MRSA. The difference in the frequency of icaA, icaD, icaB, and icaC genes between MRSA and MSSA is shown in Figure 3.

Prevalence of genes encoding microbial surface components recognizing adhesive matrix molecules

Biofilm-related genes were amplified by two multiplex PCR panels (Figs. 4 and 5). The frequency of clfA, clfB, fnbA, fnbB, fib, cna, eno, ebps, and bbp in MRSA isolates was 97%, 97%, 64%, 51%, 76%, 56%, 79%, and 12% with no track of bbp, respectively. However, in MRSA isolates, the frequency was 100%, 100%, 56%, 46%, 74%, 54%, 78%, 11%, and 1%, respectively (Fig. 6). The statistical difference between MSSA and MRSA regarding the frequency of all the biofilm-encoding genes was not significant (Table 3). There was no relationship between these genes and agr groups as depicted in Table 4.

**DISCUSSION**

All of the isolates in the present study were susceptible to vancomycin and linezolid and likewise, the majority was susceptible to trimethoprim-sulfamethoxazole. Generally, vancomycin and linezolid are completely effective in MRSA treatment; however, reduced susceptibility to both antibiotics have been reported in some studies. Vancomycin and other glycopeptides have remained the last resorts for eradication of *S. aureus* infections.

The results from the current study indicated that the antibiotic susceptibility pattern of the isolated pathogen originated from different infected areas (trachea, blood, wound etc.) was not significantly meaningful. Also, 64 isolates were MRSA, and the majority of MRSA harbored the SCCmec type III. Previous reports have also indicated that the SCCmec type III is the predominant type in Iran. Based on our findings, the majority of the isolates (both MRSA and MSSA) belonged to *agr I* (58.3%), followed by the *agr II*, *agr IV*, and the *agr III*. Previous studies have also depicted that the *agr I* is the predominant type in Iran. The relationship between *agr I* and several characteristics, such as the AST pattern, the prevalence of biofilm-related genes, and biofilm formation exhibited that the isolates belonged to *agr I* had higher antibiotic resistance compared to those with other *agr* specific groups. In the phenotypic biofilm formation, the MSSA and MRSA isolates produced biofilm, and there was no significant difference (P<0.05).

The present study indicated that ClfA and ClfB were present in more than 95% of the isolates and constituted the bound coagulase of *S. aureus*. Similar to the current study, previous surveys have determined a high prevalence of icaA and icaD genes with a relationship to phenotypic biofilm formation. For instance, Nast et al. detected the icaA and icaD genes in (34%) of catheter and blood isolates that were capable of biofilm formation. This study demonstrated that there was a relationship between the biofilm formation and the presence of these genes. The difference between MRSA and MSSA was not significant regarding the presence of these genes. This result emphasizes that the SCCmec genes are separate and independent of ica locus. A comparative analysis between these isolates in the present study would be very useful to obtain more information about the biofilm properties of *S. aureus* isolates.
Biofilm Formation in Staphylococcus aureus Isolates

Table 3. The comparison of biofilm-related genes percentage between MRSA and MSSA isolates

<table>
<thead>
<tr>
<th>Isolates</th>
<th>icaA</th>
<th>icaB</th>
<th>icaC</th>
<th>icaD</th>
<th>clfA</th>
<th>clfB</th>
<th>fnbA</th>
<th>fnbB</th>
<th>cna</th>
<th>eno</th>
<th>fib</th>
<th>ebps</th>
<th>bbp</th>
</tr>
</thead>
<tbody>
<tr>
<td>MSSA</td>
<td>71.00</td>
<td>54.00</td>
<td>69.00</td>
<td>71.00</td>
<td>100.00</td>
<td>100.00</td>
<td>56.00</td>
<td>46.00</td>
<td>54.00</td>
<td>78.00</td>
<td>74.00</td>
<td>11.00</td>
<td>1.00</td>
</tr>
<tr>
<td>MRSA</td>
<td>76.00</td>
<td>64.00</td>
<td>69.00</td>
<td>74.00</td>
<td>97.40</td>
<td>97.40</td>
<td>64.00</td>
<td>51.00</td>
<td>56.00</td>
<td>79.00</td>
<td>76.00</td>
<td>12.00</td>
<td>0.00</td>
</tr>
<tr>
<td>P value</td>
<td>0.37</td>
<td>0.26</td>
<td>0.59</td>
<td>0.42</td>
<td>0.57</td>
<td>0.32</td>
<td>0.33</td>
<td>0.56</td>
<td>0.34</td>
<td>0.36</td>
<td>0.43</td>
<td>0.53</td>
<td></td>
</tr>
</tbody>
</table>

Fig. 5. The multiplex of eno, cna, ebps, and bbp genes. Lanes 1-6, and 8. eno and cna genes with 301 and 192 bp, respectively; Lane 7, cna gene (the ebps and bbp genes not shown). M: marker 100 bp (Fermentas, USA).

Table 4. The presence of biofilm-related genes in case of each agr specific group

<table>
<thead>
<tr>
<th>Group</th>
<th>icaA</th>
<th>icaB</th>
<th>icaC</th>
<th>icaD</th>
<th>clfA</th>
<th>clfB</th>
<th>fnbA</th>
<th>fnbB</th>
<th>cna</th>
<th>eno</th>
<th>fib</th>
<th>ebps</th>
<th>bbp</th>
</tr>
</thead>
<tbody>
<tr>
<td>agrI</td>
<td>59</td>
<td>45</td>
<td>61</td>
<td>61</td>
<td>98</td>
<td>98</td>
<td>65</td>
<td>45</td>
<td>57</td>
<td>90</td>
<td>70</td>
<td>12</td>
<td>0.02</td>
</tr>
<tr>
<td>agrII</td>
<td>54</td>
<td>49</td>
<td>56</td>
<td>54</td>
<td>100</td>
<td>100</td>
<td>74</td>
<td>44</td>
<td>62</td>
<td>92</td>
<td>69</td>
<td>13</td>
<td>0</td>
</tr>
<tr>
<td>agrIII</td>
<td>79</td>
<td>64</td>
<td>71</td>
<td>70</td>
<td>100</td>
<td>100</td>
<td>76</td>
<td>43</td>
<td>56</td>
<td>76</td>
<td>66</td>
<td>7</td>
<td>0</td>
</tr>
<tr>
<td>agrIV</td>
<td>67</td>
<td>60</td>
<td>67</td>
<td>69</td>
<td>100</td>
<td>100</td>
<td>67</td>
<td>33</td>
<td>67</td>
<td>80</td>
<td>70</td>
<td>7</td>
<td>0</td>
</tr>
</tbody>
</table>

In conclusion, this study reports that there is no relation between antibiotic resistance and biofilm formation in clinical isolates of S. aureus and also there is no correlation in the distribution of MSCRAMMs and biofilm genes with biofilm formation in vitro.
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CONFLICT OF INTEREST. None declared.

REFERENCES


